

Comprehensive Invited Review

Reactive Nitrogen Species: Molecular Mechanisms and Potential Significance in Health and Disease

M. Carmen Martínez and Ramaroson Andriantsitohaina

Reviewing Editors: Markus Bachschmid, Srinivas Bharath, Maria Ciriolo, Isabella Dalle-Dorne, Sergey Dikalov, Jeffrey Keller, Pamela Maher, and Hugo Monteiro

I. Introduction	670
II. RNS Formation and Mechanisms of Action	671
A. Posttranslational modifications induced by RNS	672
1. S-Nitrosylation	672
2. Glutathionylation	672
3. Tyrosine nitration	672
B. Relation between oxidative and nitrosative stresses	673
III. RNS Neutralization	673
IV. Molecular Targets of RNS: Cellular Effects of RNS	674
A. Lipids	674
B. DNA and RNA bases	675
C. Proteins	675
1. Ion channels	676
a. Ryanodine receptors	676
b. Inositol 1,4,5-triphosphate (InsP ₃) receptors	676
c. Sarco/endoplasmic Ca ²⁺ pumps (SERCA)	676
d. Store-operated Ca ²⁺ channels	677
e. Na ⁺ /Ca ²⁺ exchangers	677
f. Voltage-dependent Ca ²⁺ channels	677
g. K ⁺ channels	677
h. Voltage-gated Na ⁺ channels	678
i. NMDA	678
j. Other ion channels	678
2. Cytoskeletal and structural proteins	678
a. Actin	678
b. α -Tubulin	678
c. Calpains	679
d. N-Ethylmaleimide sensitive factor (NSF)	679
3. Matrix metalloproteinases (MMPs)	679
4. Proteins involved in cell cycle (proliferation)	679
5. Proteins involved in cell death	680
6. Proteins implicated in cell metabolism	680
a. Enzymes playing a key role in insulin-associated cascades	681
7. Nuclear factors	681
8. Proteins transporting oxygen	681

9. Albumin	682
10. Enzymes involved in prostaglandin generation	682
11. Mn superoxide dismutase (MnSOD)	682
V. RNS in Physiologic and Pathophysiologic Conditions	682
A. Physiology	682
B. Pathophysiology	683
1. Neurodegenerative diseases	684
a. Alzheimer's disease	684
b. Parkinson's disease	684
c. Amyotrophic lateral sclerosis (ALS)	685
d. Prion diseases	685
2. Cardiovascular diseases	685
a. Heart failure	685
b. Hypertension	685
c. Pulmonary arterial hypertension	685
d. Atherosclerosis	686
e. Preeclampsia	686
3. Inflammation	686
a. Airways diseases	686
b. Sepsis	686
c. Immune responses	687
4. Cancer	687
5. Metabolic diseases	688
VI. Future Directions	689

Abstract

Reactive nitrogen species (RNS) are various nitric oxide–derived compounds, including nitroxyl anion, nitrosonium cation, higher oxides of nitrogen, *S*-nitrosothiols, and dinitrosyl iron complexes. RNS have been recognized as playing a crucial role in the physiologic regulation of many, if not all, living cells, such as smooth muscle cells, cardiomyocytes, platelets, and nervous and juxtaglomerular cells. They possess pleiotropic properties on cellular targets after both posttranslational modifications and interactions with reactive oxygen species. Elevated levels of RNS have been implicated in cell injury and death by inducing nitrosative stress. The aim of this comprehensive review is to address the mechanisms of formation and removal of RNS, highlighting their potential cellular targets: lipids, DNA, and proteins. The specific importance of RNS and their paradoxical effects, depending on their local concentration under physiologic conditions, is underscored. An increasing number of compounds that modulate RNS processing or targets are being identified. Such compounds are now undergoing preclinical and clinical evaluations in the treatment of pathologies associated with RNS-induced cellular damage. Future research should help to elucidate the involvement of RNS in the therapeutic effect of drugs used to treat neurodegenerative, cardiovascular, metabolic, and inflammatory diseases and cancer. *Antioxid. Redox Signal.* 11, 669–702.

I. Introduction

NITRIC OXIDE (NO[•]) is a ubiquitous intracellular messenger able to regulate physiologic functions such as neural and cardiovascular activities. However, during pathologic conditions, NO[•] can become deleterious because of its high reactivity with other free radicals, such as the superoxide anion (O₂^{•−}). Although both NO[•] and O₂^{•−} at low concentrations are not toxic in a physiologic environment, an imbalance between the production of these two radicals can be partially responsible for alterations of the molecular mechanisms regulating cell life. This disequilibrium is mainly due to a diminished elimination or an increased production for O₂^{•−} or a reduction in the release or enhanced scavenging of NO[•]. NO[•] and O₂^{•−} react by an enzyme-independent mechanism to form peroxynitrite (ONOO[−]), a strong oxidant that reacts with most biologic molecules, causing cell damage. ONOO[−] is not the only reactive nitrogen species (RNS); RNS

refers to various nitrogenous products, such as NO[•], nitroxyl (HNO), nitrosonium cation (NO⁺), higher oxides of nitrogen, *S*-nitrosothiols (RSNOs), ONOO[−], and dinitrosyl iron complexes, excluding NO₃[−] (225) (Table 1). Each of these compounds has distinctive properties in terms of reactivity, half-life, lipid solubility, and biologic activity (225). Thus, RNS induce reactions including nitrosylation of sulfhydryls (*S*-nitrosylation) or metals and nitration of tyrosine residues. In addition, cross-talk between reactive oxygen species (ROS) and RNS, through chemical reaction or functional interaction or both, is possible, and it can exacerbate the deleterious effects. It has been suggested that when RNS production becomes excessive, they become deleterious to target cells, and this could play a role for a variety of the degenerative processes of some human diseases (37).

By analogy with ROS and oxidative stress, when the generation of RNS in a system exceeds its ability to neutralize

TABLE 1. MAIN REACTIVE NITROGEN SPECIES IN ALPHABETIC ORDER

Name	Formula	Formation
Dinitrogen trioxide	N_2O_3	From $NO\cdot$ and O_2
Nitric oxide or nitrogen monoxide	$NO\cdot$	From NOS
Nitrite	NO_2^-	From $NO\cdot$
Nitrogen dioxide	$NO_2\cdot$	From $ONOO^-$ decomposition
Nitronium cation	NO_2^+	From $ONOOCO_2^-$ decomposition
Nitrosonium cation	NO^+	From $NO\cdot$
Nitrosoperoxycarbonate anion	$ONOOCO_2^-$	$ONOO^-$ and CO_2
Nitroxyl	HNO	From one-electron reduction of $NO\cdot$
Nitryl chloride	$Cl-NO_2$	From NO_2^- and $HOCl$
Peroxynitrite	$ONOO^-$	From $NO\cdot$ and $O_2^{\cdot-}$
S-Nitrosothiols	$RSNOs$	From covalent addition of an $NO\cdot$ group to a cysteine thiol/sulphydryl

and eliminate them, nitrosative stress occurs (233). This review summarizes the main molecular mechanisms involved in the physiologic effects of RNS as well as the potential pathophysiologic consequences subsequent to the damage induced by RNS on various molecular targets. First, we outline the mechanisms of formation and removal of RNS, highlighting the potential cellular targets and the expected cell effects. Next, we give an overview of the physiologic role of RNS in the living cell, and finally, we look at the consequences of RNS in health, including diseases such as neurodegenerative and cardiovascular pathologies, metabolic and inflammatory diseases, and cancer.

II. RNS Formation and Mechanisms of Action

$NO\cdot$ plays a dual role in the nitrosative stress. On the one hand, $NO\cdot$ is the main RNS produced by cells, and on the other hand, it is the main source for the other RNS. $NO\cdot$ is produced by NO synthases (NOS) from L-arginine and oxygen. It promptly diffuses within cells and through cell membranes, constituting a second messenger leading, for example, to the relaxation of smooth muscle cells or synaptic transmission (see later). The three isoforms of NOS are NOS 1 or nNOS (for neuronal), NOS 2 or iNOS (for inducible), and NOS 3 or eNOS (for endothelial). Although NOS isoforms catalyze the same reactions, they differ in their expression, regulation, and physiologic/pathophysiologic roles (226). Both nNOS and eNOS are constitutively expressed; nNOS is expressed predominantly in neurons, and eNOS, in endothelial cells (183). Moreover, both isoforms are present in other cell types, such as smooth muscle cells (183). In addition, constitutive eNOS is able to produce low levels of $NO\cdot$, and it is accepted that their activation is "beneficial" for cells, as demonstrated by using eNOS knockout mice (215), whereas nNOS produce moderate levels of $NO\cdot$. By contrast, when iNOS expression is induced, predominantly in macrophages, under proinflammatory conditions, large quantities of $NO\cdot$ are produced (183). Regarding the regulation of the three isoforms of NOS, both nNOS and eNOS are regulated by intracellular Ca^{2+} /calmodulin, as well as by the interaction of other proteins such as hsp90 and caveolin. In contrast, iNOS is active even at low concentrations of Ca^{2+} , and it is regulated mainly at the level of transcription.

The direct action of $NO\cdot$ is limited by its biologic half-life *in vivo* (~ 1 sec) and by the relatively short distance that this molecule can traverse. Under physiological stress conditions, $NO\cdot$ binds to the heme iron of guanylyl cyclase (GC) and activates GC and cyclic guanosine 3',5'-monophosphate (cGMP)-dependent pathways. Indeed, $NO\cdot$ released from endothelial cells diffuses toward neighboring smooth muscle cells and activates GC. The cGMP produced interacts with protein kinase G (PKG), which phosphorylates contractile proteins, decreases cytosolic Ca^{2+} levels, evokes myosin light-chain dephosphorylation, and induces vasorelaxation (219).

Although NO_2^- has been considered the final metabolite together with NO_3^- in the decomposition of $NO\cdot$, recent data show that this RNS can be considered an intermediate in the complex pathway. NO_2^- is an oxidation product of NOS-derived $NO\cdot$ (212). $NO\cdot$ can be formed by xanthine oxidase-catalyzed reduction of NO_2^- under hypoxic conditions (206, 343) and participates in its protective effects. However, Patel's group (60, 100, 101) describes the cytoprotective effect of NO_2^- independent of the xanthine oxidase pathway. NO_2^- reacts with hemoglobin, which behaves as a nitrite reductase, and decreases the ability of hemoglobin to reduce $NO\cdot$ availability (100). $NO\cdot$ can react with the heme group of hemoglobin and therefore limits its $NO\cdot$ bioactivity (60). Thus, this mechanism may have a potential role in vasodilatation during hypoxic conditions (101).

More reactive than $NO\cdot$, $ONOO^-$ is a RNS resulting from the rapid reaction between $NO\cdot$ and $O_2^{\cdot-}$. $ONOO^-$ is a strong oxidant that mainly nitrates tyrosine residues in nonenzymatic reactions. Moreover, in a biologic system, $ONOO^-$ rapidly reacts with CO_2 (in equilibrium with physiologic levels of HCO_3^-), which leads to the formation of $CO_3^{\cdot-}$ and $NO_2^{\cdot-}$ radicals that, in turn, oxidate and nitrate proteins (312). $ONOO^-$ can react with CO_2 , resulting in the formation of an unstable nitrosoperoxycarbonate anion adduct ($ONOOCO_2^-$) that can serve as a source of several reactive intermediates (326). $ONOOCO_2^-$ reacts with guanine in DNA to form a variety of oxidation and nitration products, such as 8-nitro-2'-deoxyguanosine (77). $ONOOCO_2^-$ rapidly decomposes to yield NO_2^+ and CO_3^{2-} (66). Finally, $ONOO^-$ can decompose to NO_2^- and NO_3^- . In particular, the formation of NO_3^- from $ONOO^-$ depends mainly on the iso-

merization of ONOOH and the transient ONOOCO_2^- adduct (312).

A. Posttranslational modifications induced by RNS

When NO^\bullet was identified as the molecule accounting for the endothelium-derived relaxing factor, only the modifications on protein-bound transition metals (as described earlier for GC) induced by NO^\bullet were studied. However, experiments performed in recent years have shown that RNS can induce posttranslational modifications of a large number of proteins. Like phosphorylation, the most common posttranslational protein modification, the different modifications evoked by RNS on proteins alter protein structure and, consequently, protein function and interactions with downstream signaling targets. Three main posttranslational modifications can be induced by RNS: (a) S-nitrosylation, (b) glutathionylation, and (c) tyrosine nitration.

1. **S-Nitrosylation.** NO^\bullet can induce S-nitrosylation of regulatory protein thiol groups through its autooxidation with O_2 and the formation of dinitrogen trioxide (N_2O_3), a putative nitrosylating species. S-nitrosylation is the covalent addition of an NO^\bullet group to a crucial cysteine thiol/sulfhydryl to form an S-nitrosothiol derivative (RSNO) (Fig. 1). Such posttranslational and temporary protein modifications, which are independent of enzymatic activity, regulate the function of a large number of mammalian, plant, and microbial proteins with a susceptible thiol group. Overall, the susceptibility of protein cysteine sulfhydryl to S-nitrosylation is increased by a consensus motif of amino acids comprising nucleophilic residues surrounding "the critical cysteine" (119). This mechanism of S-nitrosylation has protective effects in different systems, preventing some critical protein thiols from further irreversible oxidative modifications by ROS. Thus, Stamler's group (112) has suggested that ROS can alter the balance between phosphorylation and S-nitrosylation of key signaling proteins. Among these proteins, protein kinases and phosphatases, metabolic enzymes, membrane receptors and channels, cytoskeletal elements, and transcription factors are potential candidates to be S-nitrosylated (for a review, see ref. 119). In consequence, alterations in the regulation of protein S-nitrosylation or in the levels of S-nitrosothiols are often linked to pathophysiologic states including human diabetes (207), cystic fibrosis (105), or mouse sepsis (184).

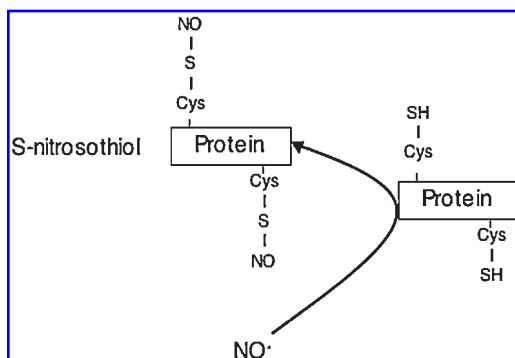


FIG. 1. S-Nitrosylation of proteins. Coupling of an NO^\bullet moiety to a reactive thiol group of cysteine residues of peptides and proteins allows the formation of S-nitrosothiols.

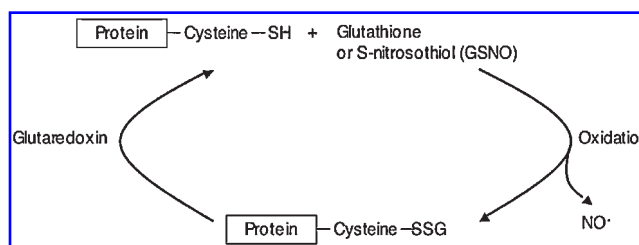


FIG. 2. Mechanism(s) of glutathionylation. Reactions between cysteine residues and glutathione or S-nitrosothiols such as GSNO are able to release NO^\bullet .

Because of the lability of the S-NO^\bullet complex, detection of S-nitrosylated proteins becomes difficult, and, in general, indirect methods of detection are used. New techniques to facilitate quantification of S-nitrosylation of proteins should allow a better understanding of the consequence of this posttranslational modification in cell function.

2. **Glutathionylation.** Glutathionylation, also referred to as glutathiolation, is the addition of glutathione (GSH), or other low-molecular-weight thiols, to the cysteine sulfhydryl residues of proteins and represents the most important thiolation. This reaction is reversible and independent of enzyme activity. After S-nitrosylation of GSH and generation of S-nitrosoglutathione (GSNO), the formed GSNO molecule can mediate posttranslational modifications (*i.e.*, modification of protein thiols *via* S-nitrosylation and glutathionylation) (32, 112) (Fig. 2). Thus, S-nitrosylation can be considered an intermediate signaling event. Then GSNO may account for remote or long-lasting effects of NO^\bullet , serving to store, transport, and deliver NO^\bullet *in vivo* to its neighboring proteins (217). Thus, GSNO has been suggested as the main physiologic mediator of NO^\bullet effects. GSNO decomposes slowly to generate NO^\bullet (349), and denitrosylation reactions can be accomplished by GSNO reductase (182). GSNO reductase, also called alcohol dehydrogenase of class III, is the major enzyme that catalyzes GSNO metabolism. GSNO reductase generates glutathione disulfide (GSSG) and ammonia, and allows the regulation of the intracellular levels of S-nitrosothiols (115, 182). By using GSNO reductase-deficient mice, Liu *et al.* (184) demonstrated that this enzyme is indispensable for S-nitrosothiol metabolism, regulation of blood pressure, and protection from nitrosative stress in response to endotoxin and bacteria. Thus, protein S-glutathionylation is considered an adaptive cellular response protecting essential proteins involved in regulatory pathways from permanent loss of function as a consequence of nitrosative stress (152). However, further studies are needed the better to understand the biologic consequences of this posttranslational modification on the regulation of the function of target proteins.

3. **Tyrosine nitration.** Tyrosine nitration is a two-step process in which the first reaction is the generation of tyrosyl radical by oxidation of tyrosine by reactive species generated from ONOO^- . In the second step, tyrosyl radical reacts with NO_2 to form 3- NO_2 -Tyr (3-nitrotyrosine) (260) (Fig. 3). Nitration of tyrosine residues is widely used as a biomarker of ONOO^- generated from the reaction between NO^\bullet and $\text{O}_2^{\bullet -}$. ONOO^- is typically more reactive and toxic than NO^\bullet *per se*.

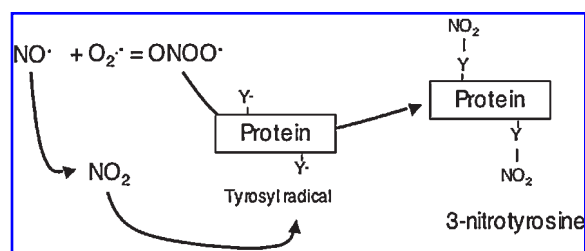


FIG 3. Formation of ONOO^- from the $\text{NO}\cdot$ and $\text{O}_2^{\cdot-}$ as reaction and the subsequent nitration of tyrosyl radical to form nitrotyrosine. High concentrations of ONOO^- lead to nitration of proteins on tyrosine residues. Tyrosine nitration is a two-step process: first, the generation of tyrosyl radical by oxidation of tyrosine by reactive species generated from ONOO^- , and second, the reaction of tyrosyl radical with NO_2 to form 3- NO_2 -Tyr(3-nitrotyrosine).

Originally, ONOO^- has been suggested to be a strong biologic oxidant that can react directly with sulfhydryls, iron-sulfur, and zinc-thiolate groups (for review, see ref. 238). In addition to tyrosine nitration, ONOO^- participates in hydroxylation and oxidation reactions (245). In particular, tyrosine nitration may alter several protein activities, and increased circulating levels of 3-nitrotyrosine are correlated with cardiovascular diseases such as coronary artery disease (294). Furthermore, it has been suggested that low amounts of ONOO^- (under physiological stress conditions) can activate pathways leading to the activation of Akt, a kinase that, in human skin primary fibroblasts, is at least in part responsible for antiapoptotic and survival signals (154). Moreover, low physiological levels of ONOO^- can interact with the tripeptide GSH, resulting in reversible S-glutathionylation of proteins (3). Another RNS, NO_2 , has been described as a nitrating molecule.

Several authors consider that tyrosine nitration is related to nitrative stress rather than to nitrosative stress. In addition, another concept, nitrooxidative stress, may replace that of nitrative stress. However, taking into consideration the number of original articles in which RNS-induced nitration is referred to as nitrosative stress, we use this term in this review.

Although data obtained in *in vitro* studies clearly show that nitration of proteins could alter protein structure and function, a large number of questions remain to be answered to explain completely the biologic significance of protein nitration *in vivo*. Comparison of proteomic posttranslational modification analysis of tissues obtained from patients *versus* "healthy" tissues could contribute to the discovery of new elements for a better understanding of disease etiology, and then, to the development of new tools for research against diseases.

Very recently, a novel mechanism was described. Sawa *et al.* (282) showed that the nitrate derivative of cGMP, 8-nitroguanosine 3',5'-cyclic monophosphate (8-nitro-cGMP), generated through iNOS activation, is able to form protein Cys-cGMP adducts by S-guanylation. In addition, 8-nitro-cGMP can regulate the redox-sensor signaling of protein Keap1, suggesting that 8-nitro-cGMP can be considered a second messenger of $\text{NO}\cdot$.

B. Relation between oxidative and nitrosative stresses

Oxidative stress is closely related to nitrosative stress. In general, the excessive production of ROS or the failure of cellular removal of these species or both are responsible for increased oxidative stress. Once ROS levels are enhanced, reactions between ROS and RNS take place and lead to nitrosative stress. Although oxidative stress is not the focus of this review, we consider it necessary to comment briefly on the main ROS implicated in the generation of nitrosative stress. Elevated levels of $\text{O}_2^{\cdot-}$ and hydrogen peroxide chiefly contribute to the development of oxidative stress, and their subsequent reactions with RNS result in nitrosative stress. The half-lives of ROS vary considerably from a few nanoseconds for the most-reactive compounds, such as $\cdot\text{OH}$, to seconds and hours for more-stable ROS, such as hypochlorous acid (311). ROS are generated by a wide array of enzymes that include NADPH oxidases, xanthine oxidase, peroxidases, lipoxygenases, and cyclooxygenases, and complex I and III of the respiratory chain in the mitochondrion. NADPH oxidases are enzymes with catalytic and regulatory subunits that catalyze single-electron reductions of molecular oxygen by using NAD(P)H as electron sources (106, 165). Conversely, xanthine oxidase produces $\text{O}_2^{\cdot-}$ or hydrogen peroxide as products of the terminal steps of purine metabolism, metabolizing hypoxanthine and xanthine to uric acid with concomitant generation of $\text{O}_2^{\cdot-}$ (25). In addition, xanthine oxidase can generate $\text{NO}\cdot$ under hypoxic situations, either from organic nitrites in the presence of xanthine as a substrate (76) or when NOS cannot produce $\text{NO}\cdot$ (175). However, the physiologic relevance of this pathway *in vivo* remains to be elucidated.

Under certain conditions, NOS themselves can generate $\text{O}_2^{\cdot-}$ as a consequence of the electron transfer within the active site becoming "uncoupled" from L-arginine oxidation; instead, molecular oxygen is reduced to form $\text{O}_2^{\cdot-}$ (331, 332). Various molecular mechanisms could contribute to NOS uncoupling. Limited availability of the substrate L-arginine may reduce $\text{NO}\cdot$ synthesis; under these conditions, the Ca^{2+} /calmodulin-activated NOS resembles the plasma membrane-associated NADPH-dependent oxidase of neutrophils (255). Also, the interaction between heat-shock protein 90 and NOS seems to regulate the balance between $\text{NO}\cdot$ and $\text{O}_2^{\cdot-}$ productions (257). Tetrahydrobiopterin (BH_4), which acts as an essential cofactor for all three NOS isoforms (6), appears to have the main role in the regulation of this equilibrium. Recent evidence suggests that BH_4 depletion is responsible for NOS uncoupling, because of its oxidation and/or reduced synthesis as described in mice with hypertension (168).

III. RNS Neutralization

The mechanisms that protect mammalian cells from the deleterious effects of nitrosative stress remain to be clarified. Nonenzymatic protection against RNS may be provided by cellular antioxidants, such as GSH, metalloporphyrins, selenium compounds, uric acid, β -carotene, and vitamins E and C, as well as by the elimination of $\text{NO}\cdot$ by its rapid reaction with hemoglobin. Proteins such as thioredoxin (243), GSH peroxidases (31), superoxide dismutases (SOD) (143), or γ -glutamyl transpeptidase (131) have been recognized as potential systems able to remove RNS. Thioredoxin proteins

are classically defined by their ability to reduce disulfides to dithiols. Besides GSH, thioredoxin constitutes a major protective mechanism. Thus, both thioredoxin and GSH reduce protein thiols and various ROS, such as $O_2^{\cdot-}$ and hydrogen peroxide (243). Both systems operate independently and, moreover, GSH levels are higher than those of thioredoxin (73, 342). GSH peroxidases are selenoproteins able to reduce hydroperoxides to the corresponding alcohols by means of GSH. In addition to their antioxidant activities (GSH peroxidases are the main intracellular antioxidant), they interact with redox-regulated transcription factors, such as the NF- κ B and Nrf2/Keap1 systems (31), suggesting that GSH peroxidases can be involved in the regulation of the inflammatory process. SODs are metalloenzymes that catalyze the conversion of $O_2^{\cdot-}$ to hydrogen peroxide, which immediately becomes a substrate for the enzyme catalase and other hydrogen peroxide-catalyzing enzymes such as GSH peroxidases and peroxiredoxins. Up to now, three different SOD isoforms have been described, and they ensure the major scavenging systems in the cytoplasm and nucleus (SOD1 or CuZn-SOD), mitochondrion (SOD2 or Mn-SOD), and extracellular space (SOD3 or EC-SOD) (143). Transcription of the *sod* genes is increased in response to cell stress, leading to increased activity to protect cells from damage.

All of these elements underscore the variety of RNS and the subsequent reactions that can be observed, and suggest that the reactions associated with RNS are, in general, more complex than those of ROS.

IV. Molecular Targets of RNS: Cellular Effects of RNS

RNS can react with lipids, DNA and RNA bases, metal cofactors, and proteins (Fig. 4).

A. Lipids

Regarding lipids, RNS can react with cellular lipids directly or indirectly, generating a spectrum of products, many of which contain functional groups capable of modifying proteins (361). The reactive lipid products can be divided into three major classes: first, lipid peroxides are products derived from the enzymatic action of 15-lipoxygenase (8);

second, reactive lipids with electrophilic properties, such as 15-deoxy- $\Delta^{12,14}$ -prostaglandin J₂, and 4-hydroxynonenal (50, 166); finally, receptor agonists including nitrolinoleic acid (LNO₂) and lysophosphatidylcholine (50). It is interesting to note that, under certain conditions, lipids reacting with RNS may become donors of NO \cdot , and, consequently, no deleterious effects are described. For example, LNO₂, a product of NO \cdot -dependent linoleic acid nitration, is the most abundant, biologically active oxide of nitrogen in the human vascular compartment (15). LNO₂ acts as a signaling mediator *via* both receptor-dependent and receptor-independent pathways. Thus, LNO₂ is a specific and a high-affinity endogenous ligand for peroxisome proliferators-activated receptor- γ (PPAR γ) (286) (Fig. 5). Besides, LNO₂ activates cAMP-dependent protein kinase pathways in cells involved in inflammation. This reaction confers cell protection against the inflammatory process. In the vascular compartment, LNO₂ elicits endothelium-independent relaxation by stimulating GC and generation of cGMP. Recently, Schopfer and colleagues (287) identified the intermediate product between LNO₂ and GC as NO \cdot and propose LNO₂ as a hydrophobically stabilized NO \cdot donor (Fig. 5). The nitro derivative of oleic acid (OA-NO₂), as well as LNO₂, is able to react with GSH and to form GS-OA-NO₂ and GS-LNO₂ (16) (Fig. 5). Baker and colleagues (16) report that the reversible adduction of thiols by nitro-fatty acids is a mechanism for reversible posttranslational regulation of protein function by nitro-fatty acids. Both OA-NO₂ and LNO₂ have been described recently as endogenous antiinflammatory mediators able to repress NF- κ B-dependent gene expression and subsequently to reduce inflammatory response in vascular cells (62).

Another aspect is the ability of nitrated products of unsaturated fatty acids, such as OA-NO₂ and LNO₂, to inhibit vascular smooth muscle cell proliferation by inducing growth arrest of these cells in the phase G₁/S of the cell cycle. This effect of the nitroalkenes (the nitration products of unsaturated fatty acids formed *via* NO \cdot -dependent oxidative reactions) is associated with an upregulation of the cyclin-dependent kinase inhibitor p27^{kip1}. In addition, modifications at the level of the translocation of the nuclear factor

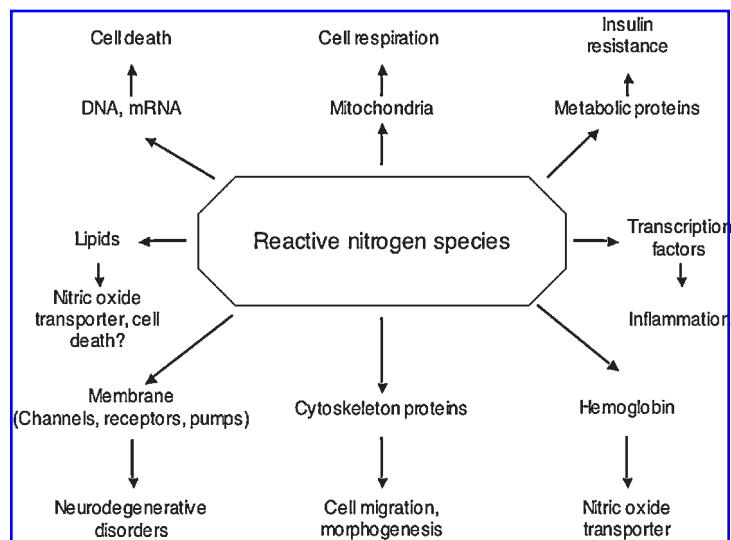


FIG. 4. Molecular targets of reactive nitrogen species (RNS). RNS are able to act on a variety of targets responsible for their pleiotropic effects.

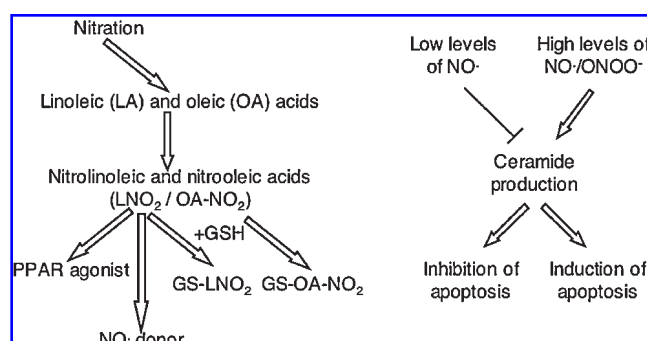


FIG. 5. Interaction of RNS with lipids. Through the nitration of linoleic (LA) and oleic (OA) acids, lipids can become NO[•] donors. Nitrolinoleic (LNO₂) and nitrooleic (OA-NO₂) acids protect cells from inflammation, through the activation of peroxisome proliferators-activated receptor-γ (PPARγ). Moreover, LNO₂ and OA-NO₂ can react with glutathione and form GS-OA-NO₂ and GS-LNO₂. The effects of RNS on ceramide metabolism can modulate cellular apoptosis.

Nrf2 have been reported (334). These findings indicate that LNO₂ could have an important role in the growth of vascular smooth muscle cells and participate in the development of novel therapeutic strategies for cardiovascular diseases.

Conversely, several lines of evidence suggest that RNS are tightly linked to the regulation of the sphingolipid metabolism through regulating the enzymes responsible for their metabolism, including sphingomyelinase (SMase) and ceramidase (346). Sphingolipids are ubiquitous constituents of membrane lipids in mammalian cells. They are involved in proliferation, differentiation, apoptosis, and inflammation (346). Sphingolipid formation of ceramide and its derivatives such as sphingosine is regulated by the action of SMase. Paradoxical effects have been observed between NO[•] and SMase activity (Fig. 5). Exogenous or endogenous NO[•] is described as an inhibitor of apoptosis through the inhibition of ceramide generation (22). However, excessive amounts of NO[•] activate apoptosis *via* the increase of ceramide production (251). Among RNS, ONOO[•] induces apoptosis through the modulation of ceramide levels in human airway epithelial cells by activating an acidic SMase (46), whereas NO[•] donors increase ceramide levels without triggering apoptosis (47). The latter mechanism implicates the increase in the protein-protein interaction between SMase and caspase-3 that prevents its cleavage. Besides, certain sphingolipids, such as ceramide, are able to induce cellular oxidative and nitrosative stresses through the activation of NADPH oxidase, NOS, and mitochondrial dysfunction (for a review, see ref. 346). These data suggest a bidirectional regulation of sphingolipid metabolism and oxidative and nitrosative stresses.

B. DNA and RNA bases

DNA damage induced by RNS may contribute to increased mutation rates, genome instability, apoptosis, and associated tissue regeneration and cell proliferation, all of which can lead to carcinogenesis (281, 313). ONOO[•] and nitrogen oxides are the two RNS mainly implicated in DNA

and RNA damage by inducing base modifications and strand breaks (281). In particular, on reaction with guanine, ONOO[•] is able to form 8-nitroguanine (8-NO₂-G), which induces G:C to T:A transversions in DNA *in vitro* (356) (Fig. 6). 8-Nitroguanosine (8-NO₂-Guo) is formed by the reaction of ONOO[•] with guanosine (355) (Fig. 6). The formation of 8-NO₂-Guo from guanosine in RNA from *in vivo* virus-infected lungs from mice suggests that 8-NO₂-Guo could be involved in the toxicity of NO[•] during infection and inflammation, at least in murine models (5). The biologic significance of 8-NO₂-G and 8-NO₂-Guo is difficult to determine because of their relative instability and the difficulty of quantitative estimation (233). Thus, both 8-nitropurines can be easily oxidized *in vitro* with ONOO[•] (171). However, several studies have shown that 8-NO₂-Guo is a highly redox-active nucleic acid derivative that strongly stimulates O₂^{•-} generation in the presence of cytochrome P450 reductase and all isoforms of NOS (5, 280). In addition, nitrated nucleic acids may be formed in the cytoplasmic nucleotide compartment or in RNA by RNS generated in cell compartments other than the nucleus. Under these conditions, 8-NO₂-G and related nucleosides and nucleotides may be incorporated into RNA to interfere with RNA function and metabolism (234), and thus, interfere with enzymes that use guanine nucleotides as substrates.

At the end of the cascade mediated by ONOO[•], the increase in DNA strand breakage triggers the activation of poly-ADP-ribose polymerase (PARP), a DNA-repair enzyme. However, this mechanism, taking place to remove the lesions induced by ONOO[•], becomes deleterious (162) and results in energy depletion (NAD⁺ and ATP depletions) and consequently in necrotic cell death (337) (Fig. 6).

C. Proteins

A large number of proteins are cellular targets of RNS. Consequent to RNS exposure, proteins are modified, lead-

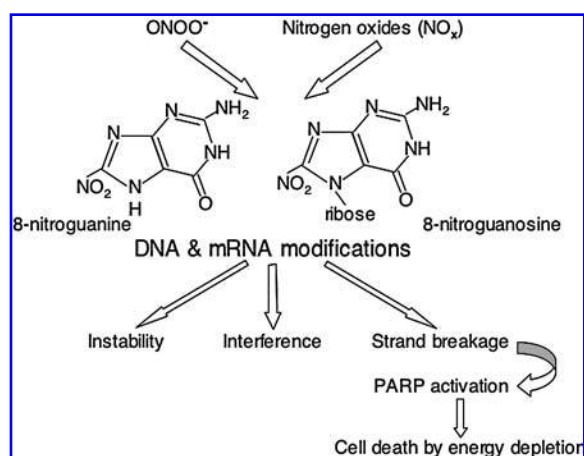


FIG. 6. Damaging effect of RNS on DNA and mRNA. Direct action of RNS [mainly ONOO[•] and nitrogen oxides (NO_x)] on DNA and mRNA, *via* the formation of 8-nitroguanine and 8-nitroguanosine, induces changes in cellular behavior that can lead to carcinogenesis, and poly-ADP-ribose polymerase (PARP) activation, inducing cell death by energy depletion.

ing to an activation or inhibition of their activities, as well as altering the protein–protein interactions. In this current review, we concentrate mainly on the molecular targets of RNS, depending on their function.

1. Ion channels. Ion channels are transmembrane proteins that are implicated in a wide variety of cellular functions by selectively controlling the passage of ions across the plasma membrane, as well as the mitochondrial outer and inner membranes, endoplasmic reticulum, and other organelles. They mainly control activity of excitable cells (neurons and muscle cells). However, ion channels may also be expressed in the so-called “nonexcitable” cell types, including lymphocytes, glia, fibroblasts, and endothelial cells (74). Depending on the structure of ion channels, they appear as susceptible targets of RNS, and the subsequent aberrant ion fluxes have been associated with cell death. We focus this part of the review on Ca^{2+} channels mainly expressed in neurons but also in other excitable and nonexcitable cells, and Na^+ and K^+ channels.

a. Ryanodine receptors. The ryanodine receptor (RyR) functions as the principal ion channel that controls the release of Ca^{2+} from the endoplasmic and sarcoplasmic reticulum and, thereby, muscle contractions. RyR contains cysteine residues, which are susceptible to modification by oxidation, S-nitrosylation, S-glutathionylation, and alkylation. Modifications induced by GSNO or NO^{\cdot} donors significantly enhance RyR activity in the skeletal muscle (309). Eu and co-workers (82) showed that Ca^{2+} flux through RyR of skeletal muscle is activated by S-nitrosylation. Moreover, the canine cardiac form of RyR can be S-nitrosylated under basal conditions, resulting in an activation of RyR, which could be associated, in isolated rabbit cardiac sarcoplasmic reticulum vesicles, with the inhibition of the Ca^{2+} -ATPase-mediated uptake of Ca^{2+} into the sarcoplasmic reticulum (347, 348). Taken together, RNS through the activation of RyR are potential inotropic agents to increase Ca^{2+} transient and therefore the strength of the contraction (Fig. 7). By contrast, other authors have described the inhibition of both cardiac and skeletal RyRs by RNS (202), depending on their concentrations (114), suggesting that, *in vivo*, initial release of NO^{\cdot} activates RyRs, but greater release inhibits RyR activity and contraction.

b. Inositol 1,4,5-triphosphate (InsP3) receptors. These receptors are a family of Ca^{2+} -release channels localized pre-

dominantly in the endoplasmic reticulum and in the nuclear envelope of all cell types. They are able to release Ca^{2+} into the cytoplasm in response to InsP3 produced from the plasma membrane after engagement of extracellular ligands. The InsP3-receptor activation generates complex local and global Ca^{2+} signals that regulate numerous cell physiologic processes (for a review, see ref. 91). To fulfill this function, the InsP3 receptors depend on interaction with accessory subunits and regulatory proteins. Little is known about the interactions of InsP3 receptors and RNS. Chung *et al.* (54) reported that NO^{\cdot} donors inhibit agonist-evoked inward cation current oscillations (see later), and these effects are mediated, in part, by functional modulation of the InsP3 receptors. Other authors demonstrated that NO^{\cdot} donors and ONOO^- induce Ca^{2+} -mobilization from InsP3-dependent internal stores through the activation of cGMP pathways in bovine chromaffin cells (333) (Fig. 7). Under hypoxic conditions, the InsP3-receptor characteristics [number of receptor sites (B_{max}) and dissociation constant (K_d)] were modified by NO^{\cdot} in neuronal nuclei of piglets (209). These NO^{\cdot} -mediated modifications of the InsP3 receptors may lead to the increase of intranuclear Ca^{2+} , resulting in altered transcription of apoptotic genes and activation of cascades of hypoxia-induced programmed neuronal death. However, further studies are needed to understand the effects of RNS on InsP3 receptors.

c. Sarco/endoplasmic Ca^{2+} pumps (SERCA). SERCAs function to maintain the pools of Ca^{2+} in either the sarcoplasmic or endoplasmic reticulum by using the energy of hydrolysis of ATP as opposed to the nonspecific leakage of Ca^{2+} from a high concentration in these organelles (277). The SERCA family includes the products of three genes, called *SERCA1*, *SERCA2*, and *SERCA3*. They are especially expressed in skeletal, cardiac and smooth muscles (mainly *SERCA1* and *SERCA2*), regulating the contraction/relaxation coupling, but also in nonmuscle tissues (*SERCA3*). These proteins present several tyrosine residues susceptible to nitration and also, cysteines able to be S-glutathionylated. Indeed, dual effects of ONOO^- on SERCA have been described, depending on the residue target. ONOO^- treatment of pig smooth muscle or endothelial cells inhibited the SERCA pump more severely in the former (281). This effect may be mediated by nitration of SERCA, as described in other models associated with different pathologies (2, 315, 350) (Fig. 7). Whereas the effects of SERCA nitration may have deleterious consequences, S-glutathionylation may be beneficial. It has been demonstrated that, under physiologic

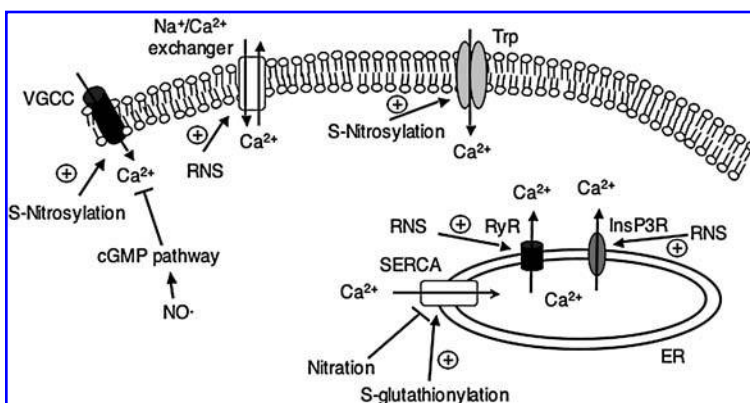


FIG. 7. Selected effects of RNS on calcium channels and pumps regulating calcium currents in living cells.

conditions, NO[•] forms S-glutathione adducts with SERCA, increasing its activity, accelerating Ca²⁺ uptake by the endoplasmic reticulum, and decreasing intracellular Ca²⁺ (3). In more detail, physiologic levels of NO[•] react with O₂^{•-} to form ONOO⁻, which, together with GSH, reacts with cysteine residues and increases SERCA activity (Fig. 7). S-Glutathionylation may be implicated in the arterial relaxation independent of cGMP, and its alteration could be related to failure processes. In pathologic states, such as atherosclerosis, an increase in ROS/RNS irreversibly oxidizes thiols of SERCA and decreases their activity (3).

Recently, it was been suggested that capsaicin-sensitive sensory neurons in isolated rat hearts could regulate myocardial function through the maintaining of S-nitrosylation of SERCA induced by ONOO⁻ (24).

d. Store-operated Ca²⁺ channels. Once InsP3 receptors are activated and Ca²⁺ is released from the endoplasmic reticulum, the resulting depletion of Ca²⁺ within this organelle serves as a message, which is returned to the plasma membrane, resulting in the slow activation of "store-operated" Ca²⁺ channels. This Ca²⁺ entry phase sustains cytosolic Ca²⁺ increase and, at the same time, replenishes intracellular stores (173).

It has been reported that Ca²⁺ entry can be directly activated by S-nitrosylation (86), but the relation between this process and the "store-operated" Ca²⁺ entry is complex. Whereas activation of "store-operated" Ca²⁺ channels appears to require InsP3 receptor as a coupling intermediary, NO[•] donors act directly on the channel (329) (Fig. 7). The nature of these channels remains to be elucidated, although a transient receptor potential channel (TRP) has been proposed. Very recently, evidence that the S-nitrosylation of TRPC5 elicits Ca²⁺ entry into endothelial cells has been proposed, suggesting that members of the TRP family can be considered NO[•] sensors (357).

e. Na⁺/Ca²⁺ exchangers. These proteins are members of a transporter family whose main role is to provide control of Ca²⁺ flux across the plasma membranes or intracellular compartments by extruding Ca²⁺ from the cytoplasm. Curiously, Na⁺/Ca²⁺ exchanger is reversible and, under certain conditions, may allow Ca²⁺ to enter the cytoplasm (for a review, see ref. 190). It was recently reported that NO[•] activates Na⁺/Ca²⁺ exchanger activity and that Na⁺/Ca²⁺ exchanger is involved in NO[•]-induced depletion of Ca²⁺ in the endoplasmic reticulum from cultured microglia, leading to endoplasmic reticulum stress (198) (Fig. 7). Maczewski and Beresewicz (192) also showed that the inhibition of Na⁺/Ca²⁺ exchanger attenuates postischemic myocardial formation of NO[•] and ROS in rat heart, suggesting that prevention of calcium overload is cardioprotective *via* the NO[•] pathway.

f. Voltage-dependent Ca²⁺ channels. Voltage-dependent Ca²⁺ channels (VGCCs) mediate Ca²⁺ entry into excitable cells in response to membrane depolarization. They are heterodimeric multiprotein complexes consisting of a pore-forming α 1 subunit and other modulatory subunits (β , γ , and α 2 δ) (352).

Although RNS exert pronounced effects on VGCC activity, no clear conclusion can be reached, inasmuch as contra-

dictory data have been reported. Indeed, depending on the subunit of VGCC affected by RNS, the observed effects can be diverse (Fig. 7). Regarding the stimulation of Ca²⁺ currents through these channels, Wang *et al.* (340) showed that NO[•] released by agonists stimulates L-type Ca²⁺ current in cat atrial myocytes. The same group showed that NO[•] can act *via* an S-nitrosylation mechanism to prevent the β 2-adrenoceptor agonist-induced inhibition of L-type Ca²⁺ current in the same model (65). The latter results indicate that NO[•] may increase the L-type Ca²⁺ current in this system and exert cardioprotective effects. In neurons, NO[•] donors enhance Ca²⁺ current amplitude and block adrenoceptor-induced Ca²⁺ current inhibition *via* stimulation of cGMP formation, suggesting that NO[•] donors can interfere with neuromodulation of Ca²⁺ channels (48). In smooth muscle cells, NO donors inhibit L-type Ca²⁺ channels through a cGMP signaling pathway, thereby suppressing the sustained increase in intracellular Ca²⁺ concentration, suggesting that these channels are important targets for nitrovasodilators (208).

S-Nitrosothiols, in particular GSNO, exert a potent inhibitory effect on L-type Ca²⁺ channels by affecting central functions (gating and ion permeation) of the pore-forming α subunit of these channels (253) by a mechanism independent of cGMP (128). NO[•] donors also inhibit L-type Ca²⁺ in rabbit carotid body sensory activity, through a direct action on the channel or an associated channel protein or both, rather than through a cGMP-dependent mechanism (307). This may explain the fact that NO[•] can modulate neurotransmitter release in these cells to maintain homeostasis during hypoxemia. Abi-Gerges and co-workers (1) proposed the hypothesis that the inhibitory effects of NO[•] donors on L-type Ca²⁺ channels in rat cardiomyocytes is dependent on the redox complex environment in cells, taking place directly on the channel itself or on other auxiliary proteins or lipids, or both.

g. K⁺ channels. K⁺ channels are a diverse and ubiquitous family of membrane proteins present in both excitable and nonexcitable cells, playing key roles in cellular signaling processes such as neurotransmitter release, heart rate, insulin secretion, neuronal excitability, epithelial electrolyte transport, smooth muscle contraction, and volume regulation (for review, see ref. 293). These channels selectively conduct K⁺ ions across the cell membrane along its electrochemical gradient. A very large number of K⁺-channel genes have already been identified and, consequently, deduced amino acid sequences and secondary structure information can be used to generate structural classification. Three groups of K⁺ channels have been characterized: (a) K⁺ channels with six or seven transmembrane segments and one pore region (voltage-activated and Ca²⁺-activated K⁺ channels); (b) large-conductance voltage-gated K⁺ channels with seven transmembrane segments; and (c) finally, the inward rectifiers K⁺ channels with two transmembrane segments and one pore, including ATP-sensitive and ATP-regulated K⁺ channels. It is obvious that many different K⁺ channels with diverse kinetics and functions exist, increasing the possibility of interaction with RNS. Although the most important posttranslational modifications of K⁺ channels are palmitoylation, glycosylation, and phosphorylation (27), RNS can affect these channels by activating or inhibiting their activities.

During the 1990s, several studies performed in vascular smooth muscle cells showed that NO[•] induces nitrosylation of sulfhydryl groups of large conductance Ca²⁺-activated K⁺ channels (4, 28). With inside-out patch recording, Lu and co-workers (189) showed that NO[•] increases whole colonic smooth muscle cell outward K⁺ current by activating Ca²⁺-activated K⁺ channels through a cGMP pathway (210); this phenomenon could play an important functional role in the control of the vascular peripheral resistance induced by NO[•]. Recently, a similar effect was described in human dermal fibroblasts (179). NO[•] may interact with the α -subunit of the channels and activate them directly (140) without any involvement of covalent modification, such as S-nitrosylation proposed by other authors.

Regarding the effects of RNS on ATP-dependent K⁺ channels in pancreatic β cells, NO[•] donors inhibit glucose-induced insulin secretion by acting directly on the activity of these channels. These effects result in the opening of the channels (321). NO[•] itself and NO[•] donors affect the activity of ATP-dependent K⁺ channels by different mechanisms (78). On the one hand, ATP-dependent K⁺ current is stimulated *via* a reduction in the ATP/ADP ratio, and on the other hand, it is inhibited by direct interference with the channel or a regulatory protein in its close vicinity (78).

Finally, it has been suggested that NO[•] donors may exert an inhibitory effect on outward K⁺ currents *via* an interaction with ROS in a cGMP-independent way (314). These results may have considerable relevance for the understanding of the early events leading to ischemia-related toxicity in excitable tissues associated with alterations in the balance between ROS and RNS.

h. Voltage-gated Na⁺ channels. Voltage-dependent Na⁺ currents are responsible for the rapid upstroke of the action potential in neurons and striated muscle and determine the membrane excitability. Taking in consideration their importance in action-potential generation, modulation of Na⁺ currents causes significant changes in membrane excitability (26). Acting on these channels by S-nitrosylation, NO[•] donors decrease Na⁺ currents in baroreceptor neurons (174), which play a crucial role in acute regulation of the arterial pressure. NO[•] donors also reduced the three types of Na⁺ currents described in C-type dorsal root ganglion neurons (268). In contrast, other authors showed that NO[•] donors increase the amplitude of the Na⁺ currents in rat hippocampal neurons involving S-nitrosylation (111). Altogether, these data suggest that NO[•] donors may modulate Na⁺ currents differently, depending on cell type.

i. NMDA. N-methyl-D-aspartate receptors (NMDA) are glutamate-gated ion channels that are essential for the regulation of synaptic function in the central nervous system. They are highly permeable to Ca²⁺, and this Ca²⁺ influx is necessary for synaptogenesis, experience-dependent synaptic remodeling, and long-lasting changes in synaptic efficacy (for a review, see ref. 170). An excessive activation of NMDA receptors is implicated in neuronal damage, resulting in exaggerated Ca²⁺ influx through the channel; this process has been called "excitotoxic activation." Lipton's group (180) was the first to show that S-nitrosylation of NMDA receptors may play a neuroprotective role. Among the five cysteine residues of the NMDA receptors, Cys399 of the NR2A subunit medi-

ates the predominant inhibitory effect of S-nitrosylation, decreasing NMDA-receptor function (52). Recent data indicate that S-nitrosylation of other proteins involved in cell death (see later) can act in synergy with NMDA S-nitrosylation and promote neuronal survival (222).

j. Other ion channels. Cystic fibrosis transmembrane conductance regulator (CFTR) is a cAMP-regulated chloride channel present in the apical membrane of epithelia in many organs. The maturation defect of CFTR presenting the δ F508 mutation is responsible for cystic fibrosis disease. Recent data open promising new therapies, because GSNO can normalize expression, maturation, and activity of CFTR (49, 291). In contrast, other studies showed that, although GSNO effectively affects CFTR activity, this effect is not mediated by S-nitrosylation but rather by induction of the expression of cysteine string proteins (358). Moreover, higher doses of NO[•] can augment CFTR breakdown by causing tyrosine nitration (23, 142) or alteration of CFTR trafficking (299).

2. Cytoskeletal and structural proteins. One of the major structural components of the cells is the cytoskeleton, a filamentous network of F-actin, microtubules, and intermediate filaments. In addition, the filamentous cytoskeletal network provides a scaffolding on which other proteins such as kinesin, dynein, and myosin can translocate to move organelles or generate internal stress (138). It is formed by a large number of proteins that play a crucial role in dynamic cellular processes, including the maintenance of the structural and mechanical integrity of cells and tissues and, more specifically, cell migration and adhesion, cytokinesis, membrane trafficking, and morphogenesis. Because a large number of cytoskeletal proteins are affected by RNS, we focus in this review on proteins with well-established functions.

a. Actin. Actin, one of the most abundant proteins in eukaryotic cells, constitutes 5% or more of cell protein and, with other cytoskeletal proteins, is a critical target for nitration-induced functional impairment because of its high percentage of tyrosine residues. The functional consequences of tyrosine nitration on actin dynamics observed in several pathologies, such as human inflammatory bowel disease or sickle cell disease, can be the loss of control of filament formation, with subsequent alterations in cell motility, attachment, and intracellular transport (10, 58, 148). ONOO⁻ is able to induce tyrosine nitration of β -actin and dysfunction of the membrane barriers *via* the disruption of the cycling of the actin monomers and actin polymerization; however, direct evidence of nitration of β -actin and dysfunction of the membrane barriers has not been demonstrated (228).

b. α -Tubulin. α -Tubulin has also been identified as a target of nitration associated with the more stable fraction of the cytoskeleton in neuron-like differentiating PC12 cells (41). Concerning this protein, it has been proposed that nitration could play a novel physiologic role in the stabilization of neurites occurring in differentiated neuronal cells (41). Moreover, disruption of microtubule assembly and architecture in intestinal epithelium as a consequence of the nitration of α -tubulin has been described *in vitro*, suggesting novel effective strategies to suppress structural damage in the intestinal epithelium (18).

c. Calpains. In skeletal muscle, calpains are thought to initiate cytoskeletal degradation by cleaving important proteins involved in linking components of the cytoskeleton together and to the cell membrane. Among these elements, talin is co-localized with calpain at the cell membrane. When talin is cleaved by calpain, a breakdown of the muscle cytoskeleton takes place (129). The inhibition of calpain activity through S-nitrosylation impairs talin proteolysis (159), suggesting that NO[•] protects against cytoskeletal degradation.

d. N-Ethylmaleimide sensitive factor (NSF). Endogenous NO[•] and NO[•] donors are able to regulate exocytosis by S-nitrosylating NSF, a cytosolic protein necessary for intracellular vesicular transport, thereby inhibiting NSF disassembly activity (199). This mechanism, which could regulate a variety of physiologic processes mediated by granule exocytosis, has been proposed as a plausible explanation for the antiinflammatory effects of NO[•].

3. Matrix metalloproteinases (MMPs). This family of extracellular soluble or membrane-bound proteases is involved in remodeling the extracellular matrix. Through S-nitrosylation, NO[•] donors induce MMP-9 activation, which can be relevant under certain pathophysiologic conditions, such as cerebral ischemia (108). Thus, extracellular proteolysis cascades triggered by MMPs can disrupt the extracellular matrix, contribute to cell detachment, and provoke apoptosis (108). MMP-2 produced by cultured vascular smooth muscle cells also is nitrated and activated by ONOO⁻; this mechanism may be important for modulation of MMP activity in cardiovascular pathologies in which extracellular matrix plays a key role, as described later (262).

4. Proteins involved in cell cycle (proliferation). Interactions of RNS with signaling molecules involved in essential pathways related to the cell cycle may affect important biologic processes such as proliferation and differentiation. RNS affect proliferation by direct or indirect interactions with different targets, such as cyclins (the elements involved directly in the cell cycle), the kinases regulating cyclins, and the growth factors or their receptors or both.

Few data concerning the specific effects of RNS on growth factors are available. Among the growth factors playing a critical role in cell growth and development, neuregulin-1 (NRG-1), a growth factor strongly expressed in lung, can be nitrated at three tyrosine residues. This posttranslational modification is critical for the binding of NRG-1 to its receptor, resulting in a loss of ligand-induced proliferation, at least in human breast adenocarcinoma *in vitro* (227). An alteration of the epidermal growth factor (EGF) by nitration has also been observed in the intestinal epithelium, in particular in Caco-2 cells. ONOO⁻ inhibits EGF-induced cell proliferation by abolishing EGF-stimulated receptor autophosphorylation and by inducing the nitration of the EGF receptor on tyrosine residues (322). Nerve growth factor (NGF), a growth factor necessary for the differentiation and survival of specific neuronal populations during development, is tyrosine nitrated by ONOO⁻, which transforms NGF into an apoptotic agent for rat motoneuron cultures (244). This gain of apoptotic function of NGF is the result of the formation of high-molecular-weight oligomers, because nitrated tyrosine facilitates covalent cross-links between

NGF molecules (244). Nitrated NGF could be formed during pathologic and inflammatory conditions under which NGF upregulation coincides with increased production of ONOO⁻ or other RNS, as observed in mice models of amyotrophic lateral sclerosis (ALS) (188). Increased S-nitrosylation of cysteines of insulin-like growth factor-1 (IGF) after *in vitro* exposure to RNS suggests that the formation of S-nitrosothiols may result in functional modifications of IGF, possibly leading to the chondrocyte insensitivity to IGF-1 observed in articular diseases (304). Finally, fibroblast growth factor (FGF-1), an inducer of angiogenesis, is nitrated *in vitro* on tyrosine residues critical for its activity, by low levels of ONOO⁻, leading to a nonreversible inactivation of FGF-1. Because FGF-1 regulates angiogenesis, these modifications may alter the ability of tissue repair of this factor (14).

Another target of RNS is the Src family of nonreceptor tyrosine kinases, which also regulate cell proliferation and are activated mainly by phosphorylation at key tyrosine residues. However, ONOO⁻-induced nitration of tyrosine residues of this family of kinases has been also described (160, 254). Nitration of Src prevents phosphorylation of these residues, leading to the decrease of Src activity in intestinal epithelial cell lines (25, 41) and *in vitro* studies (160). These results reinforce the notion that nitration may modify cascades that control signal-transduction processes and regulate cell cycles.

RNS can indirectly modify the signaling pathway of mitogen-activated protein kinases (MAP kinases). In particular, p21^{ras}, an upstream element in the MAP kinase cascade, is nitrosylated by RNS. By using human transfected T Jurkat cells, Lander's group (167) showed that RNS stimulate p21^{ras} and MAP kinase activities, suggesting that S-nitrosylation of p21^{ras} may be responsible for triggering downstream signal transduction. These effects are associated with the increase in Raf-1 kinase activity, a key effector of p21^{ras} function (70).

In parallel, RNS act directly on MAP kinase cascades. MAP kinase pathways activate ERK, JNK, and p38 kinases playing an important role in the regulation of a variety of cell functions (164). For example, angiotensin II nitrates tyrosine residues of ERK1/2 by inducing ONOO⁻ production and facilitates their activation in vascular smooth muscle cells (252). ERK proteins are also phosphorylated by ONOO⁻ in rat cardiomyocytes, but, in this case, ONOO⁻ acts through oxidative but not nitrosative reactions (249). ERK-1 also can be endogenously S-nitrosylated in endothelial cells and, under hyperglycemic conditions *in vitro*, a reduction of endogenous S-nitrosylation has been described, suggesting that alteration in protein S-nitrosylation may underlie the adverse effect of hyperglycemia on the cardiovascular system (338). Like ERK, c-Jun N-terminal kinases (JNKs) are also affected by S-nitrosylation. NO[•] donors inhibit the interaction between JNK1 and its molecular target, the transcription factor c-Jun through S-nitrosylation (242). Some contradictions concerning the direct effects of ONOO⁻ on JNK cascade have been reported. Whereas in rat hepatocytes, ONOO⁻ has no effect on JNK activity activated by Fas ligand (267), in murine alveolar epithelial cells, ONOO⁻-mediated JNK activation depends on Fas (295).

Finally, p38 MAPK can be nitrated in endothelial cells from preeclamptic pregnancies, and this was associated with reduced catalytic activity of p38 MAPK (344).

Previous studies have shown that the p85 regulatory subunit of PI3-kinase is a target for ONOO⁻-induced protein nitration on tyrosine (117). More recently, it was suggested that the increased tyrosine nitration on the p85 subunit of PI3-kinase blocks both PI3-kinase and Akt kinase activities, and this effect is accompanied by a disruption of vascular endothelial growth factor (VEGF)-induced prosurvival function in endothelial cells (80). By contrast, ONOO⁻ has been shown to promote the nitration of regulatory sites at tyrosine kinase receptors coupled to the PI3-kinase/Akt pathway, to induce its activation and thereby to prevent cell death in fibroblasts and neurons (68, 154). Some RNS can be relevant second messengers for the activation of the survival kinase Akt through a mechanism involving the tumor-suppressor phosphatase and tensin homologue (PTEN) by S-nitrosylation in bovine pulmonary endothelial cells (43). PTEN, as a phosphoinositide 3-phosphatase that antagonizes PI3-kinase action, negatively regulates cell proliferation and survival signals (193).

Not only is tyrosine nitration responsible for modifications induced by RNS on proteins involved in cell proliferation, but also, S-nitrosylation of proteins regulating cell survival has been described. Several studies have demonstrated that NO[•] donors can inhibit tyrosine phosphatase activity by S-nitrosylation, leading to their reversible inactivation (21, 177). Because tyrosine phosphatases are important in controlling the VEGF-induced endothelial-proliferation response necessary for tumor progression (36), RNS may provide possible new strategies for the development of anticancer therapies.

Erwin *et al.* (81) were the first to describe receptor-modulated reversible S-nitrosylation of eNOS in bovine aortic endothelial cells. eNOS is constitutively S-nitrosylated in resting endothelial cells; after addition of the eNOS agonist VEGF, eNOS is denitrosylated and then progressively re-nitrosylated. The re-nitrosylation of eNOS after VEGF treatment is markedly attenuated by the addition of NOS antagonists, indicating that eNOS S-nitrosylation depends acutely on eNOS activity. This elegant study suggests that changes in cellular redox state might influence the denitrosylation of eNOS and provides a partial explanation for the attenuation of eNOS activity by oxidative stress.

5. Proteins involved in cell death. Programmed cell death, or apoptosis, is a regulated, energy-dependent process of cell shrinkage, plasma membrane blebbing, chromatin condensation, and DNA fragmentation. Caspases represent the central effectors of cell death. They are a family of cysteine proteases that propagate death signals by cleaving a number of cellular protein substrates. NO[•] can prevent or induce apoptosis, depending on its concentration, cell type, or the oxidative environment. NO[•] inhibits apoptosis by S-nitrosylation of the active site of caspases (see the excellent review, ref. 147). The ability of NO[•] to S-nitrosylate caspases depends, as for a large number of other proteins, on multiple factors including the presence of free iron and intracellular redox potential. Bcl-2, an antiapoptotic protein that regulates the mitochondrial death pathway, can be S-nitrosylated by endogenous NO[•] in a model of human lung epithelial cancer cells (13). By using transfection approaches, these authors show that S-nitrosylation of Bcl-2 prevents its ubiquitin-proteasomal degradation, suggesting a novel antiapoptotic

mechanism that may represent a potential way to control cancer progression.

In contrast, in other models, NO[•] promotes apoptotic pathways in numerous cell types through indirect activation of caspases. NO[•] donors induce cytotoxic effects in bovine chromaffin cells by upregulation of p53 tumor suppressor, by increasing the ratio of proapoptotic to antiapoptotic mediators (Bax and Bcl-Xs *vs.* Bcl-2 and Bcl-XL), cytochrome *c* release, and caspase activation (247). The S-nitrosylation of cytochrome *c* during Fas-induced apoptosis of human mononuclear cell lines enhances caspase activation, resulting in the induction of apoptosis (285). Cytochrome *c* also can be nitrated (220), and in this case, inhibition of apoptosis has been observed. In rat glioma cells, ONOO⁻ is able to nitrate several tyrosine residues of cytochrome *c* and thereby decreases its effects on caspase cascade activation (220).

Apoptosis can also be influenced by tyrosine nitration. In rat and human hepatocytes, it has been shown that ONOO⁻ induces CD95-tyrosine nitration, which is associated with the prevention of CD95-tyrosine phosphorylation and apoptosis (267). *In vivo* CD95-tyrosine nitration has been described in livers from endotoxin-treated rats (267). Altogether, these results suggest that tyrosine nitration and phosphorylation of CD95 are mutually exclusive and that both mechanisms can be observed *in vivo*.

In parallel, the status of intracellular redox elements can affect the ability of the cell to become apoptotic. Several mechanisms have been proposed to explain the effects of S-nitrosylation on the antiapoptotic functions of thioredoxin, a redox regulator implicated in the control of cell growth and apoptosis inhibition, which is S-nitrosylated under basal conditions (110). First, overexpression of S-nitrosylable thioredoxin reduces ROS formation in endothelial cells (110); and second, in *in vitro* T cells, thioredoxin may act by trans-nitrosylation of proteins such as caspases, through the formation of a protein-protein complex, and inhibit their activity (211). Thus, the basal S-nitrosylation of thioredoxin is a requirement for preserving its redox regulatory activity and for scavenging ROS in endothelial cells. Moreover, NO[•] donor exposure results in thioredoxin nitration and abolishes thioredoxin activity *in vitro* (316). Thus, inactivation *via* nitration of thioredoxin might play a proapoptotic role in pathologic states in which production of RNS is increased.

6. Proteins implicated in cell metabolism. Mitochondria are the central organelles producing cellular energy. NO[•] donors and ONOO⁻, in *in vitro* tests, inhibited NADH:cytochrome *c* reductase activity by nitration, whereas succinate:cytochrome *c* reductase activity remained unaffected (275); thus, one can advance the hypothesis that RNS may limit the energy supply in tissues. Other enzymes implicated in the metabolism, such as succinyl-CoA:3-ketoacid coenzyme A transferase (the mitochondrial enzyme involved in the breakdown of ketone bodies in the extrahepatic tissues) in rat heart mitochondria (264), the purified pyruvate dehydrogenase complex (the mitochondrial matrix enzyme catalyzing the oxidative decarboxylation of pyruvate) (274), and cytosolic aconitase from murine macrophages (103) can be nitrated or S-nitrosylated by RNS. Creatine kinases are critical energy controllers that regulate cardiac myocyte contractility in both physiologic and pathologic states. An impairment of this system during cardiac failure may be the consequence of

the nitration and inactivation of creatine kinases in isolated cardiac trabeculae from mice (205). Furthermore, S-glutathionylation of creatine kinases has been proposed as a parallel mechanism explaining the temporary reversible loss in activity of this system during ischemic injury (265).

a. Enzymes playing a key role in insulin-associated cascades. Glucokinase activity is a major determinant of β -cell glucose metabolism and insulin secretion. An increase of glucokinase activity by insulin through the S-nitrosylation of this enzyme has been described in cultured β cells, as well as its association with nNOS, suggesting that this mechanism represents a sensitive means for modulating glucokinase activity and insulin secretion (276).

Much evidence indicates that S-nitrosylation as a key mechanism in the regulation of insulin secretion and resistance. In brief, when insulin binds to its receptor, dimerization and autophosphorylation of insulin receptor tyrosine kinase take place. Then, tyrosine phosphorylation of insulin-receptor substrates leads to activation of the Akt/PKB pathway. This pathway plays a central role in glucose transport, protein and glycogen synthesis, and inhibition of hepatic gluconeogenesis, and all of these effects regulate insulin actions (for a review, see ref. 145). In addition to enzymes implicated in glucose metabolism (see earlier), the Akt/PKB pathway is reversibly inactivated by S-nitrosylation *in vitro* and in intact cells (354). In this study, it has been shown that NO^\bullet donors inhibit Akt/PKB at the cysteine 224, and this process is increased in skeletal muscle of diabetic mice, suggesting that S-nitrosylation-mediated inactivation of Akt/PKB could contribute to the pathogenesis of NO^\bullet -mediated insulin resistance, at least in mice. Moreover, NO^\bullet donors may directly inhibit insulin secretion partly by S-nitrosylation of key proteins in the stimulus-secretion coupling (298). Furthermore, insulin may be a potential target of ONOO^- . The ONOO^- -mediated nitration of insulin partially affects its receptor binding and hypoglycemic capacities in a mouse model (51). At least four nitrated tyrosine residues in insulin receptor substrate-1 have been detected (230). Interestingly, among these residues, Tyr939, which is critical for association of insulin-receptor substrate-1 with the p85 subunit of PI-3 kinase, is nitrated (230). Very recently, a novel molecular mechanism resulting in insulin resistance was proposed. Thus, Clavreul *et al.* (57) showed that S-glutathionylation of p21^{ras} by ONOO^- is involved in the alterations in the insulin-signaling pathway induced by LDL, at least in bovine aortic endothelial cells. Altogether, these findings indicate that RNS may modulate insulin resistance and type 2 diabetes at different levels, leading to a sustained activation of inflammatory pathways.

7. Nuclear factors. NF- κ B family members are heterodimers consisting of p65/RelA and p50/NF- κ B1, but only the p65 subunit has transactivation domains capable of initiating transcription (69). NF- κ B is complexed in the cell cytoplasm with the inhibitory proteins, I κ B α and I κ B β . On cell stimulation, I κ B is phosphorylated, removed, and degraded, allowing NF- κ B to induce transcription. S-Nitrosylation of p65 by GSNO, which consequently inhibits NF- κ B activation, has been shown in endothelial cells (Fig. 8). In addition, S-nitrosylation of p65 is associated with an inhibition of monocyte adhesion to the activated endothelial cells *via* the

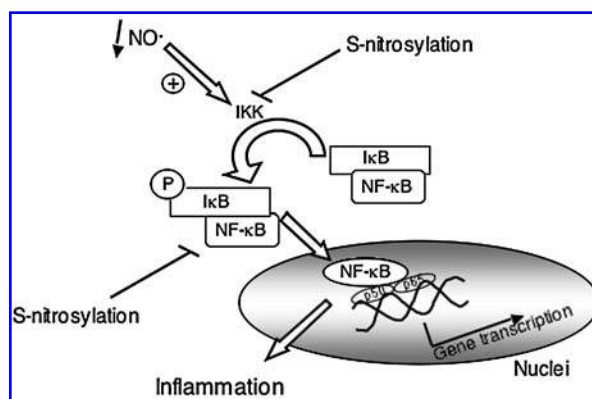


FIG. 8. Alterations of the NF- κ B pathway by RNS. The decrease in NO^\bullet production, after NOS inhibition, is able to activate IKK, which is necessary to phosphorylate I κ B and the subsequent activation of the NF- κ B pathway. S-Nitrosothiols are implicated both in the direct and indirect inhibition of NF- κ B. Arrows without heads, inhibition.

downregulation of endothelial cell adhesion molecules (256). These authors suggest that GSNO could have a protective effect in several pathophysiologic conditions by inhibition of cellular infiltration of monocytes (256). In contrast, the inhibition of NF- κ B by NO^\bullet donors could represent one mechanism by which S-nitrosylation can initiate apoptosis in A549 cells (191) (Fig. 8). By using mutant versions of inhibitory κ B kinase (IKK β), the prerequisite enzyme complex necessary to induce NF- κ B, Reynaert *et al.* (269) demonstrated that cysteine-179 is the main target of IKK β by S-nitrosothiols (S-nitroso-N-acetyl-D,L-penicillamine (SNAP) and GSNO), leading to inhibition of its enzymatic activity. Importantly, when NOS was inhibited, IKK β was activated and denitrosylated, illustrating the importance of endogenous NO^\bullet in regulating the extent of NF- κ B activation.

8. Proteins transporting oxygen. Under physiologic conditions, reactions of endothelium-derived NO^\bullet with hemoglobin are considered to be the most important pathway for limiting NO^\bullet bioactivity (102). This reaction involves the iron-containing heme groups of oxy- and deoxyhemoglobin with NO^\bullet and results in production of methemoglobin and NO_3^- ions and iron-nitrosyl-hemoglobin, respectively (100). Much evidence has shown that NO^\bullet reacts with hemoglobin to form stable metabolites, which may be considered means of transportation and subsequent release of NO^\bullet . Cannon *et al.* (40) showed that, in humans, the inhalation of NO gas is associated with increases in plasma NO_2^- levels, as well as iron-nitrosyl-hemoglobin, and they suggest that NO_2^- may be reduced to NO^\bullet in the circulation and contribute to endocrine NO^\bullet delivery. Hemoglobin possesses an allosteric regulated NO_2^- reductase activity that reduces NO_2^- to NO^\bullet along with the physiologic oxygen gradient, and this potentially contributes to hypoxic vasodilatation (101). In addition, NO^\bullet may be stabilized, transported, and released by intramolecular NO^\bullet group transfers between the heme iron and cysteine residues to form S-nitroso-hemoglobin, which, under hypoxic conditions, delivers S-nitrosothiol and induces relaxation (151). Therefore, hemoglobin derived from red cells may regulate the vascular tone, depending on tissue

oxygen tension. However, this hypothesis has been argued recently (102, 141). Thus, direct measurement by electron paramagnetic resonance of S-nitroso-hemoglobin has shown that, although its formation in either venous or arterial blood depends inversely on oxygen concentration, S-nitroso-hemoglobin gradients across the circulation are not described (141).

More recently, by using new mouse models that exclusively express either the human wild-type of hemoglobin or human hemoglobin in which the β Cys93 has been replaced with alanine, Isbell *et al.* (136) showed that S-nitroso-hemoglobin is not required for red cells to stimulate hypoxic vasodilatation. Altogether, these data suggest that S-nitroso-hemoglobin might not be critical for the coupling of red cell deoxygenation with increased NO[•] bioactivity.

It has been reported that myoglobin, an important intracellular oxygen binding hemoprotein in heart and skeletal muscle, can be nitrated and probably S-nitrosylated (229, 288). In both cases, myoglobin acts as a scavenger of NO[•] and may limit the deleterious effects of RNS.

9. Albumin. Albumin is the most abundant transport and depot protein in the mammalian vasculature. Taking into consideration that albumin is present in plasma at elevated concentrations (>0.5 mM), it is plausible to imagine that it can transport free NO[•]. Indeed, Rafikova and colleagues (261) showed that albumin accelerates formation of low-molecular-weight S-nitrosothiols by transferring NO⁺. These authors suggest that albumin could represent a major reservoir of NO[•] and its reactive oxides controlling the dynamic of NO[•]-dependent processes in the vascular system.

In addition, ONOO⁻, which can lead to protein-bound tyrosine nitration, is able to induce mild nitration on albumin and low-density lipoprotein (LDL)-albumin complex. Interestingly, human serum albumin decreases tyrosine nitration in this complex *in vitro*, raising the question of the pathophysiologic significance of these nitrations and their interactions, which may potentially prevent actions of LDL proteins on endothelial cells (320). Finally, a protective effect of albumin treatment has been reported in experimental endotoxic shock by its capacity to reduce the inflammatory process leading to oxidative and nitrosative stresses and vascular hyporeactivity in mice (204).

10. Enzymes involved in prostaglandin generation. Cyclooxygenase and prostacyclin synthase. Cyclooxygenase (COX) catalyzes the first step in the synthesis of prostaglandins, which are critically implicated in a variety of physiologic and pathophysiologic processes, including inflammation, vascular homeostasis, and immunity (61). Nitration of the proinflammatory isoform COX-2 and the increase in COX-2 expression induced by ONOO⁻ after treatment with arsenic have been detected in bovine aortic endothelial cells (34), suggesting that COX-2 may exacerbate the inflammatory state in the cardiovascular system under situations in which ONOO⁻ production is increased (Fig. 9). In contrast, NO[•] inhibits COX-2-derived prostaglandin E₂ production in mouse macrophages, and this effect is associated with both decreased expression and nitration of COX-2, indicating that the regulation of COX-2 by NO[•] is complex and may depend on the local environment of the cell (56) (Fig. 9). In human platelets, ONOO⁻ is able to induce nitration in tyrosine

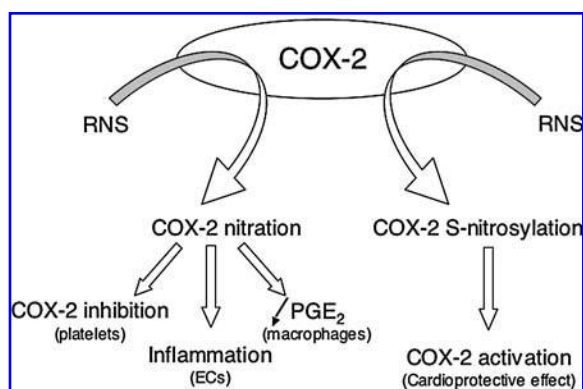


FIG. 9. Effects of RNS on cyclooxygenase (COX)-2. RNS affect COX-2 activity differently, depending on the cell type. Nitration of COX-2 leads to COX-2 inhibition in platelets, to a decrease of prostaglandin E₂ (PGE₂) production in macrophages, and to an increase of inflammatory responses in endothelial cells (ECs). By contrast, S-nitrosylation of COX-2 may have a cardioprotective effect by increasing its activity.

residues of both COX-1 and COX-2 and the subsequent inhibition of their activities (30). S-Nitrosylation of cysteine residues of COX by RNS has been described in murine macrophage cell line (RAW264.7) (325). S-Nitrosylation of COX-2 may have a cardioprotective effect, because it has been shown that, in cardiomyocytes, atorvastatin (an agent that improves cardiovascular function) is able to increase the activity of iNOS and activate COX-2 by S-nitrosylation (11) (Fig. 9).

It has been shown that prostacyclin synthase is inhibited by ONOO⁻-dependent nitration in human aortic endothelial cells (64), indicating that this mechanism may be involved in several pathologies associated with endothelial dysfunction, such as atherosclerosis or diabetes. Prostacyclin synthase in smooth muscle cells, in contrast to endothelial cells, is resistant to nitration and inhibition by ONOO⁻ (155).

11. Mn superoxide dismutase (MnSOD). SOD catalyzes the dismutation of O₂^{•-} to hydrogen peroxide and molecular oxygen. Among SODs, MnSOD, the first line of defense against O₂^{•-} generated in mitochondria, is the only one to be modified by RNS. Indeed, MnSOD is nitrated and inactivated in neurodegenerative diseases (ALS, Parkinson, and Alzheimer diseases) and atherosclerosis (9, 350). This nitration and the subsequent inactivation of MnSOD could lead to the exacerbation of oxidative stress and could explain the mitochondrial dysfunction described under these pathologic situations. Other authors suggest that the interaction of NO[•] with MnSOD in *Escherichia coli* may represent a novel mechanism by which MnSOD protects the cell from deleterious effects associated with overproduction of NO[•] (89).

V. RNS in Physiologic and Pathophysiologic Conditions

A. Physiology

Shear stress is one of the most important stimuli for the synthesis and release of NO[•] (35). Agonist-dependent recep-

tor activation also is able to release NO[•], mainly from eNOS (35) but also from nNOS (290). Whereas in the cardiovascular system, the effects of NO[•] released from eNOS are reflected by vasorelaxation, inhibition of platelet aggregation, and inhibition of vascular smooth muscle cell proliferation, NO[•] generated from nNOS, acting as a neurotransmitter in nonadrenergic, noncholinergic nerves, is involved in synaptic transmission and plasticity in the cerebral cortex and hippocampus (127). Furthermore, NO[•] from nNOS is also implicated in the regulation of basal and β -adrenergic myocardial function (44) and, in association with eNOS, in the regulation of penile erection (130). The main physiologic effect of NO[•] is the activation of GC and the subsequent cGMP production (Fig. 10). NO[•] reacts directly with the heme group of GC; thus, we can consider that the receptor for NO[•] is GC. At least seven different isoforms of GC have been identified. The soluble isoform of GC is a heterodimer with an α subunit and a β subunit. When NO[•] binds to the ferrous heme iron, the inhibition of the catalytic activity of GC by the heme is removed (195). The levels of intracellular cGMP are regulated by the ratio between its formation and the rate of its degradation and inactivation by phosphodiesterases. The latter represents a way to regulate the activity of NO[•]. Once cGMP is produced, it interacts with cGMP-dependent protein kinase, which is present at high concentrations in all smooth muscle cells, cardiomyocytes, platelets, and nervous and juxtaglomerular cells (125). Although several substrates of cGMP-dependent protein kinases have been described, this enzyme inhibits receptor-induced vascular smooth muscle contraction mainly by three mechanisms involving inositol-1,4,5-trisphosphate receptor-associated cGMP kinase substrate (IRAG), the regulator of G protein signaling (RGS2) and the myosin light-chain

phosphatase. All these mechanisms account for reducing the intracellular Ca²⁺ concentration or for decreasing the Ca²⁺ sensitivity of contraction (or both), leading to the vasorelaxant effects of NO[•] and NO[•] donors. In addition, NO[•] increases cGMP levels in platelets and inhibits platelet activation by direct interaction of cGMP with the cGMP-dependent protein kinase that phosphorylates vasodilator-stimulated phosphoprotein (VASP) and IRAG (185). Interestingly, it has also been proposed that cyclic ADP ribose (cADPR), an endogenous modulator of RyR, is involved in the regulation of endothelial function, by participating in endothelial NO[•]-dependent vasodilator response (121). Thus, in bovine coronary arterial endothelium, bradykinin-induced intracellular Ca²⁺ increase and NO[•] response are associated mainly with cADPR levels, suggesting that cADPR-mediated Ca²⁺ signaling plays an important role in the regulation of endothelial NO[•] production and could play a critical role in endothelium-dependent vasodilatation (360).

NO[•] and NO[•] donors might use pathways independent of cGMP-dependent protein kinase, such as cross-activation of cAMP-associated pathways. This has been described in smooth muscle cells and platelets, although the concentrations of cGMP needed to act on cAMP-dependent protein kinases are 1,000-fold higher than those for cGMP-dependent protein kinase (29).

Some evidence shows that alternative pathways that are cGMP independent exist. Indeed, the selective inhibitor of sGC 1H-[1,2,4]oxadiazolol [4,3- α]quinoxalin-1-one, ODQ, is not able to inhibit several effects mediated by NO[•] and NO[•] donors, at high concentrations (micromolar to millimolar range) in blood vessels and platelets (341). NO[•] and NO[•] donors can elicit vasorelaxation through a cGMP-independent mechanism, probably *via* GSNO formation and, in this way, protect cells from further oxidative and nitrosative modifications (308). Other posttranslational modifications such as protein nitration are independent of enzyme involvement. Whether protein nitration is important in normal physiologic mechanisms is unclear, but it is probable that alterations in nitration levels may contribute to different biologic outcomes (158) (see later).

ONOO⁻ generated under physiologic conditions can modulate cell signaling. In particular, tyrosine nitration by ONOO⁻ could modify phosphotyrosine-dependent cell signaling. ONOO⁻ has been reported to promote inhibition or activation of tyrosine phosphorylation in several cell types (238). Thus, ONOO⁻-mediated nitration of critical tyrosine residues in proteins could block downstream signaling, affecting essential cellular functions, as has been observed in bovine pulmonary artery endothelial cells and rat macrophages (104, 117). Conversely, in PC12 cells, ONOO⁻ activates members of MAP kinases, resulting in the gain of function in the upstream signaling of these proteins (144).

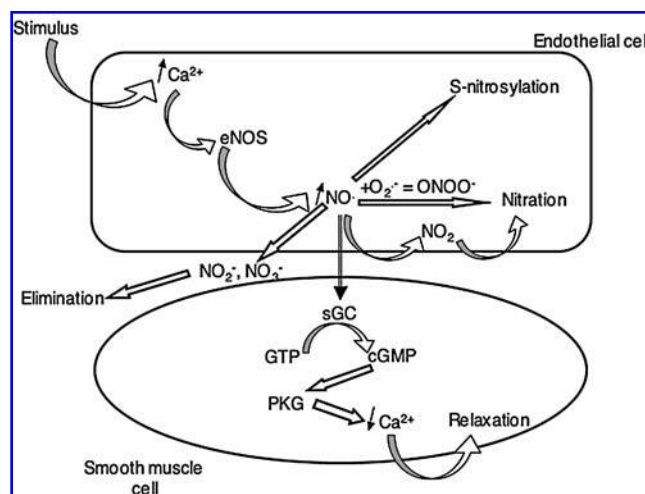


FIG. 10. Different effects of NO[•] in vascular cells. The increase of NO[•] after activation of calcium-dependent eNOS in endothelial cells is followed by activation of soluble guanylyl cyclase (sGC) in smooth muscle cells, which produces cGMP as second messenger. Through the activation of protein kinase G (PKG), cGMP is able to decrease intracellular Ca²⁺ concentration and Ca²⁺ sensitivity in smooth muscle cells, leading to vascular relaxation. NO[•] can react with O₂⁻ in endothelial cells and generate ONOO⁻, accounting for nitration, or NO[•] can induce S-nitrosylation of proteins.

B. Pathophysiology

Under pathologic conditions, RNS production increases and exceeds the cell defenses, inducing deleterious effects in a large number of pathways implicated in cell life. Here, we review the effects of RNS on four categories of pathologies. Taking into consideration that the brain is, with the cardiovascular system, the highest target of damage from nitrosative stress, it seems obvious to start this part of the re-

view by the link between RNS and neurodegenerative diseases, and then to continue with the effects of RNS on the cardiovascular system. Third, we review the role of nitrosative stress in immunologic diseases and cancer, and finally, we summarize the involvement of RNS in the development of metabolic diseases. However, other deleterious effects of RNS on other systems cannot be excluded.

1. Neurodegenerative diseases. Neurodegeneration is associated with selective neuron loss. Among the regions of the brain, hippocampus and the substantia nigra are the most susceptible to cell damage; thus, in Alzheimer's disease, hippocampal degeneration has been mainly described, whereas substantia nigra is affected in Parkinson's disease (236). Many studies indicate the participation of apoptosis in neuronal cell death and in neurodegenerative diseases (for a review, see ref. 236). In general, the excessive production of RNS (and ROS), in addition to the deficiency of antioxidant defense mechanisms, is responsible, at least in part, for the functional deterioration in aged brain and neurodegenerative disorders.

Although the three isoforms of NOS are expressed in the nervous system, NO[•] released from iNOS possesses both neuroprotective and neurotoxic effects. At low concentrations, NO[•] elicits vasodilatation and is involved in neurotransmission; by contrast, at higher concentrations, it is potentially neurotoxic (240). This is of particular significance in neurodegenerative diseases such as Alzheimer's and Parkinson's diseases, ALS, and multiple sclerosis. A common aspect of many neurodegenerative disorders is the accumulation of misfolded proteins critical for neuron life, which triggers cell death and loss (216). Indeed, Alzheimer's and Parkinson's diseases are characterized by accumulations of aggregated amyloid β peptides and tau, and α -synuclein and synphilin-1, respectively (83, 156, 197, 214). With regard to Alzheimer's disease, deposition of the long form of amyloid β , amyloid β 42, in the brain is an early and invariant feature of all forms of the disease, even before a diagnosis of Alzheimer's disease, and before the deposition of the shorter primary form of amyloid β , amyloid β 40, occurs (for a review, see ref. 90). Also, the ubiquitin-proteasome system plays an essential role by acting on the removal of aberrant proteins, but in these pathologies, this system is altered, and the accumulation of misfolded proteins can evoke neurodegeneration (216).

a. Alzheimer's disease. Amyloid β peptide can stimulate production of NO[•] through the activation of iNOS, as demonstrated by the use of specific inhibitors of iNOS and iNOS knockout mice (200). The large amount of NO[•] generated leads to ONOO⁻ production, which mediates neuronal damage. Indeed, protein tyrosine nitration can be detected in postmortem samples from Alzheimer patients (118). Specific nitrated proteins in the Alzheimer's disease hippocampus were identified immunochemically by Sultana *et al.* (306) by using a redox proteomics approach. These proteins were related to the control of energy production (α -enolase, glyceraldehyde-3-phosphate dehydrogenase, ATP synthase α chain, voltage-dependent anion channel protein 1, and carbonic anhydrase II). In addition to those in brain, peripheral lymphocytes and plasma from these patients present high levels of markers linked to nitrosative stress that are associated with increased expression of iNOS (*i.e.*, heme oxygenase-1, heat-shock protein 60, heat-shock protein 72, and thioredoxin) (37). Nitration of tau protein, which promotes tubulin polymerization and maintains microtubule stability, has been detected in Alzheimer's disease (271). This may inhibit the ability of tau to self-associate and stabilize microtubules, as demonstrated by using an *in vitro* purified protein model (270, 359) (Fig. 11). In addition, amyloid β peptide could increase the activity of NMDA-type glutamate receptor with a subsequent increase of intracellular Ca²⁺ (see earlier). This increase in intracellular Ca²⁺ could elicit the release of RNS from NOS. RNS could induce S-nitrosylation of NMDA receptors, decrease their activity, and favor neuronal survival; in this way, RNS could be neuroprotective in Alzheimer's disease (180) (Fig. 11). In contrast, S-nitrosylation of endoplasmic reticulum chaperones, such as protein-disulfide isomerase, can be deleterious by inducing accumulation of misfolded proteins or contributing to excitotoxic neuronal damage (323) (Fig. 11).

b. Parkinson's disease. Parkinson's disease, the second most common neurodegenerative disease after Alzheimer's disorder, is characterized by a prevalent degeneration of dopaminergic neurons localized in the substantia nigra. RNS-dependent mechanisms contribute to neurodegeneration in Parkinson's disease, as shown by the increased expression of iNOS in Parkinson experimental models associated with dopaminergic neuronal loss (178, 237). Intraneuronal aggregates of α -synuclein, a protein that reg-

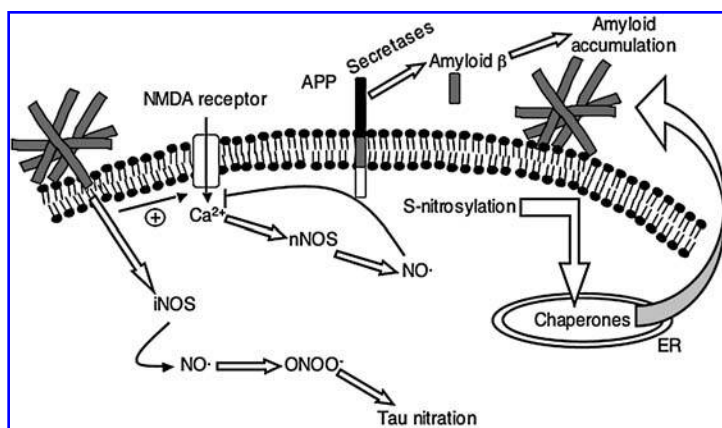


FIG. 11. Changes induced by RNS on critical proteins in Alzheimer's disease. Amyloid- β comes from the transmembrane helix of the amyloid precursor protein (APP), through the action of secretases. In neurons, excess generation of NO[•] by iNOS activated by amyloid- β peptide is responsible for the nitration of proteins such as tau. Amyloid- β increases the activity of the NMDA channel, with the subsequent increase of intracellular Ca²⁺ concentration, activation of Ca²⁺-dependent nNOS, and generation of NO[•] and other RNS that could inhibit NMDA activity by S-nitrosylation. In addition, S-nitrosylation of chaperones can elicit accumulation of misfolded proteins.

ulates the trafficking of lipid secretory vesicles, have been described in Parkinson's disease. α -Synuclein possesses several tyrosine residues, which can be nitrated, as described in postmortem human brain and in mouse models (98, 258). With structural and conformational analysis of purified human α -synuclein, it has been reported that such modifications induce the formation of α -synuclein oligomers and the inhibition of filament assembly, decrease the ability of α -synuclein to bind lipid membranes, and reduce proteolysis, favoring aggregate formation (124, 326, 351). Furthermore, other proteins can be affected by RNS in Parkinson's disease. Thus, parkin, a member of the family of E3 ubiquitin ligases that stabilizes microtubules, can be S-nitrosylated, forming SNO-parkin; this has been detected by the biotin-switch assay in Parkinson's brain from either mice or humans (353). However, contradictory data have been described. S-Nitrosylation may stimulate or inhibit E3 ubiquitin ligase activity, depending on the models (55, 353). As in Alzheimer's disease, S-nitrosylation of endoplasmic reticulum chaperones such as protein-disulfide isomerase can be involved in the protein misfolding or neurotoxicity (323).

c. Amyotrophic lateral sclerosis (ALS). ALS is a neurodegenerative disorder characterized by motoneuron death. In this pathology, increased expression of iNOS enhances the production of NO^\bullet and ONOO^- and triggers apoptosis in motor neurons (19). Other enzymes involved in the ROS pathways, such as Cu,Zn SOD and Mn SOD, have also been associated with the pathogenesis of ALS. Whereas Cu,Zn SOD catalyzes nitration by ONOO^- , Mn SOD is selectively nitrated, leading, at least in part, to the neuronal damage in ALS (272). Finally, neurofilament aggregates colocalize with nitrotyrosine proteins in upper and lower motoneurons, suggesting that nitration occurs in these deteriorating cells (53). In a transgenic model of mice overexpressing the mutant Cu,Zn SOD from humans with familial ALS, an elevated protein nitration and NO^\bullet synthesis in brain tissues and cerebrospinal fluid were reported (45, 181).

d. Prion diseases. The transmissible neurodegenerative disorders, also known as prion diseases, include Creutzfeldt-Jakob disease, bovine spongiform encephalopathy, and scrapie. These pathologies are characterized by enhanced apoptosis of neurons associated with cerebral accumulation of misfolded prion protein. In scrapie and Creutzfeldt-Jakob disease, increased expression of iNOS as well as immunoreaction for nitrotyrosine have been described during the disease progression (88, 93).

To summarize, RNS can have either beneficial or deleterious effects, depending on the species and cellular target, on the neuronal signaling pathways involved in the pathophysiology of the neurodegenerative disorders. Although many of the data concerning the effects of RNS on neurodegeneration described in the literature were obtained in animal models, one can hypothesize that RNS may play an important role in human neurodegenerative diseases. However, it remains to be elucidated whether RNS are the cause or the consequence of neuronal degeneration.

2. Cardiovascular diseases. Although the involvement of RNS in cardiac signaling has been clearly demonstrated (112), many important questions still remain unresolved. A

large number of targets for RNS exist in cardiomyocytes, mainly the ion channels and Ca^{2+} pumps implicated in myocyte contraction (1, 65, 192, 202, 340, 347, 348). Besides, NO^\bullet from eNOS inhibits L-type Ca^{2+} channels through the activation of GC (201) and reduces β -adrenergic myocardial contraction (20). In contrast, NO^\bullet from nNOS favors Ca^{2+} movements between sarcoplasmic reticulum and cytosol and increases myocardial contractility (20). Among cardiac pathologies associated with an increase in nitrosative stress, congestive heart failure and myocardial infarction are the most representative. In these diseases, nitrosative stress generated by inflammatory changes that occur in the cardiomyocytes is accompanied by contractile dysfunction (161, 186, 267).

a. Heart failure. Heart failure occurs progressively and affects the heart muscle, resulting in a loss of cardiac myocytes or in a disruption of the ability of the myocardium to generate force and a reduction of effectiveness of the pump (187). In general, it occurs after damage or disorder in the heart, as, for example, coronary endothelial dysfunction as the consequence of myocardial ischemia and infarction, but also after cardiac hypertrophy or cardiomyopathies (187). The nitrosative-oxidative imbalance could contribute to reduction of cardiac performance (112). In failing human heart, reduced NO^\bullet production from the endothelium associated with reduced eNOS activity and eNOS uncoupling have been described (75). A correlation between increased nitrotyrosine formation in myocardial specimens from patients with heart failure and increased iNOS expression has been observed (328). Besides, the production and delivery of NO^\bullet in the form of S-nitrosohemoglobin are impaired in the setting of heart failure, suggesting that NO^\bullet donors could have beneficial effects in this pathology (63). Conversely, iNOS expression is increased in the myocardium after infarction. Thus, in addition to the severe oxidative stress, the increased NO^\bullet production from iNOS increases infarct size, myocardial dysfunction, and mortality in mice (87).

b. Hypertension. Accumulating evidence suggests that alterations in NO^\bullet synthesis and pathways or a reduction in the bioavailability of NO^\bullet plays a critical role in the pathogenesis of hypertension (241). Interestingly, extensive endothelial dysfunction is observed simultaneously with an increase in eNOS expression and $\text{O}_2^{\bullet-}$ production associated with enhanced nitrotyrosine formation (99). Thus, ONOO^- formation and protein nitration have been found to be elevated in the serum, thoracic aorta, and kidneys of different hypertensive animal models and in the human plasma (38, 191, 278).

c. Pulmonary arterial hypertension. Pulmonary arterial hypertension is characterized by increased pressure in the pulmonary arteries, which is associated with right ventricular hypertrophy and remodeling of small pulmonary arteries. Pulmonary arterial hypertension can be caused by hypoxia, and the subsequent hypoxia-regulated genes in the pulmonary endothelium could play an important role (85). Very recently, it was shown that mice treated with the antioxidant N-acetylcysteine develop pulmonary arterial hypertension mimicking the effects of chronic hypoxia (increase of right ventricle pressure, right ventricle hypertrophy, and vascular

remodeling), whereas eNOS-deficient mice were protected from these effects (239). These authors demonstrated that *N*-acetylcysteine is converted to *S*-nitro-*N*-acetylcysteine. They suggest that this conversion during blood deoxygenation is necessary for the development of pulmonary arterial hypertension in this animal model and underline the potential toxicity of *N*-acetylcysteine administration.

d. Atherosclerosis. All of these diseases are the consequences of vascular atherosclerosis. In atherosclerosis and hypercholesterolemia, the NO[•] pathway regulating the vascular tone is altered. A decrease in either expression or activity of both eNOS and iNOS and a local increased scavenging of NO[•] through the reaction with ROS have been suggested. An impaired interaction of NO[•] with GC, and thereby the reduction of cGMP production, also occurs (224). Thus, strategies targeting the increase of NO[•] synthesis or activity or both may be useful in the treatment of atherosclerosis and related diseases (279). In addition to the effects on NO[•] synthesis, prostaglandin synthases located at the human atheroma plaques are inhibited by iNOS-mediated nitration (67), leading to a reduction in prostaglandin production. Furthermore, an increase of circulating nitrated HDL in plasma is correlated with an increased risk of coronary artery disease (246), whereas nitrated LDL has been detected in human thoracic aorta atheroma plaques (172), suggesting that nitrated LDL is retained within the atheroma plaque. In this pathology, MMP-2 is activated by ONOO⁻, underlining the effects of RNS on the regulation of the atherosclerotic plaque rupture (262).

e. Preeclampsia. In another type of inflammatory vascular disease such as preeclampsia, evidence exists of endothelial cell activation and dysfunction. Preeclamptic women are characterized by pregnancies associated with hypertension linked to generalized endothelial dysfunction, proteinuria, and fetal growth delay (113). Although several markers of endothelial dysfunction (reduced endothelium-dependent relaxation and NO[•] synthesis) have been detected, a novel mechanism was recently suggested. Levels of microparticles, small fragments of plasma membrane released from apoptotic or activated circulating cells or cells of the vascular wall (196), are elevated in preeclamptic women. More particularly, shed membrane particles of leukocytes and platelets have the ability to induce vascular inflammation and hyporeactivity, which is associated with increased NO[•] production and upregulation of iNOS (203). Most interestingly, whereas microparticles from both platelets and leukocytes account for nitrosative stress, only leukocyte-derived microparticles induce COX-2 metabolite generation (Fig. 12). These findings demonstrate the pathophysiologic relevance of preeclamptic platelet microparticles, which are pro-inflammatory and lead to significant signs of nitrosative stress in the vascular wall.

3. Inflammation

a. Airways diseases. In inflammation states, NO[•] is produced by iNOS induced by microorganisms, environmental products, and cytokines (for a review, see ref. 84). Together with inflammation and subsequent remodeling of airways (273), RNS are generated during the development of nu-

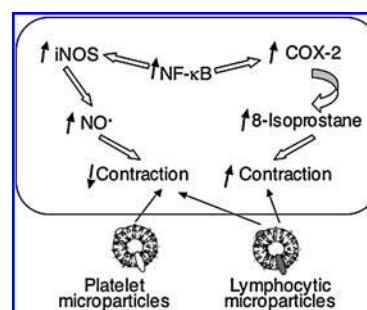


FIG. 12. Microparticles as new mediators of nitrosative stress. In preeclampsia, circulating levels of microparticles from platelets and lymphocytes are elevated. Both types of microparticles induce NF- κ B activation, leading to upregulation of pro-inflammatory proteins, such as iNOS, accounting for overproduction of NO[•] and vascular hyporeactivity. In contrast, only lymphocytic microparticles are able to upregulate COX-2 and generate 8-isoprostane, which evoke hyperreactivity. The balance between both vasorelaxant and vasoconstrictor factors released by the vessel wall results in hyporeactivity.

merous airways diseases such as asthma, chronic obstructive disease, and cystic fibrosis. For example, although all three NOS isoforms are expressed within the respiratory tract, iNOS is strongly induced in the airway epithelium in asthmatic patients (109), and subsequent alterations in the pathways dependent on NO[•] have been described, including airway hyperresponsiveness (273). Thus, elevated concentrations of NO[•] have been observed in the exhaled air of asthmatics (149). In patients with asthma and cystic fibrosis, the levels of *S*-nitrosothiols in the bronchoalveolar lavage fluids are extremely and paradoxically reduced (96, 105), suggesting a disorder of *S*-nitrosothiol metabolism. In addition, an exacerbated nitration of proteins in airway epithelial cells from asthma patients, in particular, the antioxidant enzyme catalase, was recently described (97). Altogether, these results suggest that, in asthma, RNS affect NO[•]-dependent pathways and amplify oxidative stress by decreasing *S*-nitrosothiol bioavailability and catalase activity. These effects probably contribute to the chronic inflammatory state.

Airways inflammation plays an important role in the pathogenesis of chronic obstructive pulmonary disease. In this pathology, markers of nitrosative stress, such as exhaled NO[•] and tyrosine nitration of proteins, have been detected in the sputum and lung specimens from asthma patients (133). In addition, inhibition of iNOS is associated with a reduction of nitrotyrosine, and most important, this is correlated with the improvement in forced expiratory volume and airway responsiveness to histamine, suggesting that RNS are involved in the reversible component of inflammation in chronic obstructive pulmonary disease. Therefore, specific inhibition of RNS actions could represent a novel therapeutic approach for this pathology (305) (Fig. 13). Similar results have been shown during *Candida albicans*-induced acute lung injury. Selective inhibition of iNOS suppresses the production of ONOO⁻, as well as nitrotyrosine and expression of iNOS (235).

b. Sepsis. Sepsis represents a pathology associated with the dysregulation of the NO[•] pathway. Sepsis is the result of

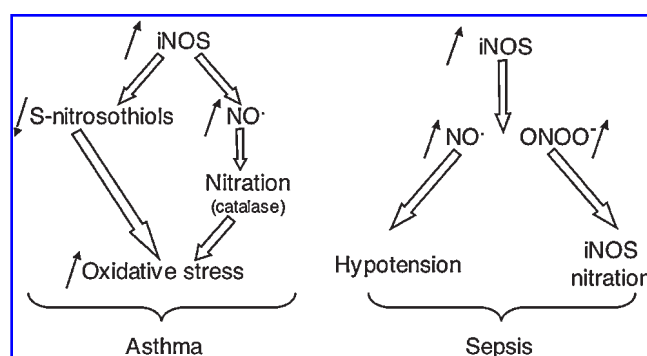


FIG. 13. RNS in inflammatory states. In asthma or sepsis, the increase of iNOS is accompanied by an augmentation of RNS production that induces nitration of proteins implicated in the defenses against oxidative stress, such as catalase or iNOS in asthma or sepsis, respectively. In addition, in asthma, oxidative stress can be increased by the reduction of S-nitrosothiols. Moreover, the excess of NO[•] generated from iNOS accounts for the hypotension in septic patients.

an acute and systemic immune response to a variety of noxious insults, in particular to bacterial infections. This response leads to the activation of a number of host mediator systems, including the cytokine, leukocyte, and hemostatic networks. Sepsis is associated with evidence of organ dysfunction characterized by tissue hypoperfusion and hypoxia, lactic acidosis, and oliguria (17). Large amounts of NO[•] and ONOO⁻, generated by enhanced expression of the iNOS, as detected in human vessels from septic patients (302), are implicated in the observed symptoms (Fig. 13). Although it is reasonable to imagine that inhibition of NO[•] synthesis could represent a potential therapeutic approach in sepsis, NO[•] inhibitors cause a progressive decrease in cardiac output, amplify organ dysfunction, and increase mortality, probably because of the lack of specificity against the NOS isoform inhibited (177, 250). Independent of the direct effects of NO[•] on GC and the subsequent exaggerated vasorelaxation accounting for hypotension in sepsis, it has been shown that accumulation of S-nitrosothiols in GSNO reductase^{-/-} mice is linked to tissue damage and mortality after endotoxemia (184). These findings indicate that GSNO reductase protects mice from nitrosative stress leading to excessive vasorelaxation and tissue injury after endotoxemia and suggest a central role of S-nitrosothiols in NO[•] biology and disease. Tyrosine nitration of iNOS by ONOO⁻ has been described in rectus abdominis muscle of septic patients (169), suggesting a modulation of activity of this enzyme by RNS. Furthermore, albumin is able to decrease nitrosative stress in a mouse endotoxemia shock model. A recent study shows that albumin prevented vascular hyporeactivity as well as endothelial dysfunction induced by endotoxin in a mouse endotoxic model (204). The effects of albumin are associated with a decreased upregulation of NF- κ B, iNOS, and ONOO⁻ in the vascular wall. In addition, albumin prevents the increases in both NO[•] and O₂^{-•}. Although this study in the mouse endotoxic model does not provide evidence of the mechanism(s) involved in the effects of albumin, it suggests that albumin displays a protective effect in experimental endotoxic shock by reducing the inflammatory process leading to

oxidative and nitrosative stresses. Conversely, evidence suggests that RNS may play a role in sepsis; in mice, deletion of myosin light-chain kinase-210 (MLCK210) isoform, which plays a central role in the control of endothelial barrier function, decreases susceptibility to endotoxin-induced mortality, and this is associated with a decrease in nitrosative stress, as demonstrated by reduction in iNOS expression and in NO[•] production in MLCK210-null mice (263). These findings may help to design new therapeutic strategies for the treatment of septic shock, based on pharmacologic inhibition of MLCK210.

c. Immune responses. Nitrosative stress is associated with the innate immune response toward infection, in which NO[•] is produced at high levels within phagocytes to assure the death of internalized bacteria, virus, and parasites (311). Several studies have shown that rat macrophages, but not mast cells, are able to produce and release NO[•] under pro-inflammatory stimulation (71, 310). In addition, NO[•], as well as hydrogen peroxide, from macrophages inhibits mast cell degranulation mediated by immunoglobulin E (71, 310).

RNS possess antimicrobial actions through several mechanisms. NO[•] and GSNO inhibit the growth of bacteria by inducing double-stranded chromosomal breaks *via* DNA-binding zinc metalloproteins (283). NO[•] donors and ONOO⁻ have been described as powerful inhibitors of hantavirus and virion replication, respectively (153). Inhibition of bacterial respiratory enzymes by NO[•] has been reported (319). Although all these results suggest that RNS can act as antimicrobial agents, it is interesting to note that microorganisms have at least partially developed mechanisms to remove or neutralize RNS, generating resistance against these compounds (84).

4. Cancer. RNS can induce DNA damage and structural and functional modifications of proteins, and thereby may play a role in carcinogenesis. Tumor cells express all isoforms of NOS, depending on the tumor type and progression, and in this way, the effect of NO[•] on tumor progression is determined by the source of cells and the type of NOS (94). Incubation of calf thymus DNA with ONOO⁻ caused DNA cleavage (135). In addition, RNS modify proteins that are essential to cell function, including enzymes involved in cell cycle, apoptosis, and DNA repair (for example, DNA ligase *in vitro*) (106). A positive correlation between iNOS activity and p53 mutations has been observed in human colorectal cancer (7) and in cancer-prone chronic inflammatory disease (132), suggesting that RNS cause mutations of cancer-related genes. In addition, studies performed by using iNOS^{-/-} mice suggested the involvement of NO[•] generated from iNOS in the initiation or promotion of tumorigenesis in melanoma or both (126). In contrast, NO[•] from iNOS inhibits transformation and tumor phenotype of mouse epidermal cells (72). These differences might be related to the different activity and localization of NOS isoforms, concentration and duration of NO[•] exposure, and cellular sensitivity to NO[•] (94). Nevertheless, NO[•] has been shown to enhance tumor cell migration and invasion *via* a cGMP-dependent pathway and to increase tumor cell proliferation in a mice mammary model and in a human colon adenocarcinoma cell line DLD-1 (137, 139), all these processes being linked to angiogenesis. Angiogenesis is a process necessary for the formation of new

vessels able to deliver oxygen and nutrients to tumor cells and to favor its growth. NO[•] from iNOS has been implicated in the angiogenic role of NO[•] through the ability of iNOS to regulate VEGF expression (296) (Fig. 14). Wang's group (339) described that, depending on the expression level of iNOS in murine pancreatic adenocarcinoma cells *in vitro*, the released NO[•] could have a metastatic potential and participate in tumor growth. These effects of NO[•] could be explained by the fact that, *in vitro*, NO[•] stimulates proliferation and migration of endothelial cells by S-nitrosylation of protein tyrosine phosphatases and reduction of their activity (34) or by activation of GC (259). Furthermore, NO[•] is not only a modulator of the expression of endogenous angiogenic factors, but also mediates their functions (79), suggesting that the regulation of the production and release of NO[•] might be a novel approach to overcoming tumor cell resistance to conventional therapeutics. Among the suggestions, strategies to increase or to decrease NO[•] pathway have been described. Thus, in rat medullary thyroid cancer, modification of *iNOS* gene delivery has been tested by using *iNOS*-expressing vectors and, in this model, a volume reduction of tumors has been observed (301). Generation of RNS at high concentrations by using NO[•] donors is directly cytotoxic to rat and human cultured glioma cells or mouse melanoma, and sensitizes to radiation and to some chemotherapeutic agents (*e.g.*, cisplatin) (163, 248). In other cases, inhibition of the NO[•] pathway is an antiangiogenic therapeutic strategy and, as described in murine plasmacytoma (324), the selective inhibition of NO[•] production from iNOS inhibits tumor growth, probably by reducing blood flow and creating a hypoxic environment that reduces the supply of nutrients and oxygen into tumors (123).

Taken together, these data suggest that NO[•] may play a key role in the regulation of cell tumor growth and progression; thus, manipulation of the NO[•] pathway in cancer could represent the potential novel opportunities in the treatment of this disease, at least for some types of cancer.

5. Metabolic diseases. Among the metabolic diseases in which the NO[•] pathway is implicated, diabetes is one of the

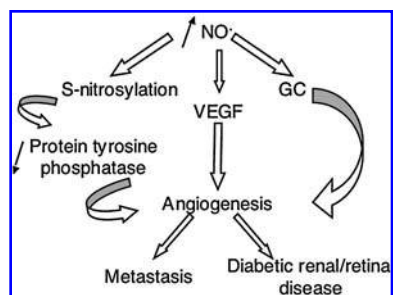


FIG. 14. RNS and angiogenesis. RNS can modulate angiogenesis through several pathways. First, NO[•] favors angiogenesis *via* the vascular endothelial growth factor (VEGF). Second, reduction of the activity of protein tyrosine phosphatase by S-nitrosylation is proangiogenic. Finally, NO[•] can promote angiogenesis through the activation of guanylyl cyclase (GC). Taken together, RNS may play a significant role in the development of metastasis and in certain symptoms of diabetic diseases associated with alteration of the angiogenesis process, such as retinopathy or nephropathy.

most studied. Diabetes-associated vascular complications are the most common major clinical problem that patients with diabetes experience, contributing to the significant morbidity and mortality rate. Thus, patients with diabetes have an elevated incidence of macrovascular complications, such as atherosclerosis, which increase the risk for myocardial infarction, stroke, and peripheral artery disease (often leading to limb amputation), and microvascular complications such as retinopathy and nephropathy, which cause blindness and renal failure. In addition, it is essential to differentiate between type 1 and type 2 diabetes. The former, also called insulin-dependent, is characterized by autoimmune destruction of pancreatic β cells producing insulin; the latter, also called insulin-independent diabetes, is often complicated by other cardiovascular risk factors (345).

Under physiologic conditions, insulin stimulates the production of NO[•] from the endothelium, leading to regulation of vascular tonus (115). However, high levels of insulin elicit an excessive generation of ONOO⁻, which triggers an impairment of endothelium-dependent relaxation *via* a decrease in SERCA function, as shown in diabetic rats (157). Endothelial dysfunction is a consistent feature of insulin-resistant states, including diabetes, obesity, and the metabolic syndrome (for review, see ref. 218). Signaling pathways involved in the coupling between insulin and the insulin receptor display similarities with the cascade regulating NO[•] production. Thus, both insulin resistance and endothelial dysfunction are related to inhibition of the PI3-kinase pathway and overstimulation of the MAP kinase pathway. Recent studies have implicated dysfunctional eNOS as a common pathogenic pathway in diabetic vascular complications (12, 213). Thus, as described earlier, almost all insulin-activated pathways (Akt/PKB, insulin-receptor substrate-1 or p21ras) can be altered by S-nitrosylation, S-glutathionylation, or nitration by RNS (57, 230, 354) (Fig. 15). Insulin secretion and receptor activation and the downstream intracellular transduction signaling can be potentially affected by NO[•] or NO[•] donors, suggesting that RNS could be regulators of the progression of this pathology, in particular of cardiovascular events associated with diabetes. Nevertheless, it should be noted that the effects of RNS in diabetes may be dependent on their source; for example, in contrast to iNOS, gene disruption of eNOS or nNOS in mice leads to insulin resistance (292).

Generation of O₂^{•-} *via* the uncoupling of eNOS has been reported in blood vessels of streptozotocin-induced diabetic rats (122). Recent data obtained in diabetic mice suggest that angiotensin II could be implicated in this process, because angiotensin II-targeted therapy improves endothelial function in diabetic mice *via* the recoupling of eNOS (231).

In bovine microvascular retinal endothelial cells, elevated glucose decreases eNOS expression and activity, as reflected by a significant dose-dependent decrease in nitrate levels in conditioned medium, as well as both laminar shear stress and pulsatile flow-induced activity (59). These effects may account for endothelial dysfunction and impaired autoregulation in diabetic retinopathy. By using a knockout mouse model, Camici and colleagues (39) showed that deletion of the p66^{Shc} adaptor protein prevents not only hyperglycemia-induced endothelial dysfunction but also the generation of ONOO⁻ and tyrosine nitration. These results suggest that p66^{Shc} adaptor protein could be part of a signal-transduction

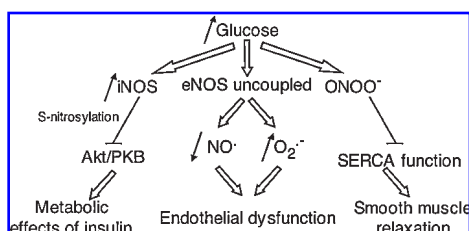


FIG. 15. Effects of RNS on insulin signaling. Elevated glycemia increases iNOS expression and activity, as well as ONOO⁻ production. iNOS inhibits the Akt/PKB pathway, contributing to reduction of metabolic effects of insulin. In parallel, glucose decreases NO[•] production and increases O₂^{-•} release through the uncoupling of eNOS, leading to the endothelial dysfunction observed in diabetes. Finally, SERCA is nitrated by ONOO⁻, leading to an impairment of endothelium-dependent relaxation. Arrows without heads, inhibition.

pathway relevant to hyperglycemia vascular damage and may represent a potential novel therapeutic target against diabetic vascular complications.

Furthermore, the NO[•]/cGMP pathway has been described as participating in the contractility of cardiomyocytes in streptozotocin-diabetic rats (33, 300). An increased production of NO[•], probably by both iNOS and constitutive NOS, and the subsequent increased production of cGMP within the myocytes from streptozotocin-diabetic rats suppressed the shortening response to β -adrenergic stimulation without affecting the Ca²⁺ current through L-type Ca²⁺ channels (300). Moreover, small coronary arteries from diabetic rats exhibit enhanced ONOO⁻ formation and concurrent impairment of voltage-dependent K⁺ channel function by direct nitration of the channels (33).

Another aspect of diabetes is the neuropathy that affects at least 30–50% of diabetic patients, independent of the type of the diabetes. Recent data show that nitrosative stress plays an important role in early experimental neuropathy in streptozotocin-induced diabetic rats (232), by inducing motor and sensory nerve-conduction velocity deficits, associated with increased nitrotyrosine in the sciatic nerve and dorsal root ganglia. The same group reported that, in leptin-deficient *ob/ob* mice, a model with type 2 diabetes with mild hyperglycemia and obesity, ONOO⁻ decomposition catalysts restore motor and sensory nerve-conduction velocity, improving nerve dysfunction induced by ONOO⁻, underlining the deleterious effects of RNS in neuropathy in type 2 diabetes, at least in mice (330).

Nephropathy also is implicated in morbidity and mortality in patients with diabetes. Kanetsuna and colleagues (146) proposed a vital role for eNOS-derived NO[•] in the prevention of diabetic nephropathy in mice. By using diabetic eNOS-knockout mice, these authors have demonstrated that deletion of eNOS leads to the development of albuminuria, hypertension, and glomerular mesangiolysis, linked to glomerular hyperfiltration and injured endothelial morphology. This has been corroborated by another group, showing that diabetic eNOS knockout mice developed nephropathy, which is associated with increased endothelial proliferation and VEGF expression. In addition, all these effects are improved with insulin therapy (220). Altogether,

these findings suggest that a relative deficiency in eNOS-derived NO[•] levels may participate in the increased susceptibility to nephropathy in diabetic subjects (146, 221).

Patients with diabetes display impairment of re-endothelialization after vascular injury, associated with a dysfunction in the recruitment of endothelial progenitor cells (EPCs) in the affected zones (134). Recently, it was shown that EPC levels in patients with diabetes are reduced compared with those of control subjects. In addition, EPCs from patients with diabetes are able to produce excessive O₂^{-•}, which is reduced when eNOS is inhibited (318). These authors propose an uncoupling of eNOS in EPCs from patients with diabetes, resulting in O₂^{-•} production. This may be responsible for the impaired neovascularization described in diabetes. Besides, a potentially deleterious role of VEGF in diabetic renal disease has been proposed, because new extra vessel formation in the glomerulus is correlated with increased VEGF expression in biopsies from human diabetic kidney (147) (Fig. 14).

Dermal application of GSNO is able to promote local vasodilatation in streptozotocin-induced diabetic rats, without inducing protein nitration or alterations in blood pressure or heart rate, suggesting that GSNO may be an effective treatment that prevents or reverses the consequences of endothelial dysfunction in patients with diabetes (289).

VI. Future Directions

During the 1980s, RNS were considered to be molecules with complex properties, at least within the cardiovascular system. However, in the last decade, the numbers of identified RNS and of potential targets of RNS have proportionately increased. In addition, RNS possess different effects (beneficial or deleterious) in relation to their cellular origin, the enzymatic source, their concentration and, finally, the cell target. The use of NO[•] or NO[•] donors (sodium nitroprusside or nitroglycerine, for example) as pulmonary or coronary vasodilators in different pathologies is now routine (317). These effects are based on the NO[•] signaling with cGMP as second messenger. In this case, modulation of expression or activity or both of eNOS, as well as modification of cGMP degradation by phosphodiesterases, remains the main axis of therapy. However, as shown in this review, much information about other intracellular transduction signaling pathways that mediate NO[•] effects is now available, and the modulation of these “new” pathways might open novel potential therapeutic strategies by neutralization of RNS, or inhibition or activation of RNS generation, depending on the pathologic situation.

Different molecules acting on a variety of targets are able to reduce RNS-induced cellular dysfunction or to inhibit enzymes on which RNS induce an excess of function. Many of the examples reported in the other sections of this review illustrate this point. As an example, acting on the angiotensin-converting enzyme by using enalapril (an inhibitor of this enzyme) reduces dysfunction of vascular smooth muscle SERCA and enhancement of tyrosine nitration in diabetic rats (315). It is plausible to consider that, in humans with diabetes, inhibition of the angiotensin II pathway could ameliorate cardiovascular complications.

Beneficial effects of PPAR agonists in the human insulin-resistance syndrome are well documented (for a review, see

ref. 302). PPARs act as central transcriptional mediators of metabolic processes. Thus, PPAR agonists are used as insulin sensitizers for their ability to correct the abnormal blood flow in patients with diabetes. Although PPAR α and PPAR γ agonists can contribute to amelioration of the macrovascular dysfunctions, the combinations of PPAR α , PPAR β , and PPAR γ may better serve to improve the microvascular defect in diabetes (335). PPAR agonists could act by reducing NO \cdot production from iNOS and the subsequent nitrosative stress (336). It remains to be determined whether the reduction of nitrosative stress by PPAR agonists correlates with an improvement of neurovascular function in the long term in diabetes.

Not many therapies exist for the microvascular complications observed in patients with diabetes. In particular, improvement of the impaired angiogenesis and vasculogenesis could represent a potential field for individuals with diabetes. Gallagher's group (95) recently showed that hyperbaric oxygen therapy, by increasing NOS-derived NO \cdot levels in perivascular tissues, enhances EPC mobilization from bone marrow in streptozotocin-treated mice. In addition, stromal cell-derived factor-1 α (SDF-1 α) mediates EPC homing, and both treatments have synergistic effects. Although the clinical relevance of this study remains to be established, it could represent a new hope for wound healing in diabetes.

As discussed earlier, S-nitrosylation of NMDA receptor could have a protective effect in neurodegenerative diseases by decreasing NMDA activity. New combinatorial drugs, nitromemantines, second-generation memantine antagonists of NMDA receptors, plus a nitro group, are able to target NO \cdot to the S-nitrosylation sites of the NMDA receptors. Preliminary studies performed by the Lipton's group have shown that nitromemantines are more effective than memantine as neuroprotectants, both *in vitro* and *in vivo* in animal models, without affecting blood pressure (for a review, see ref. 223). With these molecules, a promising research field is now opened: the modulation of S-nitrosylation sites.

In diseases such as cancer, both inhibition and overproduction of NO \cdot have been proposed as strategies for therapy. Whereas the administration of NOS inhibitors, such as N G -nitro-L-arginine methyl ester (L-NAME), can slow the growth of tumors in animal models (322), NO \cdot donors can act directly on cells to inhibit their growth (163, 248). In this way, dietary components targeting the iNOS-VEGF axis could be used as a practical approach for cancer prevention and intervention (245).

Conversely, the formation of S-nitrosothiols, which are local NO \cdot stores from which biologically active NO \cdot can be released, offers a novel and promising field in several pathologies in which NO \cdot levels can be compromised (217).

In conclusion, although the NO \cdot pathway continues to be an important target for treatment of different pathologies, application to humans of data obtained from cell or animal models requires more preclinical and clinical studies to confirm the effects of RNS in human diseases. In addition, RNS have both positive and negative effects that contribute to the physiology and pathophysiology of many disorders. Thus, measurements of new biomarkers of nitrosative stress could help us to understand the role of RNS in the regulation of homeostasis.

Acknowledgments

We thank Dr. Helen Langemann for critical reading and English corrections, and Dr. Ismail Laher, for excellent comments and suggestions.

This work was supported by institutional grants from CNRS, INSERM, and Université d'Angers. R.A. was supported by Contrat d'Interface Inserm.

Abbreviations

3-NO $_2$ -Tyr, 3-nitrotyrosine; 8-NO $_2$ -G, 8-nitroguanine; 8-NO $_2$ -Guo, 8-nitroguanosine; ALS, amyotrophic lateral sclerosis; BH $_4$, tetrahydrobiopterin; cADPR, cyclic ADP ribose; CFTR, cystic fibrosis transmembrane conductance regulator; cGMP, cyclic guanosine 3',5'-monophosphate; COX, cyclooxygenase; CuZn-SOD, copper-zinc superoxide dismutase; EC-SOD, extracellular superoxide dismutase; EGF, epidermal growth factor; eNOS, endothelial nitric oxide synthase; EPCs, endothelial progenitor cells; FGF-1, fibroblast growth factor; GC, guanylyl cyclase; GSH, glutathione; GSNO, S-nitrosoglutathione; IGF, insulin-like growth factor-1; IKK β , inhibitory κ B kinase β ; iNOS, inducible nitric oxide synthase; InsP $_3$, inositol 1,4,5-triphosphate; IRAG, inositol-1,4,5-trisphosphate receptor-associated cGMP kinase substrate; JNK, c-Jun N-terminal kinase; LDL, low-density lipoprotein; LNO $_2$, nitrolinoleic acid; MAP kinase, mitogen-activated protein kinase; MLCK210, myosin light-chain kinase-210; MMPs, matrix metalloproteinases; Mn-SOD, manganese superoxide dismutase; NF- κ B, nuclear factor- κ B; NGF, nerve growth factor; NMDA, N-methyl-D-aspartate receptors; nNOS, neuronal nitric oxide synthase; NO $^-$, nitroxyl; NO \cdot , nitric oxide; NO $^+$, nitrosonium; NOS, nitric oxide synthase; NSF, N-ethylmaleimide sensitive factor; O $_2$ $^-$, superoxide; OA-NO $_2$, oleic acid; ODQ, 1H-[1,2,4]oxadiazolo [4,3-a]quinoxalin-1-one; ONOO $^-$, peroxyntirite; PARP, poly ADP-ribose polymerase; PETN, phosphatase and tensin homologue; PKG, protein kinase G; PPARs, peroxisome proliferators-activated receptors; RGS2, regulator of G protein signaling; RNS, reactive nitrogen species; ROS, reactive oxygen species; RSNO, S-nitrosothiol derivative; RSNOs, S-nitrosothiols; RyR, ryanodine receptor; SDF-1 α , stromal cell-derived factor-1 α ; SERCA, sarco/endoplasmic calcium pumps; SMase, sphingomyelinase; SOD, superoxide dismutase; TRP, transient receptor potential channel; VASP, vasodilator-stimulated phosphoprotein; VEGF, vascular endothelial growth factor; VGCCs, voltage-dependent calcium channels.

Disclosure Statement

No competing financial interests exist.

References

1. Abi-Gerges N, Fischmeister R, and Méry PF. G protein-mediated inhibitory effect of a nitric oxide donor on the L-type Ca $^{2+}$ current in rat ventricular myocytes. *J Physiol* 531: 117–130, 2001.
2. Adachi T, Matsui R, Xu S, Kirber M, Lazar HL, Sharov VS, Schoneich C, and Cohen RA. Antioxidant improves smooth muscle sarco/endoplasmic reticulum Ca $^{2+}$ -ATP function and lowers tyrosine nitration in hypercholesterolemia and improves nitric oxide-induced relaxation. *Circ Res* 90: 1114–1121, 2002.

3. Adachi T, Weibrod RM, Pimentel DR, Ying J, Sharov VS, Schoneich C, and Cohen RA. S-Glutathiolation by peroxynitrite activates SERCA during arterial relaxation by nitric oxide: a mechanism targeted by oxidants in vascular disease. *Nat Med* 10: 1200–1207, 2004.
4. Ahern GP, Hsu SF, and Jackson MB. Direct actions of nitric oxide on rat neurohypophyseal K⁺ channels. *J Physiol* 520: 165–176, 1999.
5. Akaike T, Okamoto S, Sawa T, Yoshitake J, Tamura F, Ichimoro K, Miyazaki K, Sasamoto K, and Maeda H. 8-Nitroguanosine formation in viral pneumonia and its implication for pathogenesis. *Proc Natl Acad Sci U S A* 100: 685–690, 2003.
6. Alp NJ and Channon KM. Regulation of endothelial nitric oxide synthase by tetrahydrobiopterin in vascular disease. *Arterioscler Thromb Vasc Biol* 24: 413–420, 2004.
7. Ambs S, Bennett WP, Merriam WG, Ogunfusika MO, Oser SM, Harrington AM, Shields PG, Felly-Bosco E, Hussain SP, and Harris CC. Relationship between p53 mutations and inducible nitric oxide synthase expression in human colorectal cancer. *J Natl Cancer Inst* 91: 86–88, 1999.
8. Andersson E, Schain F, Svedling M, Claesson HE, and Forsell PK. Interaction of human 15-lipoxygenase-1 with phosphatidylinositol biphosphates results in increased enzyme activity. *Biochim Biophys Acta* 1761: 1498–1505, 2006.
9. Aoyama K, Matsubara K, Fujikawa Y, Nagahiro Y, Shimizu K, Umegae N, Hayase N, Shiono H, and Kobayashi S. Nitration of manganese superoxide dismutase in cerebrospinal fluids is a marker for peroxynitrite-mediated oxidative stress in neurodegenerative diseases. *Ann Neurol* 47: 524–527, 2000.
10. Aslan M, Ryan TM, Townes TM, Coward L, Kirk MC, Barnes S, Alexander CB, Rosenfeld SS, and Freeman BA. Nitric oxide-dependent generation of reactive species in sickle cell disease: actin tyrosine nitration induces defective cytoskeletal polymerization. *J Biol Chem* 278: 4194–4204, 2003.
11. Atar S, Ye Y, Lin Y, Freeberg SY, Nishi SP, Rosanio S, Huang MH, Uretsky BF, Perez-Polo JR, and Birnbaum Y. Atorvastatin-induced cardioprotection is mediated by increasing inducible nitric oxide synthase and consequent S-nitrosylation of cyclooxygenase-2. *Am J Physiol Heart Circ Physiol* 290: H1960–H1968, 2006.
12. Awata T, Neda T, Iizuka H, Kurihara S, Ohkubo T, Takata N, Osaki M, Watanabe M, Nakashima Y, Sawa T, Inukai K, Inoue I, Shibuya M, Mori K, Yoneya S, and Katayama S. Endothelial nitric oxide synthase gene is associated with diabetic macular edema in type 2 diabetes. *Diabetes Care* 27: 2184–2190, 2004.
13. Azad N, Vallyathan V, Wang L, Tantishyakul V, Stehlik C, Leonard SS, and Rojanasakul Y. S-Nitrosylation of Bcl-2 inhibits its ubiquitin-proteasomal degradation. *J Biol Chem* 281: 34124–34134, 2006.
14. Bagnasco P, MacMillan-Crow LA, Greendorfer JS, Young CJ, Andrews L, and Thompson JA. Peroxynitrite modulates acidic fibroblast growth factor (FGF-1) activity. *Arch Biochem Biophys* 419: 178–189, 2003.
15. Baker PR, Schopfer FJ, Sweeney FJ, and Freeman BA. Red cell membrane and plasma linoleic acid nitration products: synthesis, clinical identification, and quantitation. *Proc Natl Acad Sci U S A* 101: 11577–11582, 2004.
16. Baker LM, Baker PR, Golin-Bisello F, Schopfer FJ, Fink M, Woodcock SR, Branchaud BP, Radi R, and Freeman BA. Nitro-fatty acid reaction with glutathione and cysteine: kinetic analysis of thiol alkylation by a Michel addition reaction. *J Biol Chem* 282: 31085–31093, 2007.
17. Balk RA. Severe sepsis and septic shock. *Crit Care Clin* 16: 179–192, 2000.
18. Banan A, Zhang LJ, Shaikh M, Fields JZ, Farhadi A, and Keshavazian A. Novel effect of NF-kappaB activation: carbonylation and nitration injury to cytoskeleton and disruption of monolayer barrier in intestinal epithelium. *Am J Physiol Cell Physiol* 287: C1139–C1151, 2004.
19. Barbeito LH, Pehar M, Cassina P, Vargas MR, Peluffo H, Viera L, Estevez AG, and Beckman JS. A role for astrocytes in motor neurons loss in amyotrophic lateral sclerosis. *Brain Res Brain Res Rev* 47: 263–274, 2004.
20. Barouch LA, Harrison RW, Skaf MW, Rosas GO, Cappola TP, Kobeissi ZA, Hobai IA, Lemmon CA, Burnett AL, O'Rourke B, Rodriguez ER, Huang PL, Lima JA, Berkowitz DE, and Hare JM. Nitric oxide regulates the heart by spatial confinement of nitric oxide synthase isoforms. *Nature* 416: 337–340, 2002.
21. Barrett DM, Black SM, Todor H, Schmidt-Ullrich RK, Dawson KS, and Mikkelsen RB. Inhibition of protein-tyrosine phosphatases by mild oxidative stresses is dependent on S-nitrosylation. *J Biol Chem* 280: 14453–14461, 2005.
22. Barsacchi R, Perrotta C, Sestili P, Cantoni O, Moncada S, and Clementi E. Cyclic GMP-dependent inhibition of acid sphingomyelinase by nitric oxide: an early step in protection against apoptosis. *Cell Death Differ* 9: 1248–1255, 2002.
23. Bebek Z, Varga K, Hicks JK, Venglarik CJ, Kovacs T, Chen L, Hardiman KM, Collawn JF, Sorscher EJ, and Matalon S. Reactive oxygen nitrogen species decrease cystic fibrosis transmembrane conductance regulator expression and cAMP-mediated Cl[−] secretion in airway epithelia. *J Biol Chem* 277: 43041–43049, 2002.
24. Bencsik P, Kupai K, Giricz Z, Görbe A, Huliák I, Fürst S, Dux L, Csont T, Jancsó G, and Ferdinandy P. Cardiac capsaicin-sensitive sensory nerves regulate myocardial relaxation via S-nitrosylation of SERCA: role of peroxynitrite. *Br J Pharmacol* 153: 488–496, 2008.
25. Berry CE and Hare JM. Xanthine oxidoreductase and cardiovascular disease: molecular mechanisms and pathophysiological implications. *J Physiol* 555: 589–606, 2004.
26. Bielefeldt K, Whiteis CA, Chapleau MW, and Abboud FM. Nitric oxide enhances slow inactivation of voltage-dependent sodium currents in rat nodose neurons. *Neurosci Lett* 271: 159–162, 1999.
27. Birnbaum SG, Varga AW, Yuan LL, Anderson AE, Sweatt JD, and Schrader LA. Structure and function of Kv4-family transient potassium channels. *Physiol Rev* 84: 803–833, 2004.
28. Bolotina VM, Najibi S, Palacino JJ, Pagano PJ, and Cohen RA. Nitric oxide directly activates calcium-dependent potassium channels in vascular smooth muscle. *Nature* 368: 850–853, 1994.
29. Bonnevier J, Fässler R, Somlyo AP, Somlyo AV, and Arner A. Modulation of Ca²⁺[ops] sensitivity by cyclic nucleotides in smooth muscle from protein kinase G-deficient mice. *J Biol Chem* 279: 5146–5151, 2004.
30. Boulous C, Jiang H, and Balazy M. Diffusion of peroxynitrite into the human platelet inhibits cyclooxygenase via nitration of tyrosine residues. *J Pharmacol Exp Ther* 293: 222–229, 2000.
31. Brigelius-Flohé R. Glutathione peroxidases and redox-regulated transcription factors. *Biol Chem* 387: 1329–1335, 2006.
32. Bryan NS, Rassaf T, Maloney RE, Rodriguez CM, Saijo F, Rodriguez JR, and Feelisch M. Cellular targets and mechanism of nitrosylation: an insight into their nature and

- kinetics in vivo. *Proc Natl Acad Sci U S A* 101: 4308–4313, 2004.
33. Bubolz AH, Wu Q, Larsen BT, Gutterman DD, and Liu Y. Ebselen reduces nitration and restores voltage-gated potassium channel function in small coronary arteries of diabetic rats. *Am J Physiol Heart Circ Physiol* 293: H2231–H2237, 2007.
 34. Bunderson M, Coffin JD, and Beall HD. Arsenic induces peroxynitrite generation and cyclooxygenase-2 protein expression in aortic endothelial cells: possible role in atherosclerosis. *Toxicol Appl Pharmacol* 184: 11–18, 2002.
 35. Busse R and Fleming I. Vascular endothelium and blood flow. *Handb Exp Pharmacol* 176: 43–78, 2006.
 36. Cai J, Jiang WG, Ahmed A, and Boulton M. Vascular endothelial growth factor-induced endothelial cell proliferation is regulated by interaction between VEGFR-2, SH-PTP1 and eNOS. *Microvasc Res* 71: 20–31, 2006.
 37. Calabrese V, Sultana R, Scapagnini G, Guagliano E, Sapienza M, Bella R, Kanski J, Pennisi G, Mancuso C, Stella AMG, and Butterfield DA. Nitrosative stress, cellular stress response, and thiol homeostasis in patients with Alzheimer's disease. *Antioxid Redox Signal* 8: 1975–1986, 2006.
 38. Calò L, Giacon B, Davis PA, Pagnin E, Piccin A, Riegler P, Huber W, Antonello A, and Semplicini A. Oxidative stress and TGF β in kidney-transplanted patients with cyclosporin-induced hypertension: effect of carvedilol and nifedipine. *Clin Nephrol* 58: 103–110, 2002.
 39. Camici GG, Schiavoni M, Francia P, Bachschmid M, Martin-Padura I, Hersberger M, Tanner FC, Pelicci PG, Volpe M, Anversa P, Lüscher TF, and Cosentino F. Genetic deletion of p66^{Shc} adaptor protein prevents hyperglycemia-induced endothelial dysfunction and oxidative stress. *Proc Natl Acad Sci U S A* 104: 5217–5222, 2007.
 40. Cannon RO 3rd, Schechter AN, Panza JA, Ognibene FP, Pease-Fye ME, Wacławski MA, Shelhamer JH, and Gladwin MT. Effects of inhaled nitric oxide on regional blood flow are consistent with intravascular nitric oxide delivery. *J Clin Invest* 108: 279–287, 2001.
 41. Cappelletti G, Maggioni MG, Ronchi C, Maci R, and Tedeschi G. Protein tyrosine nitration is associated with cold- and drug-resistant microtubules in neuronal-like PC12 cells. *Neurosci Lett* 401: 159–164, 2006.
 42. Carreras MC, Franco MC, Peralta JG, and Poderoso JJ. Nitric oxide, complex I, and the modulation of mitochondrial reactive species in biology and disease. *Mol Aspects Med* 25: 125–139, 2004.
 43. Carver DJ, Gaston B, deRonde K, and Palmer LA. Akt-mediated activation of HIF-1 in pulmonary vascular endothelial cells by S-nitrosoglutathione. *Am J Respir Cell Mol Biol* 37: 255–263, 2007.
 44. Casadei B. The merging role of neuronal nitric oxide synthase in the regulation of myocardial function. *Exp Physiol* 91: 943–955, 2006.
 45. Casoni F, Basso M, Massignan T, Gianazza E, Cheroni C, Salmons M, Bendotti C, and Bonetto V. Protein nitration in a mouse model of familial myotrophic lateral sclerosis: possible multifunctional role in the pathogenesis. *J Biol Chem* 280: 16295–16304, 2005.
 46. Castillo SS, Levy M, Thaikootathil JV, and Goldkorn T. Reactive nitrogen and oxygen species activate different sphingomyelinases to induce apoptosis in airway epithelial cells. *Exp Cell Res* 313: 2680–2686, 2007.
 47. Castillo SS, Levy M, Wang C, Thaikootathil JV, Khan E, and Goldkorn T. Nitric oxide-enhanced caspase-3 and acidic sphingomyelinase interaction: a novel mechanism by which airway epithelial cells escape ceramide-induced apoptosis. *Exp Cell Res* 313: 816–823, 2007.
 48. Chen C and Schofield GG. Nitric oxide donors enhanced Ca²⁺ currents and blocked noradrenaline-induced Ca²⁺ current inhibition in rat sympathetic neurons. *J Physiol* 482: 521–531, 1995.
 49. Chen L, Patel RP, Teng X, Bosworth CA, Lancaster J Jr, and Matalon S. Mechanisms of cystic fibrosis transmembrane conductance regulator activation by S-nitrosoglutathione. *J Biol Chem* 281: 9190–9199, 2006.
 50. Chen ZH, Yoshida Y, Saito Y, Noguchi N, and Niki E. Adaptive response induced by lipid peroxidation products in cell cultures. *FEBS Lett* 580: 479–483, 2006.
 51. Chi Q, Wang T, and Huang K. Effect of insulin nitration by peroxynitrite on its biological activity. *Biochem Biophys Res Commun* 330: 791–796, 2005.
 52. Choi YB, Tenneti L, Le DA, Ortiz J, Bai G, Chen HSV, and Lipton SA. Molecular basis of NMDA receptor-coupled ion channel modulation by S-nitrosylation. *Nat Neurosci* 3: 15–21, 2000.
 53. Chou SM, Wang HS, and Komai K. Colocalization of NOS and SOD1 in neurofilament accumulation within motor neurons of amyotrophic lateral sclerosis: an immunohistochemical study. *J Chem Neuroanat* 10: 249–258, 1996.
 54. Chung SS, Ahn DS, Lee HG, Lee YH, and Nam TS. Inhibition of carbachol-evoked oscillatory currents by the NO donor sodium nitroprusside in guinea-pig ileal myocytes. *Exp Physiol* 90: 577–586, 2005.
 55. Chung KKK, Thomas B, Li X, Pletnikova O, Troncoso JC, Marsh L, Dawson VL, and Dawson TM. S-Nitrosylation of parkin regulates ubiquitination and compromises parkin's protective function. *Science* 304: 1328–1331, 2004.
 56. Clancy R, Varenika B, Huang W, Ballou L, Attur M, Amin AR, and Abramson SB. Nitric oxide synthase/COX cross-talk: nitric oxide activates COX-1 but inhibits COX-2-derived prostaglandin production. *J Immunol* 165: 1582–1587, 2000.
 57. Clavreul N, Bachschmid MM, Hou X, Shi C, Idrizovic A, Ido Y, Pimentel D, and Cohen RA. S-glutathiolation of p21ras by peroxynitrite mediates endothelial insulin resistance caused by oxidized low-density lipoprotein. *Arterioscler Thromb Vasc Biol* 26: 2454–2461, 2006.
 58. Clements MK, Siemsen DW, Swain SD, Hanson AJ, Nelson-Overton LK, Rohn TT, and Quinn MT. Inhibition of action polymerization by peroxynitrite modulates neutrophil functional responses. *J Leukoc Biol* 73: 344–355, 2003.
 59. Connell P, Walshe T, Ferguson G, Gao W, O'Brien C, and Caahill PA. Elevated glucose attenuates agonist- and flow-stimulated endothelial nitric oxide synthase activity in microvascular retinal endothelial cells. *Endothelium* 14: 17–24, 2007.
 60. Crawford JH, Chacko BK, and Patel RP. Regulation of vascular function by haemoglobin. *Biochem Soc Symp* 71: 135–142, 2004.
 61. Cuccurullo C, Fazia ML, Mezzetti A, and Cipollone F. COX-2 expression in atherosclerosis: the good, the bad or the ugly? *Curr Med Chem* 14: 1595–1605, 2007.
 62. Cul T, Schopfer FJ, Zhang J, Chen Z, Ichikawa T, Baker PRS, Batthyany C, Chacko BK, Feng X, Patel RP, Agarwall A, Freeman BA, and Chen YE. Nitrated fatty acids: endogenous anti-inflammatory signaling mediators. *J Biol Chem* 281: 35686–35698, 2006.
 63. Datta B, Tufnell-Barrett T, Bleasdale RA, Jones CJ, Beeton I, Paul V, Frenneaux M, and James P. Red blood cell nitric oxide as an endocrine vasoregulator: a potential role in congestive heart failure. *Circulation* 109: 1339–1342, 2004.

64. Davis B and Zou MH. CD40 ligand-dependent tyrosine nitration of prostacyclin synthase in vivo. *Circulation* 112: 2184–2192, 2005.
65. Dedkova EN, Wang YG, Blatter LA, and Lipsius SL. Nitric oxide signalling by selective beta(2)-adrenoceptor stimulation prevents ACh-induced inhibition of beta(2)-stimulated Ca(2+) current in cat atrial myocytes. *J Physiol* 542: 711–723, 2002.
66. Dedon PC and Tannenbaum SR. Reactive nitrogen species in the chemical biology of inflammation. *Arch Biochem Biophys* 423: 12–22, 2004.
67. Deeb RS, Shen H, Gamss C, Gavrilova T, Summers BD, Kraemer R, Hao G, Gross SS, Lainé M, Maeda N, Hajjar DP, and Upmacis RK. Inducible nitric oxide synthase mediates prostaglandin H2 synthases nitration and suppresses eicosanoid production. *Am J Pathol* 168: 349–362, 2006.
68. Delgado-Esteban M, Martin-Zanca D, Andres-Martin L, Almeida A, and Bolaños JP. Inhibition of PTEN by peroxynitrite activates the phosphoinositide-3-kinase/akt neuroprotective signaling pathway. *J Neurochem* 102: 194–205, 2007.
69. De Martin R, Hoeth M, Hofer-Warbinek R, and Schmid JA. The transcription factor NF-kappaB and the regulation of vascular cell function. *Arterioscler Thromb Vasc Biol* 20: e83–e88, 2000.
70. Deora AA, Hajjar DP, and Lander HM. Recruitment and activation of Raf-1 kinase by nitric oxide-activated Ras. *Biochemistry* 39: 9901–9908, 2000.
71. DeSchoolmeester ML, Eastmond NC, Dearman RJ, Kimber I, Basketter DA, and Coleman JW. Reciprocal effects of interleukin-4 and interferon-gamma on immunoglobulin E-mediated mast cell degranulation: a role for nitric oxide but not peroxynitrite or cyclic guanosine monophosphate. *Immunology* 96: 138–144, 1999.
72. Dhar A, Brindley JM, Stark C, Citro ML, Keefer LK, and Colburn NH. Nitric oxide, does not mediate but inhibits transformation and tumor phenotype. *Mol Cancer Ther* 2: 1285–1293, 2003.
73. Dickinson DA and Forman HJ. Glutathione in defense and signaling: lessons from a small thiol. *Ann N Y Acad Sci* 973: 488–504, 2002.
74. Diss JK, Fraser SP, and Djamgoz MB. Voltage-gated Na+ channels: multiplicity of expression, plasticity, functional implications and pathophysiological aspects. *Eur Biophys J* 33: 180–193, 2004.
75. Dixon LJ, Morgan DR, Hughes SM, McGrath LT, El-Sherbeeny NA, Plumb RD, Devine A, Leahey W, Johnston GD, and McVeigh GE. Functional consequences of endothelial nitric oxide synthase uncoupling in congestive cardiac failure. *Circulation* 107: 1725–1728, 2003.
76. Doel JJ, Godber BL, Goult TA, Eisenthal R, and Harrison R. Reduction of organic nitrites to nitric oxide catalyzed by xanthine oxidase: possible role in metabolism of nitrovasodilators. *Biochem Biophys Res Commun* 270: 880–885, 2000.
77. Dong M, Vongchampa V, Gingipalli L, Cloutier JF, Kow YW, O'Connor T, and Dedon PC. Development of enzymatic probes of oxidative and nitrosative DNA damage caused by reactive nitrogen species. *Mutat Res* 594: 120–134, 2006.
78. Drews G, Kramer C, and Krippeit-Drews P. Dual effect of NO on K+ATP current of mouse pancreatic B-cells: stimulation by deenergizing mitochondria and inhibition by direct interaction with the channel. *Biochim Biophys Acta* 1464: 62–68, 2000.
79. Dulak J and Jozkowicz A. Regulation of vascular endothelial growth factor synthesis by nitric oxide: facts and controversies. *Antioxid Redox Signal* 5: 123–132, 2003.
80. El-Remessy AB, Bartoli M, Platt D, Fulton D, and Caldwell RB. Oxidative stress inactivates VEGF survival signaling in retinal endothelial cells via PI3-kinase tyrosine nitration. *J Cell Sci* 118: 243–252, 2005.
81. Erwin PA, Lin AJ, Golan DE, and Michel T. Receptor-regulated dynamic S-nitrosylation of endothelial nitric-oxide synthase in vascular endothelial cells. *J Biol Chem* 280: 19888–19894, 2005.
82. Eu JP, Sun J, Xu L, Stamler JS, and Meissner G. The skeletal muscle calcium release channel: coupled O2 sensor and NO signaling functions. *Cell* 102: 499–509, 2000.
83. Eyal A, Szargel R, Avraham E, Liani E, Haskin J, Rott R, and Engelender S. Synphilin-1A: an aggregation-prone isoform of synphilin-1 that causes neuronal death and is present in aggregates from alpha-synucleinopathy patients. *Proc Natl Acad Sci U S A* 103: 5917–5922, 2006.
84. Fang FC. Antimicrobial reactive oxygen and nitrogen species: concepts and controversies. *Nat Rev Microbiol* 2: 820–832, 2004.
85. Farber HW and Loscalzo J. Pulmonary arterial hypertension. *N Engl J Med* 351: 1655–1665, 2004.
86. Favre CJ, Ufret-Vicenty CA, Stone MR, Ma HT, and Gill DL. Ca2+ pool emptying stimulates Ca2+ entry activated by S-nitrosylation. *J Biol Chem* 273: 30855–30858, 1998.
87. Feng Q, Lu X, Jones DL, Shen J, and Arnold JM. Increased inducible nitric oxide synthase expression contributes to myocardial dysfunction and higher mortality after myocardial infarction in mice. *Circulation* 104: 700–704, 2001.
88. Fernandez AP, Serrano J, Rodrigo J, Monleon E, Vargas A, Badiola JJ, Martinez-Murillo R, and Martinez A. Changes in the expression pattern of the nitrergic system of ovine cerebellum affected by scrapie. *J Neuropathol Exp Neurol* 66: 196–207, 2007.
89. Filipovic MR, Stanic D, Raicevic S, Spasic M, and Niketic V. Consequences of MnSOD interactions with nitric oxide: nitric oxide dismutation and the generation of peroxynitrite and hydrogen peroxide. *Free Radic Res* 41: 62–72, 2007.
90. Findeis MA. The role of amyloid beta peptide 42 in Alzheimer's disease. *Pharmacol Ther* 116: 266–286, 2007.
91. Foskett JK, White C, Cheung KH, and Mak DO. Inositol trisphosphate receptor Ca2+ release channels. *Physiol Rev* 87: 593–658, 2007.
92. Foster MW, McMahon TJ, and Stamler JS. S-nitrosylation in health and disease. *Trends Mol Med* 9: 160–168, 2003.
93. Freixes M, Rodriguez A, Dalfo E, and Ferrer I. Oxidation, glycoxidation, lipoxidation, nitration, and responses to oxidative stress in the cerebral cortex in Creutzfeldt-Jacob disease. *Neurobiol Aging* 27: 1807–1815, 2006.
94. Fukumura D, Kashiwagi S, and Jain RK. The role of nitric oxide in tumour progression. *Nat Rev Cancer* 6: 521–534, 2006.
95. Gallagher KA, Liu ZJ, Xiao M, Chen H, Goldstein LJ, Buerk DG, Nedeau A, Thom SR, and Velazquez OC. Diabetic impairments in NO-mediated endothelial progenitor cell mobilization and homing are reversed by hyperoxia and SDF-1α. *J Clin Invest* 117: 1249–1259, 2007.
96. Gaston B, Sears S, Woods J, Hunt J, Ponaman M, McMahon T, and Stamler JS. Bronchodilator S-nitrosothiol deficiency in asthmatic respiratory failure. *Lancet* 351: 1317–1319, 1998.
97. Ghosh S, Janocha AJ, Aronica MA, Swaidani S, Comhair SA, Xu W, Zheng L, Kaveti S, Kinter M, Hazen SL, and

- Erzuzum SC. Nitrotyrosine proteome survey in asthma identifies oxidative mechanism of catalase inactivation. *J Immunol* 176: 5587–5597, 2006.
98. Giasson BI, Duda JE, Murray IVJ, Chen Q, Souza JM, Hurtig HI, Ischiropoulos H, Trojanowski JQ, and Lee VM-Y. Oxidative damage linked to neurodegeneration by selective α -synuclein nitration in synucleinopathy lesions. *Science* 290: 985–989, 2000.
 99. Girouard H, Park L, Anrather J, Zhou P, and Iadecola C. Cerebrovascular nitrosative stress mediates neurovascular and endothelial dysfunction induced by angiotensin II. *Arterioscler Thromb Vasc Biol* 27: 303–309, 2007.
 100. Gladwin MT, Crawford JH, and Patel RP. The biochemistry of nitric oxide, nitrite, and hemoglobin: role in blood flow regulation. *Free Radic Biol Med* 36: 707–717, 2004.
 101. Gladwin MT, Raat NJ, Shiva S, Dezfulian C, Hogg N, Kim-Shapiro DB, and Patel RP. Nitrite as a vascular endocrine nitric oxide reservoir that contributes to hypoxic signaling, cytoprotection, and vasodilation. *Am J Physiol Heart Circ Physiol* 291: H2026–H2035, 2006.
 102. Gladwin MT and Patel RP. The role of red blood cells and hemoglobin-nitric oxide interactions on blood flow. *Am J Respir Cell Mol Biol* 38: 125–126, 2008.
 103. Gonzalez D, Drapier JC, and Bouton C. Endogenous nitration of iron regulatory protein-1 (IRP-1) in nitric oxide-producing murine macrophages: further insight into the mechanism of nitration in vivo and its impact on IRP-1 function. *J Biol Chem* 279: 43345–43351, 2004.
 104. Gow AJ, Malcolm S, and Ischiropoulos H. Effects of peroxynitrite-induced protein modifications on tyrosine phosphorylation and degradation. *FEBS Lett* 385: 63–66, 1996.
 105. Grasemann H, Gaston B, Fang K, Paul K, and Ratjen F. Decreased levels of nitrosothiols in the lower airways of patients with cystic fibrosis and normal pulmonary function. *J Pediatr* 135: 770–772, 1999.
 106. Graziewicz M, Wink DA, and Laval F. Nitric oxide inhibits DNA ligase activity: potential mechanisms for NO-mediated DNA damage. *Carcinogenesis* 17: 2501–2505, 1996.
 107. Griendling KK, Sorescu D, and Ushio-Fukai M. NAD(P)H oxidase: role in cardiovascular biology and disease. *Circ Res* 86: 494–501, 2000.
 108. Gu Z, Kaul M, Yan B, Kridel SJ, Cui J, Strongin A, Smith JW, Liddington RC, and Lipton SA. S-Nitrosylation of matrix metalloproteinases: signaling pathway to neuronal cell death. *Science* 297: 1186–1190, 2002.
 109. Guo FH, Comhair SA, Zheng S, Dweik RA, Eissa NT, Thomassen MJ, Calhoun W, and Erzuzum SC. Molecular mechanisms of increased nitric oxide (NO) in asthma: evidence for transcriptional and post-transcriptional regulation of NO synthesis. *J Immunol* 164: 5970–5980, 2000.
 110. Haendeler J, Hoffmann J, Tischler V, Berk BC, Zeiher AM, and Dimmeler S. Redox regulatory and anti-apoptotic functions of thioredoxin depend on S-nitrosylation at cysteine 69. *Nat Cell Biol* 4: 743–749, 2002.
 111. Hammarström AK and Gage PW. Nitric oxide increases persistent sodium current in rat hippocampal neurons. *J Physiol* 520: 451–461, 1999.
 112. Hare JM and Stamler JS. NO/redox disequilibrium in the failing heart and cardiovascular system. *J Clin Invest* 115: 509–517, 2005.
 113. Harskamp RE and Zeeman GG. Preeclampsia: at risk for remote cardiovascular disease. *Am J Med Sci* 334: 291–295, 2007.
 114. Hart JD and Dulhunty AF. Nitric oxide activates or inhibits skeletal muscle ryanodine receptors depending on its contraction, membrane potential and ligand binding. *J Membr Biol* 173: 227–236, 2000.
 115. Hartell NA, Archer HE, and Bailey CJ. Insulin-stimulated endothelial nitric oxide release is calcium independent and mediated via protein kinase B. *Biochem Pharmacol* 69: 781–790, 2005.
 116. Hedberg JJ, Griffiths WJ, Nilsson SJF, and Hoog JO. Reduction of S-nitrosoglutathione by human alcohol dehydrogenase 3 is an irreversible reaction as analysed by electrospray mass spectrometry. *Eur J Biochem* 270: 1249–1256, 2003.
 117. Hellberg CB, Boggs SE, and Lapetina EG. Phosphatidylinositol 3-kinase is a target for protein tyrosine nitration. *Biochem Biophys Res Commun* 252: 313–317, 1998.
 118. Hensley K, Maidt ML, Yu Z, Sang H, Markesbery WR, and Floyd RA. Electrochemical analysis of protein nitrotyrosine and dityrosine in the Alzheimer brain indicates region-specific accumulation. *J Neurosci* 18: 8126–8132, 1998.
 119. Hess DT, Matsumoto A, Kim SO, Marshall HE, and Stamler JS. Protein S-nitrosylation: purview and parameters. *Nat Rev Mol Cell Biol* 6: 150–166, 2005.
 120. Hidalgo C, Donoso P, and Carrasco MA. The ryanodine receptors Ca^{2+} release channels: cellular redox sensors? *IUBMB Life* 57: 315–322, 2005.
 121. Higashida H, Yokoyama S, Hoshi N, Egorova A, Zhong ZG, Noda M, Shahidillah M, Taketo M, Knijnik R, Kimura Y, Takahashi H, Chen XL, Shin Y, and Zhang JS. Signal transduction from bradykinin, angiotensin, adrenergic and muscarinic receptors to effector enzymes, including ADP-ribosyl cyclase. *Biol Chem* 382: 23–30, 2001.
 122. Hink U, Li H, Mollnau H, Oelze M, Matheis E, Hartmann M, Skatchkov M, Thaiss F, Stahl RA, Warnholtz A, Meinertz T, Griendling K, Harrison DG, Forstermann U, and Munzel T. Mechanisms underlying endothelial dysfunction in diabetes mellitus. *Circ Res* 88: E14–E22, 2001.
 123. Hirst DG and Robson T. Nitrosative stress in cancer therapy. *Front Biosci* 12: 3406–3418, 2007.
 124. Hodara R, Norris EH, Giasson BI, Mishizen-Eberz AJ, Lynch DR, Lee VM, and Ischiropoulos H. Functional consequences of α -synuclein tyrosine nitration: diminished binding to lipid vesicles and increased fibril formation. *J Biol Chem* 279: 47746–47753, 2004.
 125. Hoffmann F, Feil R, Kleppisch T, and Schlossmann J. Function of cGMP-dependent protein kinases as revealed by gene deletion. *Physiol Rev* 86: 1–23, 2006.
 126. Hofseth LJ, Hussain SP, Wogan GN, and Harris CC. Nitric oxide in cancer and chemoprevention. *Free Radic Biol Med* 34: 955–968, 2003.
 127. Hopper RA and Garthwaite J. Tonic and phasic nitric oxide signals in hippocampal long-term potentiation. *J Neurosci* 26: 11513–11521, 2006.
 128. Hu H, Chiamvimonvat N, Yamagishi T, and Marban E. Direct inhibition of expressed cardiac L-type Ca^{2+} channels by S-nitrosothiol nitric oxide donors. *Circ Res* 81: 742–752, 1997.
 129. Huang J and Fosberg NE. Role of calpain in skeletal muscle protein degradation. *Proc Natl Acad Sci U S A* 95: 12100–12105, 1998.
 130. Hurt KJ, Sezen SF, Champion HC, Crone JK, Palese MA, Huang PL, Sawa A, Luo X, Musicki B, Snyder SH, and Burnett AL. Alternatively spliced neuronal nitric oxide mediates penile erection. *Proc Natl Acad Sci U S A* 103: 3440–3443, 2006.
 131. Huseby NE, Asare N, Wetting S, Mikkelsen IM, Mortensen B, Sveinbjörnsson B, and Wellman M. Nitric oxide expo-

- sure of CC531 rat colon carcinoma cells induces gamma-glutamyltransferase which may counteract glutathione and cell death. *Free Radic Res* 37: 99–107, 2003.
132. Hussain SP, Amstad P, Raja K, Ambs S, Nagashima M, Bennett WP, Shields PG, Ham AJ, Swenberg JA, Marrogi AJ, and Harris CC. Increased p53 mutation load in noncancerous colon tissue from ulcerative colitis: a cancer-prone chronic inflammatory disease. *Cancer Res* 60: 3333–3337, 2000.
133. Ichinose M, Sugiura H, Yamagata S, Koarai A, and Shirato K. Increase in reactive nitrogen species production in chronic obstructive pulmonary disease airways. *Am J Respir Crit Care Med* 162: 701–706, 2000.
134. Li M, Takenaka H, Asai J, Ibusuki K, Mizukami Y, Maruyama K, Yoon YS, Wecker A, Luedemann C, Eaton E, Silver M, Thorne T, and Losordo DW. Endothelial progenitor thrombospondin-1 mediates diabetes-induced delay in reendothelialization following arterial injury. *Circ Res* 98: 697–704, 2006.
135. Inoue S and Kawanishi S. Oxidative DNA damage induced by simultaneous generation of nitric oxide and superoxide. *FEBS Lett* 31: 86–88, 1995.
136. Isbell TS, Sun CW, Wu LC, Teng X, Vitturi DA, Branch BG, Kevil CG, Peng N, Wyss JM, Ambalavanan N, Schwiebert L, Ren J, Pawlik KM, Renfrow MB, Patel RP, and Townes TM. SNO-hemoglobin is not essential for red blood cell-dependent hypoxic vasodilation. *Nature Med* (doi: 10.1038/nm1771).
137. Jadeski LC, Chakraborty C, and Lala PK. Nitric oxide-mediated promotion of mammary tumour cell migration requires sequential activation of nitric oxide synthase, guanylate cyclase and mitogen-activated protein kinase. *Int J Cancer* 106: 496–504, 2003.
138. Janmey PA. The cytoskeleton and cell signaling: component localization and mechanical coupling. *Physiol Rev* 78: 763–781, 1998.
139. Jenkins DC, Charles IG, Thomsen LL, Moss DW, Holmes LS, Baylis SA, Rhodes P, Westmore K, Emson PC, and Moncada S. Roles of nitric oxide in tumor growth. *Proc Natl Acad Sci U S A* 92: 4392–4396, 1995.
140. Jeong SY, Ha TS, Park CS, Uhm DY, and Chung S. Nitric oxide directly activates large conductance Ca^{2+} -activated K^{+} channels (rSlo). *Mol Cells* 12: 97–102, 2001.
141. Jiang J, Corbett J, Hogg N, and Mason RP. An electron paramagnetic resonance investigation of the oxygen dependence of the arterial-venous gradient of nitrosyl hemoglobin in blood circulation. *Free Radic Biol Med* 43: 1208–1215, 2007.
142. Jilling T, Haddad I, Cheng S, and Matalon S. Nitric oxide inhibits heterologous CFTR expression in polarized epithelial cells. *Am J Physiol* 277: L89–L96, 1999.
143. Johnson F and Giulivi C. Superoxide dismutases and their impact upon human health. *Mol Aspects Med* 26: 340–352, 2005.
144. Jope RS, Zhang L, and Song L. Peroxynitrite modulates the activation of p38 and extracellular regulated kinases in PC12 cells. *Arch Biochem Biophys* 376: 365–370, 2000.
145. Kaneki M, Shimizu N, Yamada D, and Chang K. Nitrosative stress and pathogenesis of insulin resistance. *Antioxid Redox Signal* 9: 319–329, 2007.
146. Kanetsuna Y, Takahashi K, Nagata M, Gannon MA, Breyer MD, Harris RC, and Takahashi T. Deficiency of endothelial nitric-oxide synthase confers susceptibility to diabetic nephropathy in nephropathy-resistant inbred mice. *Am J Pathol* 170: 1473–1484, 2007.
147. Kanesaki Y, Suzuki D, Uehara G, Toyoda M, Katoh T, Sakai H, and Watanabe T. Vascular endothelial growth factor gene expression is correlated with glomerular neovascularization in human diabetic nephropathy. *Am J Kidney Dis* 45: 288–294, 2005.
148. Keshavarzian A, Banan A, Farhadi A, Komanduri S, Mutlu E, Zhang Y, and Fields JZ. Increases in free radicals and cytoskeletal protein oxidation and nitration in the colon of patients with inflammatory bowel disease. *Gut* 52: 720–728, 2003.
149. Kharitonov SA, Yates D, Robbins RA, Logan-Sinclair R, Shinebourne EA, and Barnes PJ. Increased nitric oxide in exhaled air of asthmatic patients. *Lancet* 343: 133–135, 1994.
150. Kim PK, Kwon YG, Chung HT, and Kim YM. Regulation of caspases by nitric oxide. *Ann N Y Acad Sci* 962: 42–52, 2002.
151. Kim-Shapiro DB, Schechter AN, and Gladwin MT. Unraveling the reactions of nitric oxide, nitrite, and hemoglobin in physiology and therapeutics. *Arterioscler Thromb Vasc Biol* 26: 697–705, 2006.
152. Klatt P and Lamas S. Regulation of protein function by S-glutathiolation in response to oxidative and nitrosative stress. *Eur J Biochem* 267: 4929–4944, 2000.
153. Klingström J, Akerström S, Hardestam J, Stoltz M, Simon M, Falk KI, Mirazimi A, Rottenberg M, and Lundkvist A. Nitric oxide and peroxynitrite have different antiviral effects against hantavirus replication and free mature virions. *Eur J Immunol* 36: 2649–2657, 2006.
154. Klotz L-O, Schieke SM, Sies H, and Holbrook NJ. Peroxynitrite activates the phosphoinositide 3-kinase/Akt pathway in human skin primary fibroblasts. *Biochem J* 352: 219–225, 2000.
155. Klumpp G, Schildknecht S, Nastainczyk W, Ullrich V, and Bachschmid M. Prostacyclin in the cardiovascular system: new aspects and open questions. *Pharmacol Rep* 57: 120–126, 2005.
156. Ko LW, Rush T, Sahara N, Kersh JS, Easson C, Deture M, Lin WL, Connor YD, and Yen SH. Assembly of filamentous tau aggregates in human neuronal cells. *J Alzheimers Dis* 6: 605–622, 2004.
157. Kobayashi T, Taguchi K, Takenouchi Y, Matsumoto T, and Kamata K. Insulin-induced impairment via peroxynitrite of endothelium-dependent relaxation and sarcoplasmic/endoplasmic reticulum Ca^{2+} -ATPase function in aortas from diabetic rats. *Free Radic Biol Med* 43: 431–443, 2007.
158. Koeck T, Stuehr DJ, and Aulak KS. Mitochondria and regulated tyrosine nitration. *Biochem Soc Trans* 33: 1399–1403, 2005.
159. Koh TJ and Tidball JG. Nitric oxide inhibits calpain-mediated proteolysis of talin in skeletal muscle cells. *Am J Physiol Cell Physiol* 279: C806–C812, 2000.
160. Kong SK, Yim MB, Stadtman ER, and Chock PB. Peroxynitrite disables the tyrosine phosphorylation regulatory mechanism: lymphocyte-specific tyrosine kinase fails to phosphorylate nitrate cdc2(6-20)NH₂ peptide. *Proc Natl Acad Sci U S A* 93: 3377–3382, 1996.
161. Kooy NW, Lewis SJ, Royali JA, Ye YZ, Kelly DR, and Beckman JS. Extensive tyrosine nitration in human myocardial inflammation: evidence for the presence of peroxynitrite. *Crit Care Med* 25: 812–219, 1997.
162. Korkmaz A, Yaren H, Topal T, and Oter S. Molecular targets against mustard toxicity: implication of cell surface receptors, peroxynitrite production, and PARP activation. *Arch Toxicol* 80: 662–670, 2006.

163. Kurimoto M, Endo S, Hirashima Y, Hamada H, Ogiichi T, and Takaku A. Growth inhibition and radiosensitization of cultured glioma cells by nitric oxide generating agents. *J Neurooncol* 42: 35–44, 1999.
164. Kyriakis JM and Avruch J. Mammalian mitogen-activated protein kinase signal transduction pathways activated by stress and inflammation. *Physiol Rev* 81: 807–869, 2001.
165. Lambeth JD. NOX enzymes and the biology of reactive oxygen. *Nat Rev Immunol* 4: 181–189, 2004.
166. Landar A, Zmijewski JW, Dickinson DA, Le Goffe C, Johnson MS, Milne GL, Zanoni G, Vidari G, Morrow JD, and Darley-Usmar VM. Interaction of electrophilic lipid oxidation products with mitochondria in endothelial cells and formation of reactive oxygen species. *Am J Physiol Heart Circ Physiol* 290: H1777–H1787, 2006.
167. Lander HM, Hajjar DP, Hempstead BL, Mirza UA, Chait BT, Campbell S, and Quilliam LA. A molecular redox switch on p21(ras): structural basis for the nitric oxide-p21(ras) interaction. *J Biol Chem* 272: 4323–4326, 1997.
168. Landmesser U, Dikalov S, Price SR, McCann L, Fukai T, Holland SM, Mitch WE, and Harrison DG. Oxidation of tetrahydrobiopterin leads to uncoupling of endothelial cell nitric oxide synthase in hypertension. *J Clin Invest* 111: 1201–1209, 2003.
169. Lanone S, Manivet P, Callebort J, Launay JM, Payen D, Aubier M, Boczkowski J, and Mebazaa A. Inducible nitric oxide synthase (NOS2) expressed in septic patients is nitrated on selected tyrosine residues: implications for enzymic activity. *Biochem J* 366: 399–404, 2002.
170. Lau CG and Zukin RS. NMDA receptor trafficking in synaptic plasticity and neuropsychiatric disorders. *Nat Rev Neurosci* 8: 413–426, 2007.
171. Lee JM, Nile JC, Wishnok JS, and Tannenbaum SR. Peroxynitrite reacts with 8-nitropurines to yield 8-oxopurines. *Chem Res Toxicol* 15: 7–14, 2002.
172. Leeuwenburgh C, Hardy MM, Hazen SL, Wagner P, Ohishi S, Steinbrecher UP, and Heinecke JW. Reactive nitrogen intermediates promote low density lipoprotein oxidation in human atherosclerotic intima. *J Biol Chem* 272: 1433–1436, 1997.
173. Lewis RS. The molecular choreography of a store-operated calcium channels. *Nature* 446: 284–287, 2007.
174. Li Z, Chapleau MW, Bates JN, Bielefeldt K, Lee HC, and Abboud FM. Nitric oxide as an autocrine regulator of sodium currents in baroreceptor neurons. *Neuron* 20: 1039–1049, 1998.
175. Li H, Samouilov A, Liu X, and Zweier JL. Characterization of the magnitude and kinetics of xanthine-oxidase-catalyzed nitrate reduction: evaluation of its role in nitrite and nitric oxide generation in anoxic tissues. *Biochemistry* 42: 1150–1159, 2003.
176. Li S and Whorton AR. Regulation of protein tyrosine phosphatase 1B in intact cells by S-nitrosothiols. *Arch Biochem Biophys* 410: 269–297, 2003.
177. Liaudet L, Rosselet A, Schaller MD, Markert M, Perret C, and Feihl F. Nonselective versus selective inhibition of inducible nitric oxide synthase in experimental endotoxic shock. *J Infect Dis* 177: 127–132, 1998.
178. Liberatore GT, Jackson-Lewis V, Vukosavic S, Mandir AS, Vila M, McAuliffe WG, Dawson VL, Dawson TM, and Przedborski S. Inducible nitric oxide synthase stimulates dopaminergic neurodegeneration in the MPTP model of Parkinson disease. *Nat Med* 5: 1403–1409, 1999.
179. Lim I, Yun J, Kim S, Lee C, Seo S, Kim T, and Bang H. Nitric oxide stimulates a large-conductance Ca^{2+} -activated K^{+} channel in human skin fibroblasts through protein kinase G pathway. *Skin Pharmacol Physiol* 18: 279–287, 2005.
180. Lipton SA, Choi YB, Pan ZH, Lei SZ, Chen HS, Sucher NJ, Loscalzo J, Singel DJ, and Stamler JS. A redox-based mechanism for the neuroprotective and neurodestructive effects of nitric oxide and related nitroso-compounds. *Nature* 364: 626–632, 1993.
181. Liu D, Bao F, Wen J, and Liu J. Mutation of superoxide dismutase elevates reactive species: comparison of nitration and oxidation of proteins in different brain regions of transgenic mice with amyotrophic lateral sclerosis. *Neuroscience* 146: 255–264, 2007.
182. Liu L, Hausladen A, Zeng M, Que L, Heitman J, and Stamler JS. A metabolic enzyme for S-nitrosothiol conserved from bacteria to humans. *Nature* 410: 490–494, 2001.
183. Liu VWT and Huang PL. Cardiovascular roles of nitric oxide: a review of insights from nitric oxide synthase gene disrupted mice. *Cardiovasc Res* 77: 19–29, 2008.
184. Liu L, Yan Y, Zeng M, Zhang J, Hanes MA, Ahearn G, McMahon TJ, Dickfield T, Marshall HE, Que LG, and Stamler JS. Essential roles of S-nitrosothiols in vascular homeostasis and endotoxic shock. *Cell* 116: 617–628, 2004.
185. Lohmann SM, Vaandrager AB, Smolenski A, Walter U, and De Jonge HR. Distinct and specific functions of cGMP-dependent protein kinases. *Trends Biochem Sci* 22: 307–312, 1997.
186. Lokuta AJ, Maertz NA, Meethal SV, Potter KT, Kamp TJ, Valdivia HH, and Haworth RA. Increased nitration of sarcoplasmic reticulum Ca^{2+} -ATPase in human heart failure. *Circulation* 111: 988–995, 2005.
187. Lopez-Farré AL and Casado S. Heart failure, redox alterations, and endothelial dysfunction. *Hypertension* 38: 1400–1405, 2001.
188. Lowry KS, Murray SS, McLean CA, Talman P, Mathers S, Lopes EC, and Cheema SS. A potential role for the p75 low-affinity neurotrophin receptor in spinal motor neuron degeneration in murine and human amyotrophic lateral sclerosis. *Amyotrophic Lateral Scler Other Motor Neuron Disord* 2: 127–134, 2001.
189. Lu G, Mazet B, Sarr MG, and Szurszewski JH. Effect of nitric oxide on calcium-activated potassium channels in colonic smooth muscle of rabbits. *Am J Physiol* 274: G848–G856, 1998.
190. Lytton J. $\text{Na}^{+}/\text{Ca}^{2+}$ exchangers: three mammalian gene families control Ca^{2+} transport. *Biochem J* 406: 365–382, 2007.
191. Ma XL, Gao F, Nelson AH, Lopez BL, Christopher TA, Yue TL, and Barone FC. Oxidative inactivation of nitric oxide and endothelial dysfunction in stroke-prone spontaneous hypertensive rats. *J Pharmacol Exp Ther* 298: 879–885, 2001.
192. Maczewski M and Beresewicz A. Role of nitric oxide and free radicals in cardioprotection by blocking $\text{Na}^{+}/\text{H}^{+}$ and $\text{Na}^{+}/\text{Ca}^{2+}$ exchange in rat heart. *Eur J Pharmacol* 461: 139–147, 2003.
193. Maehama T. PETN: its deregulation and tumorigenesis. *Biol Pharm Bull* 30: 1624–1627, 2007.
194. Marshall HE and Stamler JS. Nitrosative stress-induced apoptosis through inhibition of NF- κ B. *J Biol Chem* 277: 34223–34228, 2002.
195. Martin E, Berka V, Tsai AL, and Murad F. Soluble guanylyl cyclase: the nitric oxide receptor. *Methods Enzymol* 396: 478–492, 2005.
196. Martínez MC, Tesse A, Zobairi F, and Andriantsitohaina R. Shed membrane microparticles from circulating and vas-

- cular cells in regulating vascular function. *Am J Physiol* 288: H1004–H1009, 2005.
197. Masters CL, Simms G, Weinman NA, Multhaup G, McDonald BL, and Beyreuther K. Amyloid plaque core protein in Alzheimer disease and Down syndrome. *Proc Natl Acad Sci U S A* 82: 4245–4249, 1985.
198. Matsuda T, Nagao T, Takemura M, and Baba A. Topics on the $\text{Na}^+/\text{Ca}^{2+}$ exchanger: responses of $\text{Na}^+/\text{Ca}^{2+}$ exchanger to interferon-gamma and nitric oxide in cultured microglia. *J Pharmacol Sci* 102: 22–26, 2006.
199. Matsushita K, Morrell CN, Cambien B, Yang SX, Yamakuchi M, Bao C, Hara MR, Wuick RA, Cao W, O'Rourke B, Lowenstein JM, Pevsner J, Wagenr DD, and Lowenstein CJ. Nitric oxide regulates exocytosis by S-nitrosylation of N-ethylmaleimide-sensitive factor. *Cell* 115: 139–150, 2003.
200. Medeiros R, Prediger RD, Passos GF, Pandolfo P, Duarte FS, Franco JL, Dafre AL, Di Giunta G, Figueiredo CP, Takahashi RN, Campos MM, and Calixto JB. Connecting TNF- α signaling pathways to iNOS expression in a mouse model of Alzheimer's disease: relevance for the behavioral and synaptic deficits induced by amyloid beta protein. *J Neurosci* 27: 5394–5404, 2007.
201. Méry PF, Pavoine C, Belhassen L, Pecker F, and Fischmeister R. Nitric oxide regulates cardiac Ca^{2+} current: involvement of cGMP-inhibited and cGMP-stimulated phosphodiesterases through guanylyl cyclase activation. *J Biol Chem* 268: 26286–26295, 1993.
202. Meszaros LG, Minarovic I, and Zahradnikova A. Inhibition of the skeletal muscle ryanodine receptor calcium release channel by nitric oxide. *FEBS Lett* 3: 8049–8052, 1996.
203. Meziani F, Tesse A, David E, Martínez MC, Wangesteen R, Schneider F, and Andriantsitohaina R. Shed membrane particles from preeclamptic women generate vascular wall inflammation and blunt vascular contractility. *Am J Pathol* 169: 1473–1483, 2006.
204. Meziani F, Kremer H, Tesse A, Baron-Menguy C, Mathien C, Mostefai HA, Carusio N, Schneider F, Asfar P, and Andriantsitohaina R. Human serum albumin improves arterial dysfunction during early resuscitation in mouse endotoxic model via reduced oxidative and nitrosative stresses. *Am J Pathol* 171: 1753–1761, 2007.
205. Mihm MJ, Yu F, Weinstein DM, Reiser PJ, and Bauer JA. Intracellular distribution of peroxynitrite during doxorubicin cardiomyopathy: evidence for selective impairment of myofibrillar creatine kinase. *Br J Pharmacol* 135: 581–588, 2002.
206. Millar TM, Stevens CR, Benjamin N, Eisenthal R, Harrison R, and Blake DR. Xanthine oxidoreductase catalyzes the reduction of nitrates and nitrite to nitric oxide under hypoxic conditions. *FEBS Lett* 427: 225–228, 1998.
207. Milsom AB, Jones CJ, Goodfellow J, Frenneaux MP, Peters JR, and James PE. Abnormal metabolic fate of nitric oxide in type I diabetes mellitus. *Diabetologia* 45: 1515–1522, 2002.
208. Minowa T, Miwa S, Kobayashi S, Enoki T, Zhang XF, Komuro T, Iwamuro Y, and Masaki T. Inhibitory effect of nitrovasodilators and cyclic GMP on ET-1-activated Ca^{2+} -permeable nonselective cation channel in rat aortic smooth muscle cells. *Br J Pharmacol* 120: 1536–1544, 1997.
209. Mishra OP, Qayyum I, and Delivoria-Papadopoulos M. Hypoxia-induced modification of the inositol triphosphate receptor in neuronal nuclei of newborn piglets: role of nitric oxide. *J Neurosci Res* 74: 333–338, 2003.
210. Mistry DK and Garland CJ. Nitric oxide (NO)-induced activation of large conductance Ca^{2+} -dependent K^+ channels (BK(Ca)) in smooth muscle cells isolated from the rat mesenteric artery. *Br J Pharmacol* 124: 1131–1140, 1998.
211. Mitchell DA, Morton SU, Fernhoff NB, and Marletta MA. Thioredoxin is required for S-nitrosation of pro-caspase-3 and the inhibition of apoptosis in Jurkat cells. *Proc Natl Acad Sci U S A* 104: 11609–11614, 2007.
212. Moncada S, Palmer RMJ, and Higgs EA. Nitric oxide: physiology, pathophysiology, pharmacology. *Pharmacol Rev* 43: 109–141, 1991.
213. Monti LD, Barlassina C, Citterio L, Galluccio E, Berzuini C, Setola E, Valsecchi G, Lucotti P, Pozza G, Bernardinelli L, Casari G, and Piatti P. Endothelial nitric oxide synthase polymorphisms are associated with type 2 diabetes and the insulin resistance syndrome. *Diabetes* 52: 1270–1275, 2003.
214. Mori F, Tanji K, Zhang H, Kakita A, Takahashi H, and Wakabayashi K. Alpha-Synuclein pathology in the neostriatum in Parkinson's disease. *Acta Neuropathol* 10.1007/s00401-007-0316-4, 2007.
215. Moroi M, Zhang L, Yasuda T, Virmani R, Gold HK, Fishman MC, and Huang PL. Interaction of genetic deficiency of endothelial nitric oxide, gender, and pregnancy in vascular response to injury in mice. *J Clin Invest* 101: 1225–1232, 1998.
216. Muchowski PJ and Wacker JL. Modulation of neurodegeneration by molecular chaperones. *Nat Rev Neurosci* 6: 11–22, 2005.
217. Muller B, Kleschyov AL, Alencar JL, Vanin A, and Stoclet JC. Nitric oxide transport and storage in the cardiovascular system. *Ann N Y Acad Sci* 962: 131–139, 2002.
218. Muniyappa R and Quon MJ. Insulin action and insulin resistance in vascular endothelium. *Curr Opin Clin Nutr Metab Care* 10: 523–530, 2007.
219. Murad F. Shattuck lecture: Nitric oxide and cyclic GMP in cell signaling and drug development. *N Engl J Med* 355: 2003–2011, 2006.
220. Nakagawa H, Komai N, Takusagawa M, Miura Y, Toda T, Miyata N, Ozawa T, and Ikota N. Nitration of specific tyrosine residues of cytochrome c is associated with caspase-cascade inactivation. *Biol Pharm Bull* 30: 15–20, 2007.
221. Nakagawa T, Sato W, Glushkova O, Heinig M, Clarke T, Campbell-Thompson M, Yuzawa Y, Atkinson MA, Johnson RJ, and Croker B. Diabetic endothelial nitric oxide synthase knockout mice develop advanced diabetic nephropathy. *J Am Soc Nephrol* 18: 539–550, 2007.
222. Nakamura T and Lipton SA. S-Nitrosylation and uncompetitive/fast off-rate (UFO) drug therapy in neurodegenerative disorders of protein misfolding. *Cell Death Diff* 14: 1305–1314, 2007.
223. Nakamura T and Lipton SA. Emerging roles of S-nitrosylation in protein misfolding and neurodegenerative diseases. *Antioxid Redox Signal* 10: 87–102, 2008.
224. Napoli C and Ignarro LJ. Nitric oxide and atherosclerosis. *Nitric Oxide* 5: 88–97, 2001.
225. Nathan C. Specificity of a third kind: reactive oxygen and nitrogen intermediates in cell signaling. *J Clin Invest* 111: 769–778, 2003.
226. Nathan C and Xie QW. Regulation of biosynthesis of nitric oxide. *J Biol Chem* 269: 13725–13728, 1994.
227. Nethery DE, Ghosh S, Erzurum SC, and Kern JA. Inactivation of neuregulin-1 by nitration. *Am J Physiol Lung Cell Mol Physiol* 292: L287–L294, 2007.
228. Neumann P, Gertzberg N, Vaughan E, Weisbrot J, Woodburn R, Lambert W, and Johnson A. Peroxynitrite mediates TNF- α -induced endothelial barrier dysfunction and ni-

- tration of actin. *Am J Physiol Lung Cell Mol Physiol* 290: L674–L684, 2006.
229. Nicolis S, Pennati A, Perani E, Monzani E, Sngangelantoni AM, and Casella L. Easy oxidation and nitration of human myoglobin by nitrite and hydrogen peroxide. *Chemistry* 12: 749–757, 2006.
 230. Nomiya T, Igarashi Y, Taka H, mineki R, Uchida T, Ogi-hara T, Choi JB, Uchino H, Tanaka Y, Maegawa H, Kashi-wagi A, Murayama K, Kawamori R, and Watada H. Red-uction of insulin-stimulated glucose uptake by peroxynitrite is concurrent with tyrosine nitration of in-sulin receptor substrate-1. *Biochem Biophys Res Commun* 320: 639–647, 2004.
 231. Oak JH and Cai H. Attenuation of angiotensin II signaling recouples eNOS and inhibits nonendothelial NOX activity in diabetic mice. *Diabetes* 56: 118–126, 2007.
 232. Obrosova IG, Drel VR, Oltman CL, Mashtalir N, Tibrewala J, Groves JT, and Yorek MA. Role of nitrosative stress in early neuropathy and vascular dysfunction in streptozo-tocin-diabetic rats. *Am J Physiol Endocrinol Metab* 293: E1645–E1655, 2007.
 233. Ogino K and Wang DH. Biomarkers of oxidative/nitrosative stress: an approach to disease prevention. *Acta Med Okayama* 61: 181–189, 2007.
 234. Ohshima H, Sawa T, and Akaiki T. 8-Nitroguanine, a prod-uct of nitrative DNA damage caused by reactive nitrogen species: formation, occurrence, and implications in inflam-mation and carcinogenesis. *Antioxid Redox Signal* 8: 1033–1045, 2006.
 235. Ohsugi S, Iwasaki Y, Takemura Y, Nagata K, Harada H, Yokomura I, Hosogi S, Yuba T, Niisato N, Miyazaki H, Mat-subara H, Fushiki S, and Marunaka Y. An inhaled inducible nitric oxide synthase inhibitor reduces damage of Candida-induced acute lung injury. *Biomed Res* 28: 91–99, 2007.
 236. Okouchi M, Ekshyyan O, Maracine M, and Aw TY. Neu-ronal apoptosis in neurodegeneration. *Antioxid Redox Sig-nal* 9: 1060–1096, 2007.
 237. Okuno T, Natasuji Y, Kumanogoh A, Moriya M, Ichinose H, Sumi H, Fujimura H, Kikutani H, and Sakoda S. Loss of dopaminergic neurons by the induction of inducible nitric oxide synthase and cyclooxygenase-2 via CD40: relevance to Parkinson's disease. *J Neurosci Res* 81: 874–882, 2005.
 238. Pacher P, Beckman JS, and Liaudet L. Nitric oxide and per-oxynitrite in health and disease. *Physiol Rev* 87: 315–424, 2007.
 239. Palmer LA, Doctor A, Chhabra P, Sheram ML, Laubach VE, Karlinsey MZ, Forbes MS, Macdonald T, and Gaston B. S-Nitrothiols signal hypoxia-mimetic vascular pathology. *J Clin Invest* 117: 2592–2601, 2007.
 240. Pannu R and Singh I. Pharmacological strategies for the regulation of inducible nitric oxide synthase: neurodegen-erative versus neuroprotective mechanisms. *Neurochem Int* 49: 170–182, 2006.
 241. Paravicini TM and Touyz R. Redox signaling in hyperten-sion. *Cardiovasc Res* 71: 247–258, 2006.
 242. Park HS, Mo JS, and Choi EJ. Nitric oxide inhibits an in-teraction between JNK1 and c-Jun through nitrosylation. *Biochem Biophys Res Commun* 351: 281–286, 2006.
 243. Patwari P and Lee RT. Thioredoxins, mitochondria, and hy-pertension. *Am J Pathol* 170: 805–808, 2007.
 244. Pehar M, Vargas MR, Robinson KM, Cassina P, England P, Beckman JS, Alzari PM, and Barbeito L. Peroxynitrite transforms nerve growth factor into an apoptotic factor for motor neurons. *Free Radic Biol Med* 41: 1632–1644, 2006.
 245. Peluffo G and Radi R. Biochemistry of protein tyrosine ni-tration in cardiovascular pathology. *Cardiovasc Res* 75: 291–302, 2007.
 246. Pennathur S, Bergt C, Shao B, Byun J, Kassim SY, Singh P, Green PS, McDonald TO, Brunzell J, Chait A, Oram JF, O'Brien K, Geary RL, and Heinecke JW. Human athero-sclerotic intima and blood of patients with established coro-nary artery disease contain high density lipoprotein dam-aged by reactive nitrogen species. *J Biol Chem* 279: 42977–42983, 2004.
 247. Pérez-Rodríguez R, Fuentes MP, Oliván AM, Martínez-Palacián A, Romero C, González MP, and Oset-Gasque MJ. Mechanisms of nitric oxide-induced apoptosis in bovine chromaffin cells: role of mitochondria and apoptotic pro-teins. *J Neurosci Res* 85: 2224–2238, 2007.
 248. Perrotta C, Bizzozero L, Falcone S, Rovere-Querini P, Prinetti A, Schuchman EH, Sonnino S, Manfredi AA, and Clementi E. Nitric oxide boosts chemoimmunotherapy via inhibition of acid sphingomyelinase in a mouse model of melanoma. *Cancer Res* 67: 7559–7564, 2007.
 249. Pesse B, Levrand S, Feihl F, Waeber B, Gavillet B, Pacher P, and Liaudet L. Peroxynitrite activates ERK via Raf-1 and MEK, independently from EGF receptor and p21Ras in H9C2 cardiomyocytes. *J Mol Cell Cardiol* 38: 765–775, 2005.
 250. Petros A, Lamb G, Leone A, Moncada S, Bennett D, and Vallance P. Effects of a nitric oxide synthase inhibitor in humans with septic shock. *Cardiovasc Res* 28: 34–39, 1994.
 251. Pilane CM and LaBelle EF. NO induced apoptosis of vas-cular smooth muscle cells accompanied by ceramide in-crease. *J Cell Physiol* 199: 310–315, 2004.
 252. Pinzar E, Wang T, Garrido MR, Xu W, Levy P, and Bottari SP. Angiotensin II induces tyrosine nitration and activation of ERK1/2 in vascular smooth muscle cells. *FEBS Lett* 579: 5100–5104, 2005.
 253. Poteser M, Romanin C, Schreimbayer W, Mayer B, and Groschner K. S-nitrosation controls gating and conduc-tance of the $\alpha 1$ subunit of class C L-type Ca^{2+} chan-nels. *J Biol Chem* 276: 14797–14803, 2001.
 254. Potoka DA, Upperman JS, Zhang XR, Kaplan JR, Corey SJ, Grishin A, Zamora R, and Ford HR. Peroxynitrite inhibits enterocyte proliferation and modulates Src kinase activity in vitro. *Am J Physiol Gastrointest Liver Physiol* 285: G861–G869, 2003.
 255. Pou S, Pou WS, Bredt DS, Snyder SH, and Rosen GM. Gen-eration of superoxide by purified brain nitric oxide syn-thase. *J Biol Chem* 267: 24173–24176, 1992.
 256. Prasad R, Giri S, Nath N, Singh I, and Singh AK. GSNO at-tenuates EAE disease by S-nitrosylation-mediated modu-lation of endothelial-monocyte interactions. *Glia* 55: 65–77, 2007.
 257. Pritchard KA Jr, Ackerman AW, Gross ER, Stepp DW, Shi Y, Fontana JT, Baker JE, and Sessa WC. Heat shock protein 90 mediates the balance of nitric oxide and superoxide an-ion from endothelial nitric-oxide synthase. *J Biol Chem* 276: 17621–17624, 2001.
 258. Przedborski S, Chen Q, Vila M, Giasson BI, Djaldatti R, Vukosavic S, Souza JM, Jackson-Lewis V, Lee VM, and Is-chiropoulos H. Oxidative post-translational modifications of α -synuclein in the 1-methyl-4-phenyl-1,2,3,6-tetrahy-droindole (MPTP) mouse model of Parkinson's disease. *J Neurochem* 76: 637–640, 2001.
 259. Pyriochou A, Beis D, Koika V, Potytarchou C, Papadim-itrou E, Zhou Z, and Papapetropoulos A. Soluble guanylyl cyclase activation promotes angiogenesis. *J Pharmacol Exp Ther* 319: 663–671, 2006.

260. Radi R. Nitric oxide, oxidants, and protein tyrosine nitration. *Proc Natl Acad Sci U S A* 101: 4003–4008, 2004.
261. Rafikova O, Rafikov R, and Nudler E. Catalysis of S-nitrosothiols formation by serum albumin: the mechanism and implication in vascular control. *Proc Natl Acad Sci U S A* 99: 5913–5918, 2002.
262. Rajagopalan S, Meng XP, Ramasamy S, Harrison DG, and Galis ZS. Reactive oxygen species produced by macrophage-derived foam cells regulate the activity of vascular matrix metalloproteinase in vitro. *J Clin Invest* 98: 2572–2579, 1996.
263. Ralay Ranaivo H, Carusio N, Wangenstein R, Ohlmann P, Loichot C, Tesse A, Chalupsky K, Lobysheva I, Haiech J, Watterson DM, and Andriantsitohaina R. Protection against endotoxic shock as a consequence of reduced nitrosative stress in MLCK210-null mice. *Am J Pathol* 170: 439–446, 2007.
264. Rebrin I, Br  g  re C, Kamzalov S, Gallaher TK, and Sohal RS. Nitration of tryptophan 372 in succinyl-CoA:3-ketoacid CoA transferase during aging in rat heart mitochondria. *Biochemistry* 46: 10130–10144, 2007.
265. Reddy S, Jones AD, Cross CE, Wong PS, and van der Vliet A. Inactivation of creatine kinase by S-glutathionylation of the active-site cysteine residue. *Biochem J* 347: 821–827, 2000.
266. Redout EM, Wagner MJ, Zuidwijk MJ, Boer C, Musters RJ, van Hardeveld C, Paulus WJ, and Simonides WS. Right-ventricular failure is associated with increased mitochondrial complex II activity and production of reactive oxygen species. *Cardiovasc Res* 75: 770–781, 2007.
267. Reinehr R, G  rg B, H  ngen A, and H  ussinger D. CD95-tyrosine nitration inhibits hyperosmotic and CD95 ligand-induced CD95 activation in rat hepatocytes. *J Biol Chem* 279: 10364–10373, 2004.
268. Renganathan M, Cummins TR, and Waxman SG. Nitric oxide blocks fast, slow, and persistent Na⁺ currents in C-type DRG neurons by S-nitrosylation. *J Neurophysiol* 87: 761–775, 2002.
269. Reynaert NL, Ckless K, Korn SH, Vos N, Guala AS, Wouters EFM, van der Vliet A, and Janssen-Heininger YMW. Nitric oxide represses inhibitory κ B kinase through S-nitrosylation. *Proc Natl Acad Sci U S A* 101: 8945–8950, 2004.
270. Reynolds MR, Berry RW, and Binder LI. Site-specific nitration and oxidative dihydroxylation of the tau protein by peroxynitrite: Implication for Alzheimer's disease. *Biochemistry* 44: 1690–1700, 2005.
271. Reynolds MR, Reyes JF, Fu Y, Bigio EH, Guillozet-bongaerts AI, Berry RW, and Binder LI. Tau nitration occurs at tyrosine 29 in the fibrillar lesions of Alzheimer's disease and other tauopathies. *J Neurosci* 26: 10636–10645, 2006.
272. Reynolds MR, Berry RW, and Binder LI. Nitration in neurodegeneration: deciphering the "Hows" "nYs". *Biochemistry* 46: 7325–7336, 2007.
273. Ricciardolo FLM, Sterk PJ, Gaston B, and Folkerts G. Nitric oxide in health and disease of the respiratory system. *Physiol Rev* 84: 731–765, 2004.
274. Richards EM, Rosenthal RE, Kristian T, and Fiskum G. Postischemic hyperoxia reduces hippocampal pyruvate dehydrogenase activity. *Free Radic Biol Med* 40: 1960–1970, 2006.
275. Riobo NA, Clementi E, Melani M, Boveris A, Cadenas E, Moncada S, and Poderoso JJ. Nitric oxide inhibits mitochondrial NADH:ubiquinone reductase activity through peroxynitrite formation. *Biochem J* 359: 139–145, 2001.
276. Rizzo MA and Piston DW. Regulation of beta cell glucokinase by S-nitrosylation and association with nitric oxide synthase. *J Cell Biol* 161: 243–248, 2003.
277. Rizzuto R and Pozzan T. Microdomains of intracellular Ca²⁺: molecular determinants and functional consequences. *Physiol Rev* 86: 369–408, 2006.
278. Rodr  guez-Iturbe B, Zhan CD, Quiroz Y, Sindhu RK, and Vaziri ND. Antioxidant-rich diet relieves hypertension and reduces renal immune infiltration in spontaneously hypertensive rats. *Hypertension* 41: 341–346, 2003.
279. Sardo MA, Castaldo M, Cinquegrani M, Bonaiuto M, Maesano A, Versace A, Spadar M, Campo S, Nicocia G, Altavilla D, and Saitta A. Effects of atorvastatin treatment on sICAM-1 and plasma nitric oxide levels in hypercholesterolemic subjects. *Clin Appl Thromb Hemost* 8: 257–263, 2002.
280. Sawa T, Akaike T, Ichimori K, Akuta T, Kaneko K, Nakayama H, Stuehr DJ, and Maeda H. Superoxide generation mediated by 8-nitroguanosine, a highly redox-active nucleic acid derivative. *Biochem Biophys Res Commun* 309: 567–571, 2003.
281. Sawa T and Ohshima H. Nitrate DNA damage in inflammation and its possible role in carcinogenesis. *Nitric Oxide* 14: 91–100, 2006.
282. Sawa T, Zaki MH, Okamoto T, Akuta T, Tokutomi Y, Kim-Mitsuyama S, Ihara H, Kobayashi H, Yamamoto M, Fujii S, Arimoto H, and Akaike T. Protein S-guanylation by the biological signal 8-nitroguanosine 3',5'-cyclic monophosphate. *Nat Chem Biol* 3:727–735, 2007.
283. Schapiro JM, Libby SJ, and Fang FC. Inhibition of bacterial DNA replication by zinc mobilization during nitrosative stress. *Proc Natl Acad Sci U S A* 100: 8496–8501, 2003.
284. Schmidt T, Zaib F, Samson SE, Kwan CY, and Grover AK. Peroxynitrite resistance of sarco/endoplasmic reticulum Ca²⁺ pump in pig coronary artery endothelium and smooth muscle. *Cell Calcium* 36: 77–82, 2004.
285. Schonhoff CM, Gaston B, and Mannick JB. Nitrosylation of cytochrome c during apoptosis. *J Biol Chem* 278: 18265–18270, 2003.
286. Schopfer FJ, Lin Y, Baker PR, Cui T, Garcia-Barrio M, Zhang J, Chen K, Chen YE, and Freeman BA. Nitrolinoleic acid: an endogenous peroxisome proliferators-activated receptor gamma ligand. *Proc Natl Acad Sci U S A* 102: 2340–2345, 2004.
287. Schopfer FJ, Baker PRS, Giles G, Chumley P, Batthyany C, Crawford J, Patel RP, Hogg N, Braunchaud BP, Lancaster JR Jr, and Freeman BA. Fatty acid transduction of nitric oxide signaling: nitrolinoleic acid is a hydrophobically stabilized nitric oxide donor. *J Biol Chem* 280: 19289–19297, 2005.
288. Schreiter ER, Rodr  guez MM, Weichsel A, Montfort WR, and Bonaventura J. S-nitrosylation-induced conformational change in blackfin tuna myoglobin. *J Biol Chem* 282: 19773–19780, 2007.
289. Seabra AB, Pankotai E, Feh  r M, Somlai A, Kiss L, Szabo C, Kollai M, de Oliveira MG, and Lacza Z. S-nitrosoglutathione-containing hydrogel increases dermal blood flow in streptozotocin-induced diabetic rats. *Br J Dermatol* 156: 814–818, 2007.
290. Seddon M, Shah AM, and Casadei B. Cardiomyocytes as effectors of nitric oxide signalling. *Cardiovasc Res* 75: 315–326, 2007.
291. Servetnyk Z, Krjukova J, Gaston B, Zaman K, Hjelte L, Roomans GM, and Dragomir A. Activation of chloride transport in CF airway epithelial cell lines and primary CF

- nasal epithelial cells by S-nitrosoglutathione. *Respir Res* 7: 124, 2006.
292. Shankar RR, Wu Y, Shen HQ, Zhu JS, and Baron AD. Mice with gene disruption of both endothelial and neuronal nitric oxide synthase exhibit insulin resistance. *Diabetes* 49: 684–687, 2000.
 293. Shieh CC, Coghlan M, Sullivan JP, and Gopalakrishnan M. Potassium channels: molecular defects, diseases, and therapeutics. *Pharmacol Rev* 52: 557–594, 2000.
 294. Shishehbor MH, Aviles RJ, Brennan ML, Fu X, Goormastic M, Pearce GL, Gokce N, Keaney JF Jr, Penn MS, Sprecher DL, Vita JA, and Hazen SL. Association of nitrotyrosine levels with cardiovascular disease and modulation by statin therapy. *JAMA* 289: 1675–1680, 2003.
 295. Shrivastava P, Pantano C, Watkin R, McElhinney B, Guala A, Poynter ML, Persinger RL, Budd R, and Janssen-Heininger YJ. Reactive nitrogen species-induced cell death requires Fas-dependent activation of c-Jun N-terminal kinase. *Mol Cell Biol* 24: 6763–6772, 2004.
 296. Singh RP and Agarwal R. Inducible nitric oxide synthase-vascular endothelial growth factor axis: a potential target to inhibit tumor angiogenesis by dietary agents. *Curr Cancer Durg Targets* 7: 475–483, 2007.
 297. Singh S, Das T, Ravindran A, Chaturvedi RK, Shukla Y, Agarwal AK, and Dikshit M. Involvement of nitric oxide in neurodegeneration: a study on the experimental models of Parkinson's disease. *Redox Rep* 10: 103–109, 2005.
 298. Sjöholm A. Nitric oxide donor SIN-1 inhibits insulin release. *Am J Physiol* 271: C1098–C1102, 1996.
 299. Skinn AC and MacNaughton WK. Nitric oxide inhibits cAMP-dependent CFTR trafficking in intestinal epithelial cells. *Am J Physiol Gastrointest Liver Physiol* 289: G739–G744, 2005.
 300. Smith JM, Sondgeroth KB, and Wahler GM. Inhibition of nitric oxide synthase enhances contractile response of ventricular myocytes from streptozotocin-diabetic rats. *Mol Cell Biochem* 300: 129–137, 2007.
 301. Soler MN, Bobe P, Benihoud K, Lemaire G, Roos BA, and Lausson S. Gene therapy of rat medullary thyroid cancer by naked nitric oxide synthase II DNA injection. *J Gene Med* 2: 344–352, 2000.
 302. Staels B. PPAR agonists and the metabolic syndrome. *Therapie* 62: 319–326, 2007.
 303. Stoclet JC, Martínez MC, Ohlmann P, Chasserot S, Schott C, Kleschyov AL, Schneider F, and Andriantsitohaina R. Induction of nitric oxide synthase and dual effects of nitric oxide and cyclooxygenase products in regulation of arterial contraction in human septic shock. *Circulation* 100: 107–112, 1999.
 304. Studer RK. Nitric oxide decreases IGF-1 receptor function in vitro; glutathione depletion enhances this effect in vivo. *Osteoarthritis Cartilage* 12: 863–869, 2004.
 305. Sugiura H, Ichinose M, Yamagata S, Koarai A, Shirato K, and Hattori T. Correlation between change in pulmonary function and suppression of reactive nitrogen species production following steroid treatment in COPD. *Thorax* 58: 299–305, 2003.
 306. Sultana R, Poon HF, Cai J, Pierce WM, Merchant M, Klein JB, Markesbery WR, and Butterfield DA. Identification of nitrated proteins in Alzheimer's disease brain using a redox proteomics approach. *Neurobiol Dis* 22: 76–87, 2006.
 307. Summers BA, Overholt JL, and Prabhakar NR. Nitric oxide inhibits L-type Ca²⁺ current in glomus cells of the rabbit carotid body via a cGMP-independent mechanism. *J Neurophysiol* 81: 1449–1457, 1999.
 308. Sun J, Steenbergen C, and Murphy E. S-nitrosylation: NO-related redox signaling to protect against oxidative stress. *Antioxid Redox Signal* 8: 1693–1705, 2006.
 309. Sun J, Xu L, Eu JP, Stamler JS, and Meissner G. Nitric oxide, NOC-12, and S-nitrosoglutathione modulate the skeletal muscle calcium release channel/ryanodine receptor by different mechanisms: an allosteric function for O₂ in S-nitrosylation of the channel. *J Biol Chem* 278: 8184–8189, 2003.
 310. Swindle EJ, Hunt JA, and Coleman JW. A comparison of reactive oxygen species generation by rat peritoneal macrophages and mast cells using the highly sensitive real-time chemiluminescent probe Pholasin: inhibition of antigen-induced mast cell degranulation by macrophage-derived hydrogen peroxide. *J Immunol* 169: 5866–5873, 2002.
 311. Swindle EJ and Metcalfe DD. The role of reactive oxygen species and nitric oxide in mast cell-dependent inflammatory processes. *Immunol Rev* 217: 186–205, 2007.
 312. Szabo C, Ischiropoulos H, and Radi R. Peroxynitrite: biochemistry, pathophysiology and development of therapeutics. *Nat Rev Drug Disc* 6: 662–680, 2007.
 313. Szabo C and Ohshima H. DNA damage induced by peroxynitrite: subsequent biological effects. *Nitric Oxide* 1: 373–385, 1997.
 314. Taglialatela M, Pannaccione A, Iossa S, Castaldo P, and Annunziato L. Modulation of the K⁺ channels encoded by the human ether-a-gogo-related gene-1 (hERG1) by nitric oxide. *Mol Pharmacol* 56: 1298–1308, 1999.
 315. Taguchi K, Kobayashi T, Hayashi Y, Matsumoto T, and Kamata K. Enalapril improves impairment of SERCA-derived relaxation and enhancement of tyrosine nitration in diabetic rat aorta. *Eur J Pharmacol* 556: 121–128, 2007.
 316. Tao L, Jiao X, Gao E, Lau WB, Yuan Y, Lopez B, Christopher T, Ramachandra Rao SP, Williams W, Southan G, Sharma K, Koch W, and Ma XL. Nitrate inactivation of thioredoxin-1 and its role in postischemic myocardial apoptosis. *Circulation* 114: 1395–1402, 2006.
 317. Thatcher GRJ. An introduction to NO-related therapeutic agents. *Curr Top Med Chem* 5: 597–601, 2005.
 318. Thum T, Fraccarollo D, Schultheiss M, Froese S, Galuppo P, Widder JD, Tsikas D, Ertl G, and Bauerachs J. Endothelial nitric oxide synthase uncoupling impairs endothelial progenitor cell mobilization and function in diabetes. *Diabetes* 56: 666–674, 2007.
 319. Timmins GS, Master S, Rusnak F, and Deretic V. Requirements for nitric oxide generation from isoniazid activation in vitro and inhibition of mycobacterial respiration in vivo. *J Bacteriol* 186: 5427–5431, 2004.
 320. Torres-Rasgado E, Fouret G, Carbonneau MA, and Leger CL. Peroxynitrite mild nitration of albumin and LDL-albumin complex naturally present in plasma and tyrosine nitration rate-albumin impairs LDL nitration. *Free Radic Res* 41: 367–375, 2007.
 321. Tsuura Y, Ishida H, Hayashi S, Sakamoto K, Hrie M, and Seino Y. Nitric oxide opens ATP-sensitive K⁺ channels through suppression of phosphofructokinase activity and inhibits glucose-induced insulin release in pancreatic beta cells. *J Gen Physiol* 104: 1079–1098, 1994.
 322. Uc A, Kooy NW, Conklin JL, and Bishop WP. Peroxynitrite inhibits epidermal growth factor receptor signaling in Caco-2 cells. *Dig Dis Sci* 48: 2353–2359, 2003.
 323. Uehara T, Nakamura T, Yao D, Shi ZQ, Gu Z, Ma Y, Masliah E, Nomura Y, and Lipton SA. S-Nitrosylated protein-disulphide isomerase links protein misfolding to neurodegeneration. *Nature* 441: 513–517, 2006.

324. Uneda S, Hata H, Matsuno F, Nagasaki A, Harada N, Mitsuya Y, Matsuzaki H, and Mitsuya H. A nitric oxide synthase inhibitor, N(G)-nitro-L-arginine-methyl-ester, exerts potent antiangiogenic effects on plasmacytoma in a newly established multiple myeloma severe combined immunodeficient mouse model. *Br J Haematol* 120: 396–414, 2003.
325. Upmacis RK, Deeb RS, and Hajjar DP. Reprint of “oxidative alterations of cyclooxygenase during atherogenesis.” *Prostaglandins Other Lipid Mediat* 82: I–XIV, 2007.
326. Uppu RM and Pryor WA. Carbon dioxide catalysis of the reaction of peroxynitrite with ethyl acetoacetate: an example of aliphatic nitration by peroxynitrite. *Biochem Biophys Res Commun* 229: 764–769, 1996.
327. Uversky VN, Yamin G, Munishkina LA, Karymov MA, Millet IS, Doniach S, Lyubchenko YL, and Fink AL. Effects of nitration on the structure and aggregation of α -synuclein. *Brain Res Mol Brain Res* 134: 84–102, 2005.
328. Vanderheyden M, Bartunek J, Knaepen M, Kockx M, De Bruyne B, and Goethal M. Hemodynamic effects of inducible nitric oxide synthase and nitrotyrosine generation in heart failure. *J Heart Lung Transplant* 23: 723–728, 2004.
329. van Rossum DB, Patterson RL, Ma HT, and Gill DL. Ca²⁺ entry mediated by store depletion, S-nitrosylation, and TRP3 channels: comparison of coupling and function. *J Biol Chem* 275: 28662–28668, 2000.
330. Varenik I, Pavlov IA, Drel VR, Lyzogubov VV, Ilnytska O, Bell SR, Tibrewala J, Groves JT, and Obrosova IG. Nitrosative stress and peripheral diabetic neuropathy in leptin-deficient (ob/ob) mice. *Exp Neurol* 205: 425–436, 2007.
331. Vazquez-Vivar J, Kalyanaraman B, Martaseck P, Hogg N, Masters BS, Karoui H, Tordo P, and Pritchard KA Jr. Superoxide generation by endothelial nitric oxide synthase: the influence of cofactors. *Proc Natl Acad Sci U S A* 95: 9220–9225, 1998.
332. Vazquez-Vivar J, Kalyanaraman B, and Martaseck P. The role of tetrahydrobiopterin in superoxide generation from eNOS: enzymology and physiological implications. *Free Radic Res* 37: 121–127, 2003.
333. Vicente S, Figueroa S, Perez-Rodriguez R, Gonzalez MP, and Oset-Gasque MJ. Nitric oxide donors induce calcium-mobilisation from internal stores but do not stimulate catecholamine secretion by bovine chromaffin cells in resting conditions. *Cell Calcium* 37: 163–172, 2005.
334. Villacorta L, Zhang J, Garcia-Barrio MT, Chen XL, Freeman YEC, and Cui T. Nitro-linoleic acid inhibits vascular smooth muscle cell proliferation via the Keap1/Nrf2 signaling pathway. *Am J Physiol Heart Circ Physiol* 293: H770–H776, 2007.
335. Vinik A, Parson H, and Ullal J. The role of PPARs in the microvascular dysfunction in diabetes. *Vascul Pharmacol* 45: 54–64, 2006.
336. Vinik AI, Ullal J, Parson HK, Barlow PM, and Casellini CM. Pioglitazone treatment improves nitrosative stress in type 2 diabetes. *Diabetes Care* 29: 869–876, 2006.
337. Virag L and Szabo C. The therapeutic potential of poly(ADP-ribose) polymerase inhibitors. *Pharmacol Rev* 54: 375–429, 2002.
338. Wadham C, Parker A, Wang L, and Xia P. High glucose attenuates protein S-nitrosylation in endothelial cells: role of oxidative stress. *Diabetes* 56: 2715–2721, 2007.
339. Wang B, Wei D, Crum VE, Richardson EL, Xiong HH, Luo Y, Huang S, Abbruzzese JL, and Xie K. A novel model system for studying the double-edged roles of nitric oxide production in pancreatic cancer growth and metastasis. *Oncogene* 22: 1771–1782, 2003.
340. Wang YG, Dedkova EN, Ji X, Blatter LA, and Lipsius SL. Phenylephrine acts via IP₃-dependent intracellular NO release to stimulate L-type Ca²⁺ current in cat atrial myocytes. *J Physiol* 567: 143–157, 2005.
341. Wanstall JC, Homer KL, and Doggrell SA. Evidence for, and importance of, cGMP-independent mechanisms with NO and NO donors on blood vessels and platelets. *Curr Vasc Pharmacol* 3: 41–53, 2005.
342. Watson WH, Yang X, Choi YE, Jones DP, and Kehrer JP. Thioredoxin and its role in toxicology. *Toxicol Sci* 78: 3–14, 2004.
343. Webb A, Bond R, McLean P, Uppal R, Benjamin N, and Ahluwalia A. Reduction of nitrite to nitric oxide during ischemia protects against myocardial ischemia-reperfusion damage. *Proc Natl Acad Sci U S A* 101: 13683–13688, 2004.
344. Webster RP, Brockman D, and Myatt L. Nitration of p38 MAPK in the placenta: association of nitration with reduced catalytic activity of p38 MAPK in pre-eclampsia. *Mol Hum Reprod* 12: 677–685, 2006.
345. Wild S, Roglic G, Green A, Sicree R, and King H. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. *Diabetes Care* 27: 1047–1053, 2004.
346. Won JS and Singh I. Sphingolipid signaling and redox regulation. *Free Radic Biol Med* 40: 1875–1888, 2006.
347. Xu L, Eu JP, Meissner G, and Stamler JS. Activation of the cardiac calcium release channel (ryanodine receptor) by poly-S-nitrosylation. *Science* 279: 234–237, 1998.
348. Xu KY, Huso DL, Dawson TM, Bredt DS, and Becker LC. Nitric oxide synthase in cardiac sarcoplasmic reticulum. *Proc Natl Acad Sci U S A* 96: 657–662, 1999.
349. Xu A, Vita JA, and Keaney JF Jr. Ascorbic acid and glutathione modulate the biological activity of S-nitrosoglutathione. *Hypertension* 36: 291–295, 2000.
350. Xu S, Ying J, Jiang B, Guo W, Adachi T, Sharov V, Lazar H, Menzoian J, Knyushko TV, Bigelow D, Schoneich D, and Cohen RA. Detection of sequence-specific tyrosine nitration of manganese SOD and SERCA in cardiovascular disease and aging. *Am J Physiol Heart Circ Physiol* 290: H2220–H2227, 2006.
351. Yamin G, Uversky VN, and Fink AL. Nitration inhibits fibrillation of human α -synuclein in vitro by formation of soluble oligomers. *FEBS Lett* 542: 147–152, 2003.
352. Yang SN and Berggren PO. The role of voltage-gated calcium channels in pancreatic beta-cell physiology and pathophysiology. *Endocr Rev* 27: 621–676, 2006.
353. Yao D, Gu Z, Nakamura T, Shi ZQ, Ma Y, Gasston B, Palmer LA, Rockenstein EM, Zhang Z, Masliah E, Uehara T, and Lipton SA. Nitrosative stress linked to sporadic Parkinson’s disease: S-nitrosylation of parkin regulates its E3 ubiquitin ligase activity. *Proc Natl Acad Sci U S A* 101: 10810–10814, 2004.
354. Yasukawa T, Tokunaga E, Ota H, Sugita H, Martyn JA, and Kaneki M. S-nitrosylation-dependent inactivation of Akt/protein kinase B in insulin resistance. *J Biol Chem* 280: 7511–7518, 2005.
355. Yermilov V, Rubio J, Becchi M, Friesen MD, Pignatelli B, and Ohshima H. Formation of 8-nitroguanine by the reaction of guanine with peroxynitrite *in vivo*. *Carcinogenesis* 16: 2045–2050, 1995.
356. Yermilov V, Rubio J, and Ohshima H. Formation of 8-nitroguanine in DNA treated with peroxynitrite in vitro and its rapid removal from DNA by depurination. *FEBS Lett* 376: 207–210, 1995.
357. Yoshida T, Inoue R, Morii T, Takahashi N, Yamamoto S, Hara Y, Tominaga M, Shimizu S, Sato Y, and Mori Y. Ni-

- tric oxide activates TRP channels by cysteine S-nitrosylation. *Nat Chem Biol* 2: 596–607, 2006.
358. Zaman K, Carraro S, Doherty J, Henderson EM, Lendermon E, Liu L, Verghese G, Zigler M, Ross M, Park E, Palmer LA, Doctor A, Stamler JS, and Gaston B. S-nitrosylating agents: a novel class of compounds that increase cystic fibrosis transmembrane conductance regulator expression and maturation in epithelial cells. *Mol Pharmacol* 70: 1435–1442, 2006.
359. Zhang YJ, Xu YF, Chen WQ, Wang XC, and Wang JZ. Nitration and oligomerization of tau induced by peroxynitrite inhibits its microtubule-binding activity. *FEBS LETT* 579: 2421–2427, 2005.
360. Zhang G, Teggatz EG, Zhang AY, Koeberl MJ, Yi F, Chen L, and Li PL. Cyclic ADP ribose-mediated Ca²⁺ signaling in mediating endothelial nitric oxide production in bovine coronary arteries. *Am J Physiol Heart Circ Physiol* 290: H1172–H1181, 2006.
361. Zmijewski JW, Landar A, Watanabe N, Dickinson DA, Noguchi N, and Darley-Usmar VM. Cell signalling by oxidized lipids and the role of reactive oxygen species in the endothelium. *Biochem Soc Trans* 33: 1385–1389, 2005.

Address reprint requests to:
Ramaroson Andriantsitohaina, Ph.D.
INSERM, U771
Faculté de Médecine
Rue Haute de Reculée
Angers
F-49045 France

E-mail: ramaroson.andriantsitohaina@univ-angers.fr

Date of first submission to ARS Central, November 29, 2007;
date of final revised submission, August 20, 2008; date of acceptance, August 20, 2008.

This article has been cited by:

1. Ramaroson Andriantsitohaina, Lucie Duluc, Julio C. García#Rodríguez, Lizette Gil#del Valle, Mariela Guevara#Garcia, Gilles Simard, Raffaella Soleti, Ding#Feng Su, Luis Velásquez#Pérez, John X. Wilson, Ismail Laher. 2012. Systems biology of antioxidants. *Clinical Science* **123**:3, 173-192. [[CrossRef](#)]
2. Massimo Bertinaria, Barbara Rolando, Marta Giorgis, Gabriele Montanaro, Elisabetta Marini, Massimo Collino, Elisa Benetti, Pier Giuseppe Daniele, Roberta Fruttero, Alberto Gasco. 2012. Carnosine analogues containing NO-donor substructures: Synthesis, physico-chemical characterization and preliminary pharmacological profile. *European Journal of Medicinal Chemistry* **54**, 103-112. [[CrossRef](#)]
3. Neelam Shahani, Akira Sawa. 2012. Protein S-nitrosylation: Role for nitric oxide signaling in neuronal death. *Biochimica et Biophysica Acta (BBA) - General Subjects* **1820**:6, 736-742. [[CrossRef](#)]
4. Frederick A Villamena, Amlan Das, Kevin M Nash. 2012. Potential implication of the chemical properties and bioactivity of nitron spin traps for therapeutics. *Future Medicinal Chemistry* **4**:9, 1171-1207. [[CrossRef](#)]
5. Janet M. Dowding, Talib Dosani, Amit Kumar, Sudipta Seal, William T. Self. 2012. Cerium oxide nanoparticles scavenge nitric oxide radical (#NO). *Chemical Communications* . [[CrossRef](#)]
6. Donna L. Cioffi . 2011. Redox Regulation of Endothelial Canonical Transient Receptor Potential Channels. *Antioxidants & Redox Signaling* **15**:6, 1567-1582. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
7. Hélène Kremer, Céline Baron-Menguy, Angela Tesse, Yves Gallois, Alain Mercat, Daniel Henrion, Ramaroson Andriantsitohaina, Pierre Asfar, Ferhat Meziani. 2011. Human serum albumin improves endothelial dysfunction and survival during experimental endotoxemia: Concentration-dependent properties*. *Critical Care Medicine* **39**:6, 1414-1422. [[CrossRef](#)]
8. Taeko Horinouchi, Hidehiko Nakagawa, Takayoshi Suzuki, Kiyoshi Fukuhara, Naoki Miyata. 2011. Photoinduced Nitric Oxide Release from a Nitrobenzene Derivative in Mitochondria. *Chemistry - A European Journal* **17**:17, 4809-4813. [[CrossRef](#)]
9. C. B. Pattillo, S. Bir, V. Rajaram, C. G. Kevil. 2011. Inorganic nitrite and chronic tissue ischaemia: a novel therapeutic modality for peripheral vascular diseases. *Cardiovascular Research* **89**:3, 533-541. [[CrossRef](#)]
10. Anna Tillmann, Neil A.R. Gow, Alistair J.P. Brown. 2011. Nitric oxide and nitrosative stress tolerance in yeast. *Biochemical Society Transactions* **39**:1, 219-223. [[CrossRef](#)]
11. Samantha McLean, Lesley A.H. Bowman, Robert K. Poole. 2010. KatG from Salmonella Typhimurium is a peroxynitritase. *FEBS Letters* **584**:8, 1628-1632. [[CrossRef](#)]
12. Vittorio Calabrese , Carolin Cornelius , Enrico Rizzarelli , Joshua B. Owen , Alben T. Dinkova-Kostova , D. Allan Butterfield . 2009. Nitric Oxide in Cell Survival: A Janus Molecule. *Antioxidants & Redox Signaling* **11**:11, 2717-2739. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
13. Christy L. White, Paul J. Pistell, Megan N. Purpera, Sunita Gupta, Sun-Ok Fernandez-Kim, Taylor L. Hise, Jeffrey N. Keller, Donald K. Ingram, Christopher D. Morrison, Annadora J. Bruce-Keller. 2009. Effects of high fat diet on Morris maze performance, oxidative stress, and inflammation in rats: Contributions of maternal diet. *Neurobiology of Disease* **35**:1, 3-13. [[CrossRef](#)]